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### 56 Abstract

Atmospheric carbon dioxide enrichment (eCO<sub>2</sub>) can enhance plant carbon uptake and 57 growth<sup>1,2,3,4,5</sup>, thereby providing an important negative feedback to climate change by slowing 58 the rate of increase of the atmospheric CO<sub>2</sub> concentration<sup>6</sup>. While evidence gathered from 59 young aggrading forests has generally indicated a strong CO<sub>2</sub> fertilization effect on biomass 60 growth<sup>3,4,5</sup>, it is unclear whether mature forests respond to eCO<sub>2</sub> in a similar way. In mature 61 trees and forest stands<sup>7,8,9,10</sup>, photosynthetic uptake has been found to increase under eCO<sub>2</sub> 62 63 without any apparent accompanying growth response, leaving an open question about the fate of additional carbon fixed under  $eCO_2^{4,5,7,8,9,10,11}$ . Here, using data from the first ecosystem-64 65 scale Free-Air CO<sub>2</sub> Enrichment (FACE) experiment in a mature forest, we constructed a 66 comprehensive ecosystem carbon budget to track the fate of carbon as the forest responds to four years of  $eCO_2$  exposure. We show that, although the  $eCO_2$  treatment of ambient +150 67 ppm (+38%) induced a 12% (+247 g C m<sup>-2</sup> yr<sup>-1</sup>) increase in carbon uptake through gross 68 69 primary production, this additional carbon uptake did not lead to increased carbon 70 sequestration at the ecosystem level. Instead, the majority of the extra carbon was emitted 71 back into the atmosphere via several respiratory fluxes, with increased soil respiration alone 72 accounting for ~50% of the total uptake surplus. Our results call into question the 73 predominant thinking that the capacity of forests to act as carbon sinks will be generally 74 enhanced under eCO<sub>2</sub>, and challenge the efficacy of climate mitigation strategies that rely on 75 ubiquitous CO<sub>2</sub> fertilization as a driver of increased carbon sinks in global forests.

76

# 77 Main text

Globally, forests act as a large carbon sink, absorbing a significant portion of the anthropogenic  $CO_2$  emissions<sup>1,12</sup>, an ecosystem service that has tremendous social and

80 economic value. Whether mature forests will remain carbon sinks into the future is of critical importance for aspirations to limit climate warming to no more than 1.5 °C above pre-81 industrial levels<sup>13</sup>. Free-Air CO<sub>2</sub> Enrichment (FACE) experiments provide an opportunity to 82 83 determine the capacity of ecosystems to sequester carbon under the higher atmospheric CO<sub>2</sub> concentrations expected in the future<sup>3,4,5,7,8,10,11</sup>. Evidence gathered from the four first-84 85 generation forest FACE experiments, which all measured responses of rapidly-growing 86 young forest plantations, has generally indicated a strong CO<sub>2</sub> fertilization effect on biomass growth<sup>3,4</sup>. This CO<sub>2</sub> fertilization effect has been hypothesized to be one of the largest drivers 87 of the terrestrial carbon sink and its acceleration in recent decades<sup>14</sup>, potentially accounting 88 for up to 60% of present-day terrestrial carbon sequestration<sup>2</sup>. However, younger trees are 89 generally more responsive to rising  $CO_2$  than mature trees<sup>11</sup>, potentially because nutrient 90 limitation increases with stand age<sup>15</sup>. Thus, extrapolating evidence collected from these 91 92 experiments may be argued to provide an upper limit on how much carbon can be stored by global forests under  $eCO_2^{16}$ . Evidence from experiments with older trees on nutrient-poor 93 94 soils suggests that although eCO<sub>2</sub> increases leaf photosynthesis to a similar degree as in 95 young forests, stimulation of biomass growth and carbon storage may be lower or absent<sup>7,8,9,10</sup>. Reconciling these conflicting observations is a crucial step towards quantifying 96 97 the carbon sequestration capacity of mature forests in the future. It requires that we identify 98 the fate of the extra carbon fixed under eCO<sub>2</sub> in mature forests, which are expected to be 99 closer to a state of equilibrium between carbon uptake and turnover, compared to young 100 aggrading stands.

The *Eucalyptus* FACE (EucFACE) experiment is the world's first replicated, ecosystem-scale
mature forest FACE experiment (Extended Data Figure 1, 2). It is located in a warmtemperate evergreen forest that has remained undisturbed for the past 90 years, is dominated

105 by the regionally widespread tree *Eucalyptus tereticornis* and has an understorey composed 106 principally of native grasses and shrubs. The low-fertility soil has been shown to limit tree growth in an adjacent phosphorus-fertilization experiment<sup>17</sup>. Seven ecosystem-scale models 107 were used to predict the  $eCO_2$  response at EucFACE in advance of the experiment<sup>18</sup>, 108 109 highlighting three alternative hypotheses for the expected ecosystem response based on plausible assumptions incorporated in different models<sup>19</sup>. These hypotheses were: (i) 110 111 enhanced photosynthesis under eCO<sub>2</sub> would lead to increased biomass accumulation; (ii) 112 eCO<sub>2</sub>-induced increase in photosynthesis would be directly down-regulated by limited 113 nutrient availability; or (iii) eCO<sub>2</sub>-induced increase in photosynthesis would lead to increased autotrophic respiration<sup>18</sup>. This range of predictions among a suite of well-tested models 114 115 indicated a prognostic knowledge gap as to how the carbon cycling of mature forests would respond to the expected rise in atmospheric CO<sub>2</sub> concentration<sup>11</sup>, which is crucial to resolve in 116 the face of future carbon-climate uncertainty $^{20}$ . 117

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119 To date, both canopy trees and understorey plants at EucFACE have shown increased rates of 120 leaf photosynthesis but the canopy trees showed no significant increase in aboveground biomass growth under  $eCO_2^7$ , reflecting a similar lack of response observed in other  $eCO_2$ 121 experiments on mature trees<sup>8,9,10</sup>. Incorporating leaf-scale gas exchange measurements into a 122 123 process-based tree stand model, it was estimated that the observed +19% stimulation of light-124 saturated overstorey leaf photosynthesis<sup>7</sup> corresponded to a +11% stimulation of wholecanopy gross primary production (GPP) in response to  $eCO_2^{21}$ . However, the probable fate of 125 126 the extra carbon fixed under  $eCO_2$  remained undetermined. Where did the extra carbon go? 127

To answer this question, we compiled measurements on all major carbon pools and fluxes collected over four years of experimental treatment (2013-2016), including individual and

130 aggregated biomass and associated fluxes measured or inferred from plants, litter, soil, 131 microbes, and insects, and constructed an ecosystem carbon budget (Figure 1) under both 132 ambient (aCO<sub>2</sub>) and eCO<sub>2</sub> conditions (+150 ppm). We first confirmed mass balance of the 133 ecosystem carbon budget by checking agreement between independent estimates of GPP and 134 soil respiration (R<sub>soil</sub>) derived from separate data streams (Extended Data Figure 3; see Methods). For GPP of the aCO<sub>2</sub> plots, we confirmed that a process-based model estimate of 135 overstorey and understorey GPP (2059  $\pm$  211 g C m<sup>-2</sup> yr<sup>-1</sup>), driven by site-specific 136 meteorology and treatment-specific physiological data, broadly agreed with the sum of data-137 138 driven estimates of net primary production (NPP) and autotrophic respiration (2068  $\pm$  61 g C m<sup>-2</sup> yr<sup>-1</sup>). The carbon-use efficiency (NPP/GPP) of this mature forest was estimated to be 0.31 139 140  $\pm$  0.03, which is on the low end of global forest estimates, but consistent with studies that have observed this ratio to decline with stand  $age^{22}$  (Extended Data Figure 2). We further 141 142 confirmed carbon mass balance for R<sub>soil</sub> of the aCO<sub>2</sub> plots by comparing soil chamber-based estimates  $(1097 \pm 86 \text{ g C m}^{-2} \text{ yr}^{-1})$  with the sum of litterfall and independently estimated root 143 respiration ( $1086 \pm 14$  g C m<sup>-2</sup> yr<sup>-1</sup>), assuming no change in soil carbon pool (see Methods). 144 145 This agreement between independent estimates of components of the ecosystem carbon 146 budget gives confidence that our measurements captured the pools and fluxes of carbon with 147 low aggregate uncertainty and hence allow us to infer the fate of the extra carbon fixed under 148 eCO<sub>2</sub>.

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To accommodate the inherent pre-treatment plot differences (see Methods), we normalized the  $CO_2$  responses across plots by using a linear mixed-model with plot-specific pretreatment leaf area index as a covariate<sup>23,24</sup>. The non-normalized eCO<sub>2</sub> responses are provided in Extended Data Figure 4, and generally confirm the findings but with larger uncertainty. Our normalized responses (Figure 2, Extended Data Figure 5) showed that eCO<sub>2</sub> induced an

average of 12% increase (+247  $\pm$  195 g C m<sup>-2</sup> yr<sup>-1</sup>, mean  $\pm$  one standard deviation) in carbon 155 uptake, including contributions of overstorey (+192  $\pm$  193 g C m<sup>-2</sup> yr<sup>-1</sup>) and understorey GPP 156  $(+55 \pm 21 \text{ g C m}^{-2} \text{ yr}^{-1})$ . The fate of this additional carbon entering the system under eCO<sub>2</sub> 157 was primarily traced to an increase in  $R_{soil}$  (+128.8 ± 116.7 g C m<sup>-2</sup> yr<sup>-1</sup>, or 52% of the carbon 158 uptake surplus), followed by a smaller increase in tree stem respiration ( $R_{stem}$ ; +40.0 ± 43.6 g 159 C m<sup>-2</sup> yr<sup>-1</sup>, or 16% of the carbon uptake surplus). In comparison, the increase in total NPP 160  $(+67.3 \pm 12.7 \text{ g C m}^{-2} \text{ yr}^{-1})$ , or 28% of the carbon uptake surplus) corresponded to a smaller 161 increase in storage of the total carbon pools at the ecosystem-level ( $\Delta C_{\text{pools}}$ ; +31.6 ± 188.8 g C 162 m<sup>-2</sup> yr<sup>-1</sup>, or 12.8% of the carbon uptake surplus, Extended Data Figure 6). There was thus 163 164 little evidence of additional carbon accumulation under eCO<sub>2</sub> in this mature forest ecosystem. 165 We then compared three alternative methods (see Methods) of estimating net ecosystem 166 production (NEP; Figure 3). All three indicated that the ecosystem remained close to carbon-167 neutral under ambient CO<sub>2</sub> over the experimental period (mean  $\pm$  SD for the methods: 28  $\pm$ 225,  $21 \pm 129$ ,  $-73 \pm 50$  g C m<sup>-2</sup> yr<sup>-1</sup>, respectively), and that eCO<sub>2</sub> of +150 ppm did not result 168 in statistically significant increases in ecosystem carbon storage ( $109 \pm 258$ ,  $-19 \pm 171$ ,  $-42 \pm$ 169 262 g C m<sup>-2</sup> yr<sup>-1</sup>, respectively). However, the variability reported here means that we cannot 170 fully rule out the possibility of additional carbon storage under eCO<sub>2</sub>, but we stress that our 171 individual and aggregated responses consistently suggest a lack of CO2 response in this 172 173 mature forest (Figure 2 & 3, Extended Data Figure 5).

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The relatively small but positive NPP response to  $eCO_2$  was mainly driven by the understorey aboveground NPP response (NPP<sub>ua</sub>; +50.3 ± 17.9 g C m<sup>-2</sup> yr<sup>-1</sup>), which was 75% of the net NPP response (Figure 2). However, this significant NPP<sub>ua</sub> response did not result in an equivalent  $eCO_2$  effect on understorey aboveground biomass increment (+27.2 ± 29.7 g C m<sup>-2</sup> yr<sup>-1</sup>), suggesting a possible higher understorey biomass turnover under  $eCO_2$ . Smaller fluxes, 180 often neglected in other ecosystem carbon budgets, such as leaf consumption by insect herbivores (NPP<sub>ins</sub>;  $25.5 \pm 4.3$  vs.  $27.8 \pm 6.3$  g C m<sup>-2</sup> yr<sup>-1</sup>, aCO<sub>2</sub> vs. eCO<sub>2</sub> mean  $\pm$  SD), insect 181 frass production (Frass;  $10.5 \pm 1.8$  vs.  $11.4 \pm 2.6$  g C m<sup>-2</sup> yr<sup>-1</sup>), vegetation volatile carbon 182 emission (VC;  $2.63 \pm 0.18$  vs.  $2.45 \pm 0.13$  g C m<sup>-2</sup> yr<sup>-1</sup>), net ecosystem methane uptake (CH<sub>4</sub>; 183  $0.18 \pm 0.0009$  vs.  $0.19 \pm 0.0003$  g C m<sup>-2</sup> yr<sup>-1</sup>), and leaching of dissolved organic carbon (DOC; 184  $0.16 \pm 0.017$  vs.  $0.17 \pm 0.024$  g C m<sup>-2</sup> yr<sup>-1</sup>), contributed to the closure of the overall 185 186 ecosystem carbon budget (Figure 1; Extended Data Figure 3), but were not quantitatively 187 important in explaining pathways of the carbon uptake surplus under  $eCO_2$  (Figure 2, 188 Extended Data Figure 5, Extended Data Figure 6).

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190 Here we provide some of the first replicated experimental evidence on the probable fate of 191 carbon under eCO<sub>2</sub> in intact mature forest. We found that increased R<sub>soil</sub> accounted for ~50% 192 of the extra photosynthate produced by plants under eCO<sub>2</sub>. It has been suggested that the 193 increase in R<sub>soil</sub> at EucFACE was likely a consequence of increased root and rhizosphere respiration<sup>25,26</sup>, in contrast to other FACE sites where increased R<sub>soil</sub> was attributed to 194 enhanced soil organic matter decomposition (e.g. DukeFACE<sup>27</sup>). Here, the eCO<sub>2</sub>-induced 195 increase in  $R_{soil}$  was not accompanied by substantial changes in root respiration (18.6 ± 20.1 g 196 C m<sup>-2</sup> yr<sup>-1</sup>) or in carbon pools associated with fine roots (+7.0  $\pm$  12.5 g C m<sup>-2</sup> yr<sup>-1</sup>), microbes 197  $(+1.9 \pm 3.5 \text{ g C m}^{-2} \text{ yr}^{-1})$ , mycorrhizae  $(+0.4 \pm 0.5 \text{ g C m}^{-2} \text{ yr}^{-1})$ , leaf litter  $(+27.1 \pm 38.6 \text{ g C})$ 198  $m^{-2} yr^{-1}$ ) or soil (-23.8 ± 159.6 g C  $m^{-2} yr^{-1}$ ), suggesting that the additional carbon fixed under 199 200 eCO<sub>2</sub> may have led to an enhanced carbon transport belowground and a rapid belowground 201 turnover of this flux. Assimilation of these data into a carbon balance model supports this inference (Extended Data Figure 7, see Methods for details). An initial enhancement in 202 nitrogen and phosphorus mineralization was observed<sup>28</sup>, which suggested that the increased 203 R<sub>soil</sub> with eCO<sub>2</sub> could reflect soil organic matter priming with the potential to alleviate plant 204

nutrient stress in this low-phosphorus soil<sup>28,29</sup>. However, the enhanced soil mineralization rate and associated increase in nutrient availability did not persist over time<sup>28</sup>, indicating that this increased belowground carbon allocation and the rapid turnover of this flux was not effective in increasing phosphorus availability to the plants<sup>30</sup>.

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210 The ecosystem carbon budget presented here provides an opportunity to confront the three alternative hypotheses of the response of this system to eCO2 treatment that emerged from 211 model predictions made in advance of the experiment<sup>18</sup>. Our data do not support any of the 212 213 three hypotheses. The eCO<sub>2</sub>-induced increase in photosynthesis was not strongly downregulated by low nutrient availability<sup>7,21</sup>; nor did the eCO<sub>2</sub>-induced additional carbon uptake 214 215 lead to additional biomass accumulation, or enhanced aboveground respiration. These 216 predictions reflect common mechanisms by which terrestrial vegetation models implement nutrient limitation of the eCO<sub>2</sub> response<sup>18,19,31,32</sup>. In contrast, our results suggest a direct 217 218 connection between plant photosynthesis and belowground activity (Extended Data Figure 7), 219 in which increased belowground carbon allocation increased soil respiration at a rate that 220 accounted for half of the extra carbon fixed under  $eCO_2$  (Figure 2). Predictions made in 221 advance of the experiment did not capture this additional belowground carbon flux, despite 222 their general agreement with data on turnover rates of major carbon pools (Extended Data 223 Figure 8). This increased soil respiration has been demonstrated by some models to be an 224 important and often overlooked mechanism that reduces global soil carbon sequestration relative to estimates by many current models<sup>33</sup>. As a consequence of including this rapid 225 226 turnover of the increased belowground carbon allocation in terrestrial biosphere models, the 227 time-lag in emitting some of the extra carbon via biomass accumulation and litterfall input into the soils may be reduced, thereby leading to faster cycling of carbon<sup>34</sup> and therefore 228 229 possible different trajectories of carbon-climate predictions for the future.

231 A major form of land-based climate mitigation actions envisaged in the 2015 Paris 232 Agreement is to enhance forest biomass carbon stocks globally through the protection of 233 existing, largely mature, forests, and through afforestation of new areas. The mitigation 234 potential of forests lies in the accumulated stock of ecosystem carbon, not in the short-term 235 rate of forest photosynthesis. The probable fate of additional carbon determined in our study 236 (Figure 2) challenges the current thinking that all non-aggrading mature forests will contribute to enhanced carbon sinks due to CO<sub>2</sub> fertilization<sup>35</sup>, which further questions the 237 allowable CO<sub>2</sub> emission targets sourced from existing carbon cycle models<sup>13,36</sup>. Given that 238 the effect of CO<sub>2</sub> fertilization may be one of diminishing returns over time<sup>14</sup>, the statistically 239 240 non-significant eCO<sub>2</sub> effect on NEP (Figure 3), if representative of nutrient-limited mature forest ecosystems generally, suggests an even weaker carbon sink in the future, especially in 241 242 low-phosphorus systems such as EucFACE. Future research efforts should target a deeper 243 understanding of the nutrient-carbon feedbacks that likely constrain the carbon sink potential 244 of mature forests under  $eCO_2$ , and evaluate the implications of a potentially weaker terrestrial 245 land carbon sink in the development of robust mitigation strategies in the face of climate 246 change. More importantly, whilst the terrestrial carbon sink is integral to current strategies for 247 climate change mitigation, our results call for more active reductions of anthropogenic 248 emissions to meet the targets of the Paris Agreement.

### 249 Methods

### 250 EucFACE site description

The EucFACE facility (Extended Data Figure 1) is located in a mature evergreen Eucalyptus 251 252 forest on an alluvial spodosol in western Sydney, Australia (33°36'S, 150°44'E). The site has 253 been a remnant patch of native Cumberland Plain woodland since the 1880's and has 254 remained unmanaged for at least the past 90 years, with *Eucalyptus tereticornis* Sm. as the 255 dominant tree species (98% of the overstorey basal area). Eucalyptus trees occur naturally across Australia, accounting for 78% of native forest area in Australia<sup>37</sup> and are planted 256 widely around the globe<sup>38</sup>. Infrastructure for six large circular plots (490 m<sup>2</sup> each) was 257 established in 2010. Starting on 18<sup>th</sup> September 2012, three plots were subjected to free-air 258 259  $CO_2$  enrichment treatment using a computer-controlled pre-dilution method. The  $CO_2$ 260 concentrations at EucFACE were ramped up over a six-month period, increasing by +30 ppm 261 every five weeks in discrete steps (+30, 60, 90, 120, and 150 ppm). The full elevated  $CO_2$ treatment of +150 ppm started on 6<sup>th</sup> February 2013 during daylight hours over all days of the 262 263 year. The site is characterized by a humid temperate-subtropical transitional climate with a mean annual temperature of 17.5°C and a mean annual precipitation of 800 mm (Figure S1). 264 The soil is a Holocene alluvial soil of low fertility with low phosphorus content<sup>7,17</sup>. Soil 265 266 texture is a loamy sand (> 75% sand content) up to 50 cm in depth. From ca. 50 to 300 cm 267 depth, soils are sandy clay loam, with > 30% silt and clay. Average bulk density is 1.39, 1.69 and 1.71 g cm<sup>-3</sup> for depths of 0-10, 10-20 and 20-30 cm, respectively (Figure S2). Permanent 268 groundwater depth is  $\sim 11$  m below the soil surface<sup>39</sup>. Understorey vegetation is a diverse 269 mixture of 86 species including forbs, graminoids and shrubs<sup>40</sup>. The dominant understorey 270 271 species is *Microlaena stipoides*, a  $C_3$  perennial grass that accounted for ~70% of herbaceous biomass and responded rapidly to rainfall variability<sup>41</sup>. 272

# 273

## 274 Estimates of carbon pools and fluxes

275 We estimated plot-specific carbon pools and fluxes at EucFACE over 2013-2016 (Extended 276 Data Table 1). We defined pools as a carbon reservoir and annual increments as the annual 277 changes in the size of each reservoir. We compartmentalized the ecosystem into 11 carbon 278 pools, namely overstorey leaf (Col), stem (Cstem), fine root (Cfroot), coarse root (Ccroot), intermediate root (Ciroot), understorey aboveground (Cua), soil (Csoil), microbe (Cmicr), 279 mycorrhizae (Cmyco), leaf litter (Clit), and aboveground insect (Cins) carbon pools, and reported 280 pool size in the unit of g C  $m^{-2}$ . We defined fluxes as components of the carbon flow through 281 the system, and report them in the unit of g C m<sup>-2</sup> yr<sup>-1</sup>. All annual incremental changes in 282 carbon pools were reported in g C m<sup>-2</sup> yr<sup>-1</sup> with a symbol  $\Delta$ . We converted estimates of 283 biomass into carbon content using variable-specific carbon fractions (f) defined in Extended 284 285 Data Table 2. Below we describe how each pool and flux was estimated.

286

287 <u>Pools</u>

Soil carbon pool ( $C_{soil}$ ; Figure S2) was estimated based on quarterly sampled soil carbon content (oven-dried at 40 °C for 48 hours) and plot-specific soil bulk density at three depths (0 - 10 cm, 10 - 20 cm, 20 - 30 cm). Out of the 15 dates when samples were taken, soil carbon content below the top 10 cm of soil was measured on three dates. To obtain a more accurate estimate of annual incremental change in soil carbon pool, we therefore reported soil carbon pool for the top 10 cm only. There were no temporal and eCO<sub>2</sub> trends in soil carbon content at deeper depths.

296 **Overstorey leaf carbon pool (C\_{ol};** Figure S3) was estimated based on continuous measures of leaf area index (LAI) and specific leaf area (SLA, m<sup>2</sup> leaf area g<sup>-1</sup> leaf DM), following C<sub>ol</sub> 297 = LAI × SLA ×  $f_{ol}$ , where  $f_{ol}$  is a carbon fraction constant for overstorey leaves (Extended 298 299 Data Table 2). Daily averages of plot-specific LAI were estimated based on the attenuation of diffuse radiation in a homogenous canopy<sup>24</sup>. The number of observations varies between days, 300 301 depending on the number of 30-minute cloudy periods. SLA was estimated based on time-302 series measures of leaf mass per area (LMA), and was then linearly interpolated to plot-303 specific daily values over time.

304

Stem carbon pool (Cstem; Figure S4) was estimated based on tree-specific height and 305 306 diameter at breast height (DBH) measurements, and an allometric scaling relationship derived for *E. tereticornis*<sup>7,42</sup>. DBH changes were measured repeatedly at roughly monthly intervals, 307 308 at 1.3 m height. Bark was periodically removed from under the dendrometer bands - this 309 effect on DBH was considered by calculating biomass once per year using December data 310 only. Stem biomass data were summed for each plot and averaged over the plot area to obtain 311 ground-based estimates, and was then converted into C<sub>stem</sub> using treatment-specific carbon 312 fraction (Extended Data Table 2).

313

Understorey aboveground carbon pool ( $C_{ua}$ ; Figure S5) was estimated at 1-3 month intervals between February 2015 and December 2016 using non-destructive measurements of plant height obtained from stereo-photography<sup>43</sup>. In each of the four 2m × 2m understorey monitoring subplots within each plot, stereo photographs were collected using a Bumblebee XB3 stereo camera (Point Grey Research) mounted ~2.4 m above the ground surface and facing vertically downwards towards the center of the subplot. Stereo images were taken at dusk under diffuse light conditions to avoid measurement errors related to shadows from 321 trees and EucFACE infrastructure. On each sampling date, three sets of stereo photographs 322 were taken in each subplot to produce a large number (i.e. 100,000 s) of understorey plant 323 height estimates from which mean plant height (H<sub>mean</sub>, in m) was calculated for each plot. Understorey aboveground biomass (Bua, in kg m<sup>-2</sup>) for each plot was predicted from Hmean 324 using an empirical model developed for the grassy understorey vegetation at EucFACE ( $B_{ua}$  = 325  $1.72 \times H_{mean} - 0.05)^{43}$ . The four subplot-level estimates were averaged to obtain a plot-level 326 estimate of B<sub>ua</sub>, and then converted to an estimate of C<sub>ua</sub> using a carbon fraction constant 327 328 (Extended Data Table 2).

329

**Root carbon pool (** $C_{root}$ **)** consists of fine root ( $C_{froot}$ **)**, intermediate root ( $C_{iroot}$ **)**, and coarse root ( $C_{croot}$ ) pools, with  $C_{froot}$  defined as roots with diameter of < 2 mm,  $C_{iroot}$  defined as roots with diameter of 2 – 3 mm, and the remaining roots defined as  $C_{croot}$  (Figure S6). The  $C_{root}$ pool includes roots of both overstorey and understorey vegetation. Total root biomass ( $B_{root}$ ) was estimated based on an allometric relationship with stand basal area (derived from DBH) derived for Australian forest species<sup>44</sup>, as follows: ln( $B_{root}$ ) = 0.787 × ln (DBH) + 1.218.

336

337 Standing intermediate root (2-3 mm in diameter) and fine root biomass ( $\leq 2$  mm in diameter) 338 were sampled in four subplots per plot at two depths (0 - 10 cm and 10 - 30 cm) in year 2017, 339 whereas only fine root biomass at the same depths with the same number of subplots was repeatedly sampled over the period of 2014-2016<sup>29</sup>. We estimated a depth-specific 340 341 relationship between fine root biomass (< 2 mm in diameter) and total root biomass less than 342 3 mm in diameter based on samples collected in 2017, and calculated the intermediate root 343 biomass for the period of 2014-2016 based on its corresponding fine root biomass. Coarse 344 root biomass was then estimated as the net difference between total allometrically-derived 345 root biomass and that of roots with diameter < 3mm. The fine, intermediate, and coarse root

biomass were multiplied by the corresponding carbon fraction constants to obtain C<sub>froot</sub>, C<sub>iroot</sub>,
 and C<sub>croot</sub>, respectively (Extended Data Table 2).

348

Microbial carbon pool ( $C_{micr}$ ) was estimated based on fumigation extraction and 0.5 M K<sub>2</sub>SO<sub>4</sub> extraction as in Ref. 25 using samples taken at 0-10 cm soil depth over the period of 2012 - 2015. Total organic carbon was determined on a Shimadzu TOC analyzer (TOC-L TNM-L; Shimadzu, Sydney, Australia), which was then multiplied by soil bulk density over the same soil depth to obtain the  $C_{micr}$  (Figure S7a).

354

355 Mycorrhizal carbon pool ( $C_{myco}$ ) for the top 10 cm of soil was estimated via measurements 356 of colonization of mycorrhizal in-growth bags, carbon isotopic partitioning, microbial 357 phospholipid fatty acid abundance and  $C_{micr}$ . Nine 45 µm nylon mesh bags (4 × 5 cm) filled with sand, which excluded roots but allowed access of fungi<sup>45</sup>, were buried in November 358 359 2014 in each experimental plot and three bags were subsequently collected every four months 360 for one year. Phospholipid-derived fatty acids (PLFA), a proxy for total microbial biomass 361 abundance, were quantified in sand bags and native field soil following the protocol by Ref 46.  $\delta^{13}$ C values of ground subsamples of this sand, native soil carbon, and aboveground plant 362 363 tissue (leaves of Eucalypts in April 2014) were used to estimate the fraction of the 364 accumulated carbon in sand bags that was derived from plant carbon using isotopic mass 365 balance. Due to the exclusion of roots, plant-derived carbon in bags can be attributed to 366 mycorrhiza. This plant-derived unitless fraction was then multiplied by the total 367 concentration of PLFA in sand bags to obtain the amount of the total PLFA contributed by 368 mycorrhiza ( $\mu g$  PLFA / g sand). To scale this to native soil PLFA concentrations we then 369 calculated the ratio between mycorrhizal PLFA in sand bags to total PLFA in soil 370 (representing the total microbial pool). Subsequently, to estimate  $C_{myco}$ , this ratio was 371 multiplied by the  $C_{micr}$  in each plot (Figure S7b).

372

373 Leaflitter carbon pool (Clit) was estimated based on leaf litter decomposition rate and leaf litterfall data collected by litter baskets (Figure S8)<sup>24</sup>. Leaf litter decomposition rates were 374 375 estimated over 24 months using litter bags. Briefly, 2 g air-dried Eucalyptus litter was added 376 to  $10 \times 15$  cm litter bags with a 2-mm mesh size. Twelve litter bags were randomly allocated to 4 subplots within each treatment plot, and two litter bags were collected at 3, 6, 9, 12, 18 377 378 and 24 months to calculate mass loss over time (mass loss was averaged across the two 379 replicates from each subplot). A leaflitter exponential decay function was estimated for each 380 plot, based on data collected over this 24-month period. Leaf litterfall was estimated from monthly collections of material from circular fine-mesh traps (each  $0.2 \text{ m}^2$ ) at eight random 381 382 locations for each plot. We then applied the exponential decay function with litterfall biomass 383 to obtain C<sub>lit</sub>, assuming a carbon fraction constant (Extended Data Table 2).

384

Insect carbon pool (Cins) was estimated based on two different sampling techniques, with 385 386 aerial insects partially estimated based on monthly dead insect data collected from circular fine-mesh traps of 0.2  $\text{m}^2$  at eight random locations for each plot<sup>47</sup>, and understory insects 387 estimated based on vacuum suction sampling from two locations for each plot<sup>48</sup>. The insect 388 389 biomass estimated based on these two sampling techniques may be a conservative estimate (the frass produced would suggest presence of a larger insect biomass<sup>49</sup>); nevertheless, they 390 391 provided a direct estimate based on data collected in situ. The vacuum suction method 392 collected invertebrates from understorey vegetation in two  $1 \times 1$  m subplots using a petrolpowered 'G-Vac' vacuum device run on full-throttle for 20 s, for a total of five sampling 393 394 campaigns. Trapping locations were randomly chosen and fixed between sampling 395 campaigns. All invertebrates were sorted from debris, dried to constant weight at 60 °C and 396 weighed on a microbalance with a precision of 1  $\mu$ g. We assumed that vacuum samples as 397 well as fine-mesh trap samples represent point estimates of invertebrate abundance. Then, the 398 total biomass of sampled invertebrates was summed across sampling methods within each 399 plot. A constant carbon fraction based on Ref 50 (Extended Data Table 2) was used to 400 convert biomass into C<sub>ins</sub> pool (Figure S9).

# 401 *Ecosystem carbon uptake fluxes*

402 Overstorey gross primary production (GPP<sub>0</sub>) for each plot was provided by a stand-level 403 model simulation (MAESPA), forced by hourly meteorological data, daily plot-specific leaf 404 area index and leaf-scale treatment-specific photosynthetic parameters measured at the site (Figure S10a)<sup>7,21</sup>. In short, MAESPA was used as a tool to up-scale leaf-level gas exchange 405 406 measurements to the whole canopy. In MAESPA, each plot consists of individual tree crowns 407 that are located and parameterized with measured coordinates, crown size, and LAI. Each 408 crown is divided into six layers, with leaf area uniformly distributed in each layer. Within 409 each layer, the model simulates twelve grid points. The incident radiation on the sunlit and 410 shaded leaf area at each grid point is calculated considering shading from upper crown and 411 surrounding trees, solar angle (zenith and azimuth), and light source (diffuse or direct). Incident radiation is then used to calculate gas exchange using a Farquhar<sup>51</sup> formulation for 412 photosynthesis and a Medlyn formulation<sup>52</sup> for stomatal conductance. The model was 413 parameterized with treatment-specific leaf gas exchange measurements made in situ<sup>7,53</sup>. Leaf 414 415 respiration and its temperature dependence were also quantified using data collected on site, 416 then up-scaled to the canopy using MAESPA. The performance of the model was evaluated by comparing the simulated transpiration flux to sap flow data<sup>54</sup>. 417

419 Similarly, understorey GPP (GPP<sub>u</sub>) (Figure S10b) was simulated using MAESPA with photosynthetic parameters taken for the dominant grass Microlaena stipoides<sup>41</sup>. The 420 421 parameterization of understory vegetation is different from that of the canopy. In each plot, 422 the understory was assumed to form a single crown covering the whole plot (i.e., a circle with 423 12.5 m radius) at a height of 1.5 m. The LAI of the understory was estimated using 424 phenology camera digital photographs taken at four permanent understorey vegetation monitoring subplots in each plot<sup>43</sup>. The average green pixel content was calculated from three 425 426 photos in each subplot, and assumed to be the same as the fraction of absorbed PAR. We then 427 assumed a light extinction coefficient of 0.5 in Beers' Law and calculated understorey LAI. 428 Before 2014 there were 3 campaigns per year while from 2014 the cameras were automated, 429 and we used the fortnightly averages. Leaf gas exchange parameters were obtained from Ref 430 41 and covered four to six campaigns per year from 2013 to 2016. We estimated a one-time  $g_1$  parameter<sup>52</sup> for all plots and time, and assumed constant carboxylation rate ( $V_{cmax}$ ) and 431 432 electron transport rate  $(J_{max})$  values at 25 °C across plots. Basal leaf respiration rate and the 433 temperature dependence of photosynthesis and respiration were assumed to be the same as 434 those for the canopy. The understory simulation was conducted separately from the canopy, 435 with canopy LAI from Ref 24 included to account for the shading from the canopy, branches 436 and stems on the understory.

437

For the **methane net flux (CH<sub>4</sub>),** air samples were collected following the closed-chamber method (or Non-Flow-Through Non-Steady-State [NFT-NSS] method). Seven replicated chambers were available for each plot. Headspace samples were collected monthly, over a period of one hour and analyzed by gas chromatography. Fluxes were estimated by a mixture of linear and quadratic regressions (depending on goodness-of-fit), assuming a constant air pressure of one atmosphere and correcting the air temperature inside the chambers for each 444 air sample<sup>55</sup>. The CH<sub>4</sub> fluxes are net fluxes, which represent the sum of: 1) CH<sub>4</sub> efflux 445 (emissions from the soil into the atmosphere); 2) CH<sub>4</sub> influx (uptake from the atmosphere 446 into soil). Here, the annual net CH<sub>4</sub> flux was an ecosystem influx and was presented as 447 positive values (Figure S11a).

448

### 449 <u>Production fluxes</u>

450 Plant net primary production (NPP) is the sum of overstorey leaf (NPP<sub>ol</sub>), stem (NPP<sub>stem</sub>), 451 fine root (NPP<sub>froot</sub>), intermediate root (NPP<sub>iroot</sub>), coarse root (NPP<sub>croot</sub>), other (including twigs, 452 barks, and seeds; NPP<sub>other</sub>), understorey aboveground (NPP<sub>ua</sub>), and consumption of overstorey leaf by insect herbivores (NPP<sub>ins</sub>). NPP<sub>ol</sub> and NPP<sub>other</sub> were estimated based on monthly litter 453 data collected from circular fine-mesh traps of 0.2 m<sup>2</sup> at eight random locations for each plot 454 455 (Figure S12). Litter was sorted into leaf, twigs, bark, and seeds, dried to constant mass at 456 40 °C and weighed. A subsample was reweighed when dried to constant mass at 70 °C and a small moisture correction<sup>7</sup> was applied to the leaf component of the whole dataset. NPP<sub>ol</sub> was 457 458 computed as the sum of annual leaf litter, which excluded leaf consumption by insects. For 459 twigs, we assumed strictly annual turnover across the years. NPPstem (Figure S13) and 460 NPP<sub>croot</sub> (Figure S14) were estimated based on annual incremental change of stem biomass 461 and coarse root biomass, respectively. NPPfroot was estimated based on samples collected from in-growth cores at four different locations per plot (Figure S14). NPP<sub>iroot</sub> was estimated 462 based on a global mean coarse root turnover rate (0.3605 yr<sup>-1</sup>) for evergreen broadleaf 463 forests<sup>56</sup>, and the C<sub>iroot</sub> pool in our dataset (Figure S14). 464

465

466 NPP<sub>ua</sub> was estimated based on biomass clippings taken between 2015 - 2017, assuming one
467 understorey turnover per harvest interval (Figure S15). We used a clip-strip method of
468 biomass harvest as has been applied previously at the BioCON experiment<sup>57</sup>. Specifically,

four narrow strips, each with a size of  $1 \text{ m} \times 0.1 \text{ m}$ , were situated in each of the experimental plots at least 2 m away from the vertical pipes for FACE, while avoiding the understory shrubs. The understory herbaceous species were clipped approximately 1 cm above soil level. The total mass per harvest represents the total production. Biomass samples were oven dried for two days at 60 °C, and converted into carbon mass by applying a constant fraction (Extended Data Table 2).

475

476 NPP lost to overstorey leaf consumption by insect herbivores (NPP<sub>ins</sub>) was estimated based 477 on insect frass data (Frass) collected from the circular fine-mesh traps, and a relationship 478 between frass mass and insect-consumed leaf mass derived based on multiple *Eucalyptus* tree 479 species at different CO<sub>2</sub> concentrations (Figure S16a)<sup>58,59</sup>. Frass was estimated based on 480 annual collection of frass biomass collected from the circular fine-mesh litter traps with their 481 associated carbon content (Extended Data Table 2; Figure S16c).

482

# 483 <u>Outfluxes</u>

Leaching lost as **dissolved organic carbon (DOC)** from soils was estimated based on concentrations of DOC in soil solutions, provided by water suction lysimeter measurements<sup>28</sup>. Lysimeters were installed to two depths (0 - 15 cm and 35 - 75 cm, which is immediately above the impermeable layer). Here we assumed that DOC reaching deeper depth is lost from the system at a rate of 20 ml m<sup>-2</sup> d<sup>-1</sup>, which is an estimate of the daily drainage rate at the site (Figure S11b).

490

491 Plant autotrophic respiration (R<sub>a</sub>) consists of overstorey leaf (R<sub>ol</sub>), stem (R<sub>stem</sub>), root (R<sub>root</sub>),
492 understorey aboveground (R<sub>ua</sub>) (Figure S17), and growth respiration (R<sub>grow</sub>) (Figure S18). R<sub>ol</sub>

493 and  $R_{ua}$  were based on MAESPA simulation (Figure S17a, c), as described in the respective 494 GPP sections.  $R_{grow}$  was estimated by taking a constant fraction of 30% of total NPP as 495 measured directly on *E. tereticornis* trees<sup>60</sup>.

496

R<sub>stem</sub> was estimated from measurements of stem CO<sub>2</sub> efflux performed in three dominant 497 498 trees per plot (Figure S17b). Collars were horizontally attached to the stem at an approximate height of 0.75 m, and  $R_{stem}$  (nmol CO<sub>2</sub> m<sup>-2</sup> s<sup>-1</sup>) was measured with a portable infrared gas 499 analyzer coupled to a soil respiration chamber adapted for this purpose<sup>61</sup>. Measurement 500 501 campaigns were performed every one or two months from December 2017 to October 2018, and the relationship between R<sub>stem</sub> and air temperature (T<sub>air</sub>) was used to extrapolate R<sub>stem</sub> 502 across the surveyed period, following  $R_{stem} = 0.1866 \times 2.84^{Tair/10}$  ( $r^2 = 0.42$ , p < 0.0001).  $R_{stem}$ 503 504 was then upscaled to the stand level considering the ratio of stem axial surface per unit of soil 505 surface measured per plot. Stem surface area was inferred from the measured tree diameter 506 based on dendrometer, and a relationship between diameter and stem surface area estimated 507 from the Terrestrial Laser Scanning (TLS) data. Stem surface area and diameter in the TLS 508 data was estimated through quantitative structure models presented in Ref. 62 and 63. TLS 509 data were acquired with a RIEGL VC-400 terrestrial laser scanner (RIEGL Laser 510 Measurement Systems GmbH). Stem surface area was derived from the TLS data following a 511 two-step approach: (i) manually extracting single trees from the registered TLS point cloud; 512 and (ii) deriving parameters for an extracted single tree. Once a tree is extracted from the 513 point cloud, the next step was to strip off the leaves, and segment the point cloud into stem 514 and branches. Finally, the surface of the segments was reconstructed with geometric 515 primitives (cylinders). The method used a cover set approach, where the point cloud was 516 partitioned into small subsets, which correspond to small connected patches in the tree 517 surface.

519 R<sub>root</sub> was partitioned into fine root (R<sub>froot</sub>), intermediate root (R<sub>iroot</sub>), and coarse root (R<sub>croot</sub>) 520 respiration (Figure S17d). Mass-based rates of fine root and intermediate root respiration (nmol CO<sub>2</sub> DM g<sup>-1</sup> s<sup>-1</sup>) were measured for detached roots sampled by soil cores at 10 cm soil 521 522 depth at four subplots per plot with a portable infrared gas analyzer coupled to a small root 523 chamber. Measurement campaigns were performed every one or two months from November 524 2018 to July 2019. The relationship between root respiration and soil temperature (T<sub>soil</sub>) at 10 cm soil depth was used to extrapolate the corresponding root respiration rates across the 525 surveyed period, following the equations:  $R_{froot} = 1.138 \times 1.614^{0.0479 \times Tsoil}$  ( $r^2 = 0.36$ , p < 526 0.0001, RMSE = 1.054), and  $R_{iroot} = 0.9764 \times 1.586^{0.0641 \times Tsoil}$  ( $r^2 = 0.52$ , p < 0.0001, RMSE = 527 0.597). The mass-based rate of coarse root respiration was assumed to be the same as the 528 529 mass-based rate of stem respiration. R<sub>froot</sub>, R<sub>iroot</sub> and R<sub>croot</sub> were then upscaled to the stand level to obtain R<sub>root</sub> with fine root, intermediate root, and coarse root biomass, respectively. 530

531

532 Carbon efflux due to insect respiration (R<sub>ins</sub>) was estimated as the net difference between
533 NPP<sub>ins</sub> and Frass, assuming no net change in insect biomass (Figure S16b).

534

535 Soil respiration ( $\mathbf{R}_{soil}$ ): The rate of soil CO<sub>2</sub> efflux was measured at eight locations within 536 each plot, where a permanent PVC collar inserted into the soil was co-located with soil TDR 537 probes for continuous measurements of soil temperature (5-cm-depth) and volumetric water 538 content (0 to 21-cm-depth; CS650-L; Campbell Scientific, Logan, UT, USA). R<sub>soil</sub> was 539 measured manually at all collar locations every 2-3 weeks, in addition to 30-minute 540 measurements using automated chambers (Li-8100-103; Licor) at one location within each plot, resulting in >300,000 observations over the study period<sup>26</sup>. These data were used to 541 parameterize a semi-mechanistic model of R<sub>soil</sub>, in which R<sub>soil</sub> was predicted based on 542

543 measurements of soil properties, soil physics, and measured soil temperature and volumetric water content<sup>64</sup>. This model successfully recreated the observed fluxes ( $r^2$  between predicted 544 and observed survey  $R_{soil}$  was 0.65)<sup>26</sup>. Annual sums of  $R_{soil}$  were derived by summing the 545 546 averaged daily fluxes over eight locations within each plot, where daily fluxes at each 547 location were predicted based on the semi-mechanistic model and daily soil temperature and 548 volumetric water content data taken adjacent to each measurement collar. Soil heterotrophic respiration (R<sub>hetero</sub>) was taken as the net difference between R<sub>soil</sub> and R<sub>root</sub> (Figure S19). Total 549 ecosystem respiration (R) was calculated as the sum of R<sub>a</sub>, R<sub>hetero</sub>, R<sub>ins</sub>, and VC. 550

551

**Volatile carbon (VC;** Figure S20) flux as isoprene ( $C_5H_8$ ) and monoterpenes was estimated using the Model of Emissions of Gases and Aerosols from Nature (MEGAN)<sup>65</sup>. Isoprene represents over half of all volatile organic carbon species emitted by vegetation globally, and is the dominant source of VC emission at our site. A MEGAN box-model was built from the version used in Ref. 66, centered on the EucFACE facility to calculate hourly emissions of isoprene across the period 2013-2016 for all six plots:

558

559 Where EF is the compound-specific basal emission factor,  $\gamma$  is the emission activity factor, 560 accounting for changes in the emission response due to light, temperature, leaf age and soil 561 moisture. The MEGAN simulations were driven by daily input data of LAI, soil moisture, 562 and hourly input data of photosynthetic active radiation, temperature, atmospheric pressure, 563 wind speed and relative humidity.

 $VC = EF \times LAI \times \gamma$ 

564

565 The isoprene EFs for ambient and elevated  $CO_2$  plots were derived from in-line 566 photosynthetic gas-exchange measurements coupled with simultaneous volatile isoprenoid 567 sampling. The isoprene was collected onto sterile stainless steel thermal desorption tubes at

568 the same time as gas exchange was measured, and these were capped and later thermally 569 desorbed for off-line volatile analysis in the laboratory using a Shimadzu 2010 Plus GC-MS 570 system connected to a Shimadzu TD20 automated cartridge desorber. The sampling and GC-571 MS analysis methodology is described in detail in Ref 67. The chromatographic peaks were 572 identified by comparing them to an isoprene standard and reference mass spectra in the NIST 573 Mass Spectral Library (https://www.nist.gov/srd). Monoterpene emissions were sampled during February 2018 using a push-pull headspace technique<sup>68</sup> from enclosed branches 574 575 containing approximately 10 leaves and trapped on adsorbent cartridges (150 mg Tenax TA 576 and 200 mg Carbograph 1TD, Markets International Limited, United Kingdom) at an outflow rate of 200 ml min<sup>-1</sup> for 15 min. Before each measurement, the sampling system was 577 578 equilibrated for 15 min at an inflow rate of 1000 mol min<sup>-1</sup>. Monoterpenes were analyzed by 579 gas chromatography-mass spectrometry (R7890A Series GC coupled with a 5975C inert 580 MSD/DS Performance Turbo EI System, Agilent Technologies, Inc., Santa Clara, CA, USA), 581 as described by Ref 69. The obtained chromatograms were deconvoluted, analyzed and data retrieved using the software PARADISe<sup>70</sup> version 3.88. Identification of compounds was 582 583 performed using analytical standards and according to their mass spectra in the NIST11 584 library. Pure analytical standards were used for quantification. The box-model produced 585 isoprene and monoterpenes were converted to carbon content using the respective molecular 586 mass ratios.

587

# 588 <u>Net Ecosystem Production</u>

Net ecosystem production (NEP) was estimated based on three different methods that estimated NEP in relatively independent ways (Figure 3), similar to Ref 71. The first method considered NEP as the difference between total ecosystem influx and total ecosystem outflux (i.e. In - Out), which relied on both process-based modeling and empirical upscaling of

respiratory fluxes collected from the field. The second method considered NEP as NPP minus R<sub>hetero</sub> (i.e. NPP - R<sub>hetero</sub>), with NPP relying mostly on litter-based production estimates, and R<sub>hetero</sub> relying on R<sub>soil</sub> and R<sub>root</sub> estimates. The third method considers NEP as the sum of changes in carbon pools over time in the ecosystem (i.e.  $\Delta C_{pools}$ ), which was mostly determined by biomass estimates. Equations for each method are provided below:

Method	NEP =
In - Out	$GPP_{o} + GPP_{u} + CH_{4} - R_{ol} - R_{stem} - R_{soil} - R_{ua} - R_{ins} - DOC - VC - R_{grow}$
NPP - R <sub>hetero</sub>	$NPP_{ol} + NPP_{stem} + NPP_{froot} + NPP_{iroot} + NPP_{croot} + NPP_{other} + NPP_{ua} + NPP_{ins}$ -
	R <sub>hetero</sub>
$\Delta C_{\text{pools}}$	$\Delta C_{soil} + \Delta C_{ol} + \Delta C_{stem} + \Delta C_{croot} + \Delta C_{froot} + \Delta C_{iroot} + \Delta C_{ua} + \Delta C_{lit} + \Delta C_{ins} + \Delta C_{micr} + \Delta C_{myco}$

598

#### 599 Carbon budget evaluation

600 We evaluated the mass balance of our estimated ecosystem carbon budget in two ways. 601 Firstly, we compared model simulated GPP with the aggregated sum of NPP and  $R_a$ 602 (Extended Data Figure 3a, b). GPP was simulated by a stand-level ecophysiological model, driven by hourly meteorological data and parameterized with site-specific ecological data<sup>20</sup>. 603 This GPP should equal to the aggregation of NPP ( $NPP_{ol} + NPP_{stem} + NPP_{froot} + NPP_{iroot} + NPP_{iroot}$ 604 605  $NPP_{croot} + NPP_{other} + NPP_{ua} + NPP_{ins}$  and  $R_a$  fluxes ( $R_{ol} + R_{stem} + R_{root} + R_{ua} + R_{grow}$ ), which 606 were mostly extrapolated based on field data. Secondly, R<sub>soil</sub> estimated based on soil collar flux measurements  $^{24}$  was evaluated against the sum of litterfall and  $R_{root}$  (Extended Data 607 608 Figure 3c, d), assuming minimal changes in soil carbon stock (as change over this short 609 period of time is beyond the detection limit in a complex and slow-growing mature forest 610 ecosystem like EucFACE). Here, litterfall was the sum of NPPol + NPPfroot + NPPiroot +

611 NPP<sub>other</sub> + NPP<sub>ua</sub> + Frass, and R<sub>root</sub> was extrapolated based on root biomass and temperature
612 functions.

613

# 614 Statistical analyses

We performed linear mixed-model analysis using the "lmer" function within the "lme4" 615 package<sup>72</sup> in software R<sup>73</sup> to determine the CO<sub>2</sub> treatment effect on all reported variables. All 616 fluxes were reported at an annual rate (g C m<sup>-2</sup> yr<sup>-1</sup>). In our model, date and CO<sub>2</sub> treatment 617 were considered as fixed factors, plot as a random factor, and plot-specific pre-treatment LAI 618 619 (i.e. 4-month average LAI before full  $CO_2$  treatment was switched on) as a covariate to 620 account for pre-treatment differences among treatment plots. Normalizing all response 621 variables with a covariate that integrates light, water and nutrient constraints helps to isolate the CO<sub>2</sub> effect<sup>23</sup>, as has been done previously at the site<sup>24</sup> and elsewhere<sup>8,23</sup>. Confidence 622 623 intervals for the CO<sub>2</sub> effect size of individual variables were reported using the function 624 "confint", which applies quantile functions for the t-distribution after model fitting. 625 Confidence intervals for the predicted flux and pool were reported as the standard deviation of the plot-specific totals (n = 3). Similarly, confidence intervals for the aggregated fluxes 626 627 (e.g. NPP) were reported by summing individual component fluxes that constitutes the 628 aggregated flux for each plot and computing the standard deviations across plots (n = 3). Finally, confidence intervals for the  $CO_2$  effect size (SD<sub>agg</sub>) of some aggregated fluxes (e.g. 629 630 NPP) were calculated by pooling the standard deviations of the aggregated fluxes for ambient 631 (SD<sub>amb</sub>) and elevated CO<sub>2</sub> treatment (SD<sub>ele</sub>), following:

$$SD_{agg} = \sqrt{\frac{SD_{amb}^2 + SD_{ele}^2}{2}}$$

## 633 Uncertainty analysis

634 We applied a Markov Chain Monte Carlo (MCMC) data assimilation algorithm to a 635 simplified carbon cycle framework to make inference of the uncertainties around the fate of 636 carbon in our carbon budget. We simplified our carbon budget into eight pools (Extended 637 Data Figure 7), namely, leaf (C'<sub>leaf</sub>, which includes overstorey and understorey), wood 638 (C'wood, which includes stem and coarse root), root (C'root, which includes fine root and intermediate root), aboveground litter (C'aglit), belowground litter (C'bglit), mycorrhizae 639 640 (C'<sub>myco</sub>), microbe (C'<sub>micr</sub>), and soil (C'<sub>soil</sub>). Here, C'<sub>aglit</sub> and C'<sub>bglit</sub> were assumed unknowns 641 and inferred from the analysis. Net primary production (NPP) was calculated as the 642 difference of gross primary production (GPP) and autotrophic respiration ( $R_a$ ). NPP was then 643 allocated into the four plant carbon pools (C'leaf, C'wood, C'root, and C'myco), with the 644 respective fitted allocation coefficients (aleaf, awood, aroot, and amyco) being inferred. It has been 645 shown that plant carbon allocation to mycorrhizal fungi may be an important flux in forest carbon budget calculation<sup>74</sup>. Turnover rates of C'<sub>leaf</sub>, C'<sub>root</sub>, C'<sub>myco</sub>, C'<sub>aglit</sub>, C'<sub>bglit</sub>, C'<sub>micr</sub> and 646 C'soil were represented by the corresponding turnover coefficients ( $\tau_{leaf}$ ,  $\tau_{wood}$ ,  $\tau_{root}$ ,  $\tau_{myco}$ ,  $\tau_{aglit}$ , 647  $\tau_{\text{bglit}}$ ,  $\tau_{\text{micr}}$ ,  $\tau_{\text{soil}}$ ), all of which were assumed unknowns except  $\tau_{\text{wood}}$  (estimated based on litter 648 649 basket data of twigs, barks and seeds) and  $\tau_{aglit}$  (estimated from the leaf litter decomposition data). For carbon leaving from C'aglit, C'bglit and C'micr, we inferred the corresponding 650 651 fractional coefficient that determines the fraction of carbon entering into the next pool (f'aglit, f'bglit, and f'micr), and assumed the remainder to be respired as part of Rhetero. The turnover of 652 653 soil carbon (i.e.  $\tau_{soil}$ ) also contributed to R<sub>hetero</sub>. In total, we fitted 2 pools, 4 allocation 654 coefficients, 6 turnover rates, and 3 fractional coefficients using the MCMC algorithm.

655

656 We used plot-level estimates of GPP,  $R_a$ ,  $R_{hetero}$ , carbon pools and changes in pools to 657 constrain the MCMC fitting. We assumed uniform parameter distributions and a burn-in 658 coefficient of 10%. Chain lengths were set at 200,000 for the ambient  $CO_2$  plots and 500,000 659 for the elevated plots. The longer chain length for the elevated plots was due to the smaller 660 proposal step size for these plots to meet an acceptance rate of around 20%. We reported the 661 means and standard deviation of the estimated parameters at the treatment level in Extended 662 Data Figure 7.

663

# 664 Data statement

- 665 Data will be available via Figshare (DOI: 10.6084/m9.figshare.11634315) with the 666 publication of the manuscript. Code to process the data is available via GitHub
- 667 (<u>https://github.com/mingkaijiang/EucFACE\_Carbon\_Budget/releases/tag/V20200120</u>).

### 668 References for main text

- 669 1. Le Quéré, C.L. *et al.* Global carbon budget 2018. *Earth Syst. Sci. Data* 10, 2141-2194
  670 (2018).
- 671 2. Schimel, D., Stephens, B.B., and Fisher, J.B. Effect of increasing CO<sub>2</sub> on the
  672 terrestrial carbon cycle. *Proc. Natl. Acad. Sci. USA* 112, 436-441 (2015).
- Walker, A.P. *et al.* Decadal biomass increment in early secondary successional woody
  ecosystems is increased by CO<sub>2</sub> enrichment. *Nat. Commun.* 10, 454,
  https://doi.org/10.1038/s41467-019-08348-1 (2019).
- 4. Norby, R.J. and Zak, D.R. Ecological lessons from Free-Air CO<sub>2</sub> Enrichment (FACE)
  experiments. *Annu. Rev. Ecol. Evol. Syst.* 42, 181-203 (2011).
- 5. Leuzinger, S. and Hattenschwiler, S. Beyond global change: lessons from 25 years of
  CO<sub>2</sub> research. *Oecologia* 171, 639-651 (2013).
- 680 6. Arora, V.K. *et al.* Carbon-concentration and carbon-climate feedbacks in CMIP5
  681 Earth system models. *J. Clim.* 26, 5289-5214 (2013).
- 682 7. Ellsworth, D.S. *et al.* Elevated CO<sub>2</sub> does not increase eucalypt forest productivity on a
  683 low-phosphorus soil. *Nat. Clim. Change* 7, 279-282 (2017).
- 684 8. Körner, C. *et al.* Carbon flux and growth in mature deciduous forest trees exposed to
  685 elevated CO<sub>2</sub>. *Science* **309**, 1360-1362 (2005).
- 686 9. Ryan, M.G. Three decades of research at Flakaliden advancing whole-tree physiology,
  687 forest ecosystem and global change research. *Tree Physiol.* 33, 1123-1131 (2013).
- 688 10. Klein, T. *et al.* Growth and carbon relations of mature *Picea abies* trees under 5 years
  689 of free-air CO<sub>2</sub> enrichment. *J. Ecol.* 104, 1720-1733 (2016).
- 690 11. Norby, R.J. *et al.* Model-data synthesis for the next generation of forest free-air CO<sub>2</sub>
  691 enrichment (FACE) experiments. *New Phytol.* 209, 17-28 (2016).

- 692 12. Pugh, T.A.M. *et al.* Role of forest regrowth in global carbon sink dynamics. *Proc.*693 *Natl. Acad. Sci. USA* 116, 4382-4387 (2019).
- 694 13. Grassi, G. *et al.* The key role of forests in meeting climate targets requires science for
  695 credible mitigation. *Nat. Clim. Change* 7, 220-226 (2017).
- 696 14. Peñuelas, J. *et al.* Shifting from a fertilization-dominated to a warming-dominated
  697 period. *Nat. Ecol. Evol.* 1, 1438-1445 (2017).
- 698 15. Luo, Y. *et al.* Progressive nitrogen limitation of ecosystem response to rising
  699 atmospheric carbon dioxide. *BioScience* 54, 731-739.
- 700 16. DeLucia, E.H. *et al.* Net primary production of a forest ecosystem with experimental
  701 CO<sub>2</sub> enrichment. *Science* 285, 1177-1179 (1999).
- 702 17. Crous, K., Ósvaldsson, A., and Ellsworth, D.S. Is phosphorus limiting in a mature
  703 *Eucalyptus* woodland? Phosphorus fertilization stimulates stem growth. *Plant Soil*704 **391**, 293-305 (2015).
- 18. Medlyn, B.E. *et al.* Using models to guide field experiments: a priori predictions for
  the CO<sub>2</sub> response of a nutrient- and water-limited native Eucalypt woodland. *Global Change Biol.* 22, 2834-2851 (2016).
- 708 19. Medlyn, B.E. *et al.* Using ecosystem experiments to improve vegetation models. *Nat.*709 *Clim. Change* 5, 528-534 (2015).
- 710 20. Friedlingstein, P. *et al.* Uncertainties in CMIP5 climate projections due to carbon
  711 cycle feedbacks. *J. Clim.* 27, 511-526 (2014).
- 712 21. Yang, J. *et al.* Low sensitivity of gross primary production to elevated CO<sub>2</sub> in a
  713 mature Eucalypt woodland. *Biogeosciences*. DOI: <u>https://doi.org/10.5194/bg-2019-</u>
  714 <u>272</u> (2020).

- 715 22. De Lucia, E.H., Drake, J.E., Thomas, R.B., and Gonzalez-Meler, M. Forest carbon
- vise efficiency: is respiration a constant fraction of gross primary production? *Global Change Biol.* 13, 1157-1167 (2007).
- 718 23. Norby, R.J. Forest canopy productivity index. *Nature* **381**, 564 (1996).
- 719 24. Duursma, R.A. *et al.* Canopy leaf area of a mature evergreen Eucalyptus woodland
  720 does not respond to elevated atmospheric CO<sub>2</sub> but tracks water availability. *Global*721 *Change Biol.* 22, 1666-1676 (2016).
- 722 25. Drake, J.E. *et al.* Short-term carbon cycling responses of a mature eucalypt woodland
  723 to gradual stepwise enrichment of atmospheric CO<sub>2</sub> concentration. *Global Change*724 *Biol.* 22, 380-390 (2016).
- 725 26. Drake, J.E. *et al.* Three years of soil respiration in a mature eucalypt woodland
  726 exposed to atmospheric CO<sub>2</sub> enrichment. *Biogeochemistry* 139, 85-101 (2018).
- 727 27. Drake, J.E. *et al.* Increases in the flux of carbon belowground stimulate nitrogen
  728 uptake and sustain the long-term enhancement of forest productivity under elevated
  729 CO<sub>2</sub>. *Ecol. Lett.* 14, 349-357 (2011).
- 730 28. Hasegawa, S., Macdonald, C.A., and Power, S.A. Elevated carbon dioxide increases
  731 soil nitrogen and phosphorus availability in a phosphorus-limited *Eucalyptus*732 woodland. *Global Change Biol.* 22, 1628-1643 (2016).
- 29. Ochoa-Hueso, R. *et al.* Rhizosphere-driven increase in nitrogen and phosphorus
  availability under elevated atmospheric CO<sub>2</sub> in a mature *Eucalyptus* woodland. *Plant Soil* 416, 283-295 (2017).
- 30. Crous, K.Y., Wujeska-Klause, A., Jiang, M., Medlyn, B.E. and Ellsworth, D.S.
  Nitrogen and phosphorus retranslocation of leaves and stemwood in a mature *Eucalyptus* forest exposed to 5 years of elevated CO<sub>2</sub>. *Front. Plant Sci.* 10:664, doi:
  10.3389/fpls.2019.00664 (2019).

740	31. Zaehle, S. et al. Evaluation of 11 terrestrial carbon-nitrogen cycle models against
741	observations from two temperature Free-Air CO2 Enrichment studies. New Phytol.
742	<b>202</b> , 803-822 (2014).
743	32. Fleischer, K. et al. Amazon forest response to CO2 fertilization dependent on plant
744	phosphorus acquisition. Nat. Geosci. 12, 736-741 (2019).
745	33. Todd-Brown, K.E.O. et al. Changes in soil organic carbon storage predicted by earth
746	system models during the 21 <sup>st</sup> century. <i>Biogeosciences</i> , <b>11</b> , 2341-2356 (2014).
747	34. Kuzyakov, Y., Horwath, W.R., Dorodnikov, M. and Blagodatskaya, E. Review and
748	synthesis of the effects of elevated atmospheric CO <sub>2</sub> on soil processes: no changes in
749	pools, but increased fluxes and accelerated cycles. Soil Biol. Biochem. 128, 66-78
750	(2019).
751	35. Luyssaert, S. et al. Old-growth forests as global carbon sinks. Nature 455, 213-215
752	(2008).
753	36. Jones, C. et al. 21st century compatible CO2 emissions and airborne fraction
754	simulated by CMIP5 Earth System models under 4 representative concentration
755	pathways. J. Clim. 26, doi:10.1175-JCLI-D-12-00554.1 (2013).
756	
757	
758	Reference for Methods
759	37. Australia Government Department of Agriculture, Fisheries and Forestry. Australia's
760	agriculture, fisheries and forestry at a glance 2012. Canberra, Australia (2012).
761	38. Food and Agricultural Organization of the United Nations. Global Forest Resources
762	Assessment 2000. FAO Forestry Paper 140. Rome, Italy (2001).

- 39. Gimeno, T.E., McVicar, T.R., O'Grady, A.P., Tissue, D.T. and Ellsworth, D.S.
  Elevated CO<sub>2</sub> did not affect the hydrological balance of a mature native *Eucalyptus*woodland. *Global Change Biol.* 24, 3010-3024 (2018).
- 40. Hasegawa, S. *et al.* Elevated CO<sub>2</sub> concentrations reduce C<sub>4</sub> cover and decrease
  diversity of understorey plant community in a *Eucalyptus* woodland. *J. Ecol.* 106,
  1483–1494 (2018).
- 769 41. Pathare, V.S. *et al.* Water availability affects seasonal CO<sub>2</sub>-induced photosynthetic
  770 enhancement in herbaceous species in a periodically dry woodland. *Global Change*771 *Biol.* 23, 5164–5178 (2017).
- 42. Paul, K.I. *et al.* Development and testing of allometric equations for estimating aboveground biomass of mixed-species environmental plantings. *For. Ecol. Manage.* 310,
  483-494 (2013).
- 43. Collins, L. *et al.* Understorey productivity in temperate grassy woodland responds to
  soil water availability but not to elevated CO<sub>2</sub>. *Global Change Biol.* 24, 2366-2376
  (2018).
- 44. Snowdon, P. *et al.* National carbon accounting system technical report no. 17.
  Australian Greenhouse Office, Canberra, Australia (2000).
- 45. Wallander, H. *et al.* Evaluation of methods to estimate production, biomass and
  turnover of ectomycorrhizal mycelium in forests soils A review. *Soil Biol. Biochem.*57, 1034–1047 (2013).
- 46. Buyer, J.S. and Sasser, M. High throughput phospholipid fatty acid analysis of soils. *Appl. Soil Ecol.* 61, 127–130 (2012).
- 47. Gherlenda, A.N., Esveld, J.L., Hall, A.A.G., Duursma, R.A. and Riegler, M. Boom
  and bust: rapid feedback responses between insect outbreak dynamics and canopy leaf
  area impacted by rainfall and CO<sub>2</sub>. *Global Change Biol.* 22, 3632-3641 (2016).

- 788 48. Facey, S.L. *et al.* Atmospheric change causes declines in woodland arthropods and
  789 impacts specific trophic groups. *Agr. Forest Entomol.* 19, 101-112 (2017).
- 49. Murray, T.J., Tissue, D.T., Ellsworth, D.S. and Riegler, M. Interactive effects of preindustrial, current and future CO<sub>2</sub> and temperature on an insect herbivore of *Eucalyptus. Oecologia* 171, 1025-1035 (2013).
- 50. Trakimas, G. *et al.* Ecological Stoichiometry: a link between developmental speed
  and physiological stress in an omnivorous insect. *Front. Behav. Neurosci.*13:42, https://doi.org/10.3389/fnbeh.2019.00042 (2019).
- Farquhar, G.D. von Caemmerer, S. and Berry, J.A. A biochemical model of
  photosynthetic CO<sub>2</sub> assimilation in leaves of C3 species. *Planta* 149, 78–90 (1980).
- 52. Medlyn, B.E. *et al.* Reconciling the optimal and empirical approaches to modelling
  stomatal conductance. *Global Change Biol.* 17, 2134–2144 (2011).
- 800 53. Gimeno, T.E. *et al.* Conserved stomatal behavior under elevated CO<sub>2</sub> and varying
  801 water availability in a mature woodland. *Funct. Ecol.* **30**, 700-709 (2016).
- 54. Yang, J. *et al.* Incorporating non-stomatal limitation improves the performance of leaf
  and canopy models at high vapor pressure deficit. *Tree Physiol.* DOI:
  <u>https://doi.org/10.1093/treephys/tpz103</u> (2019).
- 805 55. Martins, C.S.C. *et al.* Identifying environmental drivers of greenhouse gas emissions
  806 under warming and reduced rainfall in boreal-temperate forests. *Funct. Ecol.* 31,
  807 2356–2368 (2017).
- 56. Zhang, X. and Wang, W. The decomposition of fine and coarse roots: their global
  patterns and controlling factors. *Sci. Rep.* 5:09940 (2015).
- 810 57. Reich, P.B. *et al.* Plant diversity enhances ecosystem responses to elevated CO<sub>2</sub> and
  811 nitrogen deposition. *Nature* 410, 809-810 (2001).

- S8. Gherlenda, A.N., Moore, B.D., Haigh, A.M., Johnson, S.N. and Riegler, M. Insect
  herbivory in a mature *Eucalyptus* woodland canopy depends on leaf phenology but
  not CO<sub>2</sub> enrichment. *BMC Ecol.* 16, 47 (2016).
- Sp. Gherlenda, A.N. *et al.* Precipitation, not CO<sub>2</sub> enrichment, drives insect herbivore frass
  deposition and subsequent nutrient dynamics in a mature *Eucalyptus* woodland. *Plant Soil* 399, 29-39 (2016).
- 60. Drake, J.E. *et al.* The partitioning of gross primary production for young *Eucalyptus tereticornis* trees under experimental warming and altered water availability. *New Phytol.* 222, 1298-1312 (2019).
- 821 61. Salomón, R.L., Steppe, K., Crous, K.Y., Noh, N.J. and Ellsworth, D.S. Elevated CO<sub>2</sub>
  822 does not affect stem CO<sub>2</sub> efflux nor stem respiration in dry *Eucalyptus* woodland, but
  823 it shifts the vertical gradient in xylem CO<sub>2</sub>. *Plant Cell Environ.* 42, 2151-2164 (2019).
- 824 62. Raumonen, P. *et al.* Fast Automatic Precision Tree Models from Terrestrial Laser
  825 Scanner Data. *Remote Sens.* 5, 491-520 (2013).
- 63. Calders, K. *et al.* Nondestructive estimates of above-ground biomass using terrestrial
  laser scanning. *Methods Ecol. Evol.* 6, 198-208 (2015).
- 64. Davidson, E.A., Samanta, S., Caramori, S.S. and Savage, K. The Dual Arrhenius and
  Michaelis–Menten kinetics model for decomposition of soil organic matter at hourly
  to seasonal time scales. *Global Change Biol.* 18, 371-384 (2012).
- 65. Guenther, A.B. *et al.* The Model of Emissions of Gases and Aerosols from Nature
  version 2.1 (MEGAN2.1): an extended and updated framework for modeling biogenic
  emissions. *GeoSci. Model Dev.* 5, 1471-1492 (2012).
- 834 66. Emmerson, K.M., Palmer, P.I., Thatcher, M., Haverd, V. and Guenther, A.B.
  835 Sensitivity of isoprene emissions to drought over south-eastern Australia: Integrating

- models and satellite observations of soil moisture, *Atmos. Environ.* 209, 112-124
  (2019).
- 67. Kännaste, A., Copolovici, L. and Niinemets, Ü. Gas chromatography massspectrometry method for determination of biogenic volatile organic compounds
  emitted by plants. In: Plant isoprenoids: methods and protocols. Eds. M. RodríguezConcepción, Humana Press, New York. pp. 161-169 (2014).
- 842 68. Tholl, D. *et al.* Practical approaches to plant volatile analysis. *Plant J.* 45, 540 560
  843 (2006).
- 844 69. Li, T., Holst, T., Michelsen, A. and Rinnan, R. Amplification of plant volatile defense
  845 against insect herbivory in a warming Arctic tundra. *Nat. Plants* 5, 568-475 (2019).
- 70. Johnsen, L.G., Skou, P.B., Khakimov, B., and Bro, R. Gas chromatography mass
  spectrometry data processing made easy. *J. Chromatogr. A.* 1503, 57-64 (2017).
- 848 71. Keith, H. *et al.* Multiple measurements constrain estimates of net carbon exchange by
  849 a *Eucalyptus* forest. *Agric. For. Meteorol.* 149, 535-558 (2009).
- 850 72. Bates, D., Machler, M., Bolker, B.M., and Walker, S.C. Fitting linear mixed-effects
  851 models using lme4. *J. Stat. Softw.* 67, 1-48 (2015).
- 852 73. R Core Team. R: A language and environment for statistical computing. R Foundation
  853 for Statistical Computing, Vienna, Austria. URL https://www.R-project.org/ (2018).
- 854 74. Ouimette, A.P. *et al.* Accounting for carbon flux to mycorrhizal fungi may resolve
  855 discrepancies in forest carbon budgets. *Ecosystems* <u>https://doi.org/10.1007/s10021-</u>
  856 019-00440-3 (2019).
- 857 75. Harrison, I., Jones, P.D., Osborn, T.J. and Lister, D.H. Updated high-resolution grids
  858 of monthly climatic observations the CRU TS3.10 dataset. *Int. J. Climatol.* 34, 623859 642 (2014).

- 860 76. Olson, D.M. *et al.* Terrestrial ecoregions of the world: a new map of life on Earth.
  861 *BioScience* 51, 933-938 (2001).
- 862 77. Jiang, M., Felzer, B.S., Nielsen, U.N. and Medlyn, B.E. Biome-specific climatic space
  863 defined by temperature and precipitation predictability. *Global Ecol. Biogeogr.* 26,
  864 1270-1282 (2017).
- 865 78. Scarascia-Mugnozza, G. *et al.* Response to elevated CO<sub>2</sub> of a short rotation,
  866 multispecies Poplar plantation: the POPFACE/EUROFACE experiment. In Managed
  867 Ecosystems and CO<sub>2</sub>. Eds. Nösberger, J., *et al.* Berlin, Heidelberg, New York:
  868 Springer Verlag. pp 173–195 (2006).
- 79. Linden, S. NPP Boreal Forest: Flakaliden, Sweden, 1986-1996, R1. Data set.
  Available on-line [http://daac.ornl.gov] from Oak Ridge National Laboratory
  Distributed Active Archive Center, Oak Ridge, Tennessee, USA.
  DOI:10.3334/ORNLDAAC/201 (2013).
- 873 80. Anderson-Teixeira, K.J. *et al.* ForC: a global database of forest carbon stock and
  874 fluxes. *Ecology* 99, 1507-1507 (2018).
- 875 81. Shangguan, W. Dai, Y., Duan, Q., Liu, B. and Yuan, H. A global soil data set for
  876 earth system modelling. *J. Adv. Model Earth Sys.* 6, 249-263 (2014).
- 877 82. Yang, X., Post, W.M., Thornton, P.E. and Jain, A. Global gridded soil phosphorus
  878 distribution maps at 0.5-degree resolution. Data set. Available on-line
  879 [http://daac.ornl.gov] from Oak Ridge National Laboratory Distributed Active
  880 Archive Center, Oak Ridge, Tennessee, USA.
  881 <u>http://dx.doi.org/10.3334/ORNLDAAC/1223</u> (2014).
- 882

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#### 905 Author contributions

906 MJ, BEM, RAD and JED designed the synthesis, compiled the data, and performed the

- analyses. MJ, BEM, JED, RAD, ICA, CVMB, MMB, YC, LC-G, LC, KYC, BMDS, SLF,
- 908 ANG, TEG, SH, SNJ, AK, CAM, KM, BDM, LN, EHJN, UNN, ÜN, NJN, RO-H, VSP, EP,
- 909 JP, JP, JRP, SAP, PBR, AAR, MR, RR, PR, RLS, BKS, BS, MGT, JKMW, AW-K, JY and
- 910 DSE collected data and contributed to data analyses. MJ performed data assimilation analysis,
- 911 with contributions from MGDK and BEM. JY and BEM performed the MAESPA model

simulations, with contributions from MGDK and RAD. JED and AAR performed soil

- 913 respiration gap-filling and modelling. KME performed the MEGAN model simulation. MJ
- and LC-G conceptualized Figure 1, and LC-G implemented the graphic design. MJ wrote the
- 915 initial manuscript, with significant input from BEM, JED, BS, PBR, SZ, MGDK, MGT and
- 916 DSE. All authors edited and approved the manuscript.
- 917

912

### 918 Competing financial interests

- 919 None declared.
- 920

## 921 Materials and Correspondence

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926	Figure 1. A comprehensive carbon budget under ambient and elevated CO <sub>2</sub> treatment
927	in a mature forest ecosystem. Diamond boxes are gross primary production for overstorey
928	$(GPP_o)$ and understorey $(GPP_u)$ , respectively. Squared boxes are average carbon stocks over
929	the experimental period ( $C_{pools}$ , g C m <sup>-2</sup> ), including overstorey leaf ( $C_{ol}$ ), stem ( $C_{stem}$ ), coarse
930	root ( $C_{croot}$ ), fine root ( $C_{froot}$ ), intermediate root ( $C_{iroot}$ ), understorey aboveground ( $C_{ua}$ ), leaf
931	litter (C <sub>lit</sub> ), soil (C <sub>soil</sub> ), microbe (C <sub>micr</sub> ), aboveground insect (C <sub>ins</sub> ), and mycorrhizae (C <sub>myco</sub> ).
932	Unboxed variables are carbon fluxes (g C m <sup>-2</sup> yr <sup>-1</sup> ), including net primary production of
933	overstorey leaf (NPPol), stem (NPPstem), coarse root (NPPcroot), fine root (NPPfroot),
934	intermediate root (NPP <sub>iroot</sub> ), and understorey aboveground (NPP <sub>ua</sub> ), overstorey leaf
935	consumption by insects (NPP <sub>ins</sub> ), respiration fluxes of overstorey leaf ( $R_{ol}$ ), stem ( $R_{stem}$ ), root
936	( $R_{root}$ ), understorey aboveground ( $R_{ua}$ ), growth ( $R_{grow}$ ), insect ( $R_{ins}$ ), heterotroph ( $R_{hetero}$ ), and
937	soil (R <sub>soil</sub> ), and volatile carbon emission (VC), frass production (Frass), dissolved organic
938	carbon (DOC), and soil methane net uptake (CH <sub>4</sub> ). Solid arrow lines are fluxes entering a
939	pool, dotted arrow lines are fluxes leaving a pool. The changes in each carbon pool over time
940	$(\Delta C_{\text{pools}}, \text{g C m}^{-2} \text{ yr}^{-1})$ are reported in Extended Data Figure 6. Blue italic values are means $\pm$
941	one standard deviation of the ambient CO <sub>2</sub> treatment (n=3), whereas red values are means $\pm$
942	one standard deviation of the elevated $CO_2$ treatment (n=3). All values are normalized by a
943	linear mixed-model with plot-specific pre-treatment leaf area index as a covariate to account
944	for pre-existing differences. A summary of variable definitions and data availability is
945	provided in Extended Data Table 1.

946 Figure 2. The fate of additional carbon fixed under elevated  $CO_2$  (eCO<sub>2</sub>) in a mature 947 forest ecosystem. a) Column "GPP" represents the total eCO2-induced increases in 948 overstorey and understorey gross primary production (GPP<sub>o</sub> and GPP<sub>u</sub>, respectively), "NPP + 949  $R_a$ " represents the sum of net primary production and autotrophic respiration response, "R +  $\Delta C_{\text{pools}}$ " represents the sum of ecosystem respiration and change in carbon storage response. 950 951 **b**) The relative contributions of individual NPP fluxes to the aggregated NPP response to 952 eCO<sub>2</sub>, including NPP responses of overstorey leaf (NPP<sub>ol</sub>), twigs, barks and seeds (NPP<sub>other</sub>), 953 fine root (NPP<sub>froot</sub>), and understorey aboveground (NPP<sub>ua</sub>); c) The relative contributions of 954 individual respiratory fluxes to the aggregated R response to eCO<sub>2</sub>, including respiration responses of stem (Rstem), root (Rroot), understorey aboveground (Rua), growth (Rgrow), and soil 955 956 heterotroph (R<sub>hetero</sub>); and d) The relative contributions of individual change in carbon storage 957 to the aggregated  $\Delta C_{\text{pools}}$  response to eCO<sub>2</sub>, including changes in pool of stem ( $\Delta C_{\text{stem}}$ ), 958 understorey aboveground ( $\Delta C_{ua}$ ), fine root ( $\Delta C_{froot}$ ), leaf litter ( $\Delta C_{lit}$ ), and soil ( $\Delta C_{soil}$ ). Variables with an absolute mean CO<sub>2</sub> effect of  $< 5 \text{ g C m}^{-2} \text{ yr}^{-1}$  are not reported in the bar 959 960 chart for better visual clarification. Individual CO<sub>2</sub> responses are reported in Extended Data 961 Figure 5. Each color represents the  $CO_2$  response of a flux variable, the point indicates the net 962 sum of all variables for a column, and the grey error bar represents one standard deviation of 963 the estimated column sum at the plot-level (see Methods). The CO<sub>2</sub> effect is estimated using a 964 linear mixed-model analysis with plot-specific pre-treatment leaf area index as a covariate to 965 account for pre-existing differences (see Methods). The non-normalized response is provided 966 in Extended Data Figure 4, which generally agrees with findings present in this figure, but 967 with larger uncertainty.

### 968 Figure 3. Estimates of net ecosystem production (NEP) under ambient and elevated CO<sub>2</sub>

969 treatment at EucFACE. Positive values indicate ecosystem net carbon uptake by the 970 ecosystem. "In - Out" calculates NEP based on the difference between total influxes and total 971 outfluxes. "NPP - Rhetero" calculates NEP based on the difference between net primary 972 production (NPP) and heterotrophic respiration (R<sub>hetero</sub>). " $\Delta C_{pools}$ " derives NEP based on 973 incremental changes in all ecosystem carbon pools. Colored bars indicate treatment means 974 based on each method (n=3), with blue representing ambient and red representing elevated 975 CO<sub>2</sub> treatment. Individual dots are plot-level NEP, derived based on different methods (see 976 Methods). Values are normalized by a linear mixed-model with plot-specific pre-treatment 977 leaf area index as a covariate to account for pre-existing differences. Horizontal dotted line 978 indicates NEP equals zero. The inset figure includes an inferred production allocation flux to 979 mycorrhizal fungi (NPP<sub>myco</sub>) based on data assimilation (Methods), which affected NEP 980 estimates based on the NPP – R<sub>hetero</sub> method only.

981 Extended Data Table 1. Definition and data availability of variables. Data availability 982 includes start and end year of data included in this study. Time points indicate the number of 983 data collections over the available data period. Within plot sub-replicate indicate the number 984 of replicates within each treatment plot. The detailed methods for estimating each variable is 985 provided in the Method section.

Variable	Data coverage				
Name	Symbol	Start year	End year	Time points	Within plot sub- replicate (plot <sup>-1</sup> )
Specific Leaf Area	SLA	2013	2016	50	3
Leaf Area Index	LAI	2012	2016	303	1
Soil bulk density	ВК	2017	2017	2	3
Diameter at breast height	DBH	2013	2016	4	Individual tree
Overstorey leaf pool	C <sub>ol</sub>	2012	2016	303	1
Understorey aboveground pool	C <sub>ua</sub>	2015	2016	16	4
Overstorey stem C pool	C <sub>stem</sub>	2013	2016	4	Individual tree
Fine root C pool	C <sub>froot</sub>	2014	2016	7	4
Intermediate root C pool	Ciroot	2014	2016	7	4
Coarse root C pool	C <sub>croot</sub>	2013	2016	4	Individual tree
Forest floor leaf litter C pool	C <sub>lit</sub>	2013	2016	46	-
Microbial C pool	C <sub>micr</sub>	2012	2015	15	4
Soil C pool	C <sub>soil</sub>	2012	2014	11	4

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Mycorrhizal C pool	C <sub>myco</sub>	2015	2015	3	-
Insect C pool (aerial)	C <sub>ins</sub>	2013	2016	43	8
Insect C pool (understorey)	C <sub>ins</sub>	2014	2015	5	2
Overstorey gross primary production	GPPo	2013	2016	Annual	1
Understorey gross primary production	GPP <sub>u</sub>	2013	2016	Annual	1
Overstorey leaf respiration	R <sub>ol</sub>	2013	2016	Annual	1
Understorey leaf respiration	R <sub>ua</sub>	2013	2016	Annual	1
Stem respiration	R <sub>stem</sub>	2012	2016	Daily	3
Root respiration	R <sub>root</sub>	2012	2015	Daily	-
Methane net flux	CH <sub>4</sub>	2013	2016	35	7
Volatile C emission flux	VC	2013	2016	Daily	1
Insect herbivore respiration	R <sub>ins</sub>	2012	2014	22	-
Dissolved organic C loss flux	DOC	2012	2014	12	4
Soil respiration	R <sub>soil</sub>	2012	2015	Daily	8
Growth respiration	R <sub>grow</sub>	2012	2016	Annual	1
Overstorey leaf net primary production	NPP <sub>ol</sub>	2012	2016	49	8
Stem net primary production	NPP <sub>stem</sub>	2012	2016	4	Individual tree
Fine root net primary production	NPP <sub>froot</sub>	2014	2016	6	4
Intermediate root net primary production	NPP <sub>iroot</sub>	2014	2016	6	4

Coarse root net primary production	NPP <sub>croot</sub>	2012	2016	4	Individual tree
Other net primary production (sum of twigs, bark, seeds)	NPP <sub>other</sub>	2012	2016	49	8
Twig net primary production	NPP <sub>twig</sub>	2012	2016	49	8
Bark net primary production	NPP <sub>bark</sub>	2012	2016	49	8
Seed net primary production	NPP <sub>seed</sub>	2012	2016	49	8
Understorey aboveground net primary production	NPP <sub>ua</sub>	2015	2016	3	4
Frass production	Frass	2012	2014	22	8
Heterotrophic respiration	R <sub>hetero</sub>	2012	2016	Daily	8
Overstorey leaf insect consumption flux	NPP <sub>ins</sub>	2012	2014	22	-

# 987 Extended Data Table 2. Carbon (C) fraction used to convert from biomass into C

988 content.

Variable	Symbol	Mean value	Data source
C fraction of overstorey leaf pool	$f_{ol}$	0.5	EucFACE data
C fraction of understorey aboveground pool	fua	0.456	EucFACE data
C fraction of stem pool	<i>f</i> stem	0.445 (ambient plots) 0.448 (elevated plots)	EucFACE data
C fraction of coarse root pool	$f_{croot}$	0.445 (ambient plots) 0.448 (elevated plots)	Assumed the same as $f_{\text{stem}}$
C fraction of fine root pool	$f_{\it froot}$	0.40 (ambient plots) 0.42 (elevated plots)	EucFACE data
C fraction of intermediate root pool	Firoot	0.40 (ambient plots) 0.42 (elevated plots)	Assumed the same as $f_{\rm froot}$
C fraction of overstorey leaflitter pool	$f_{lit}$	0.5	EucFACE data
C fraction of aboveground insect pool	$f_{ins}$	0.5	Ref 49
C fraction of frass production	$f_{\it frass}$	0.53	EucFACE data
C fraction of microbial pool	$f_{micr}$	0.534 (ambient plots) 0.493 (elevated plots)	EucFACE data
C fraction of mycorrhizal pool	$f_{myco}$	0.534 (ambient plots) 0.493 (elevated plots)	Assumed the same as $f_{micr}$
C fraction of soil pool	$f_{soil}$	0.016 (ambient plots) 0.017 (elevated plots)	EucFACE data
C fraction of twigs, barks and seeds production	fother	0.5	Assumed

989

- 991 Extended Data Figure 1. The Eucalyptus Free Air Carbon dioxide Enrichment
- 992 experiment facility (EucFACE). a) View of the forest and facility from above (photo credit:
- 993 David S. Ellsworth), b) view of the understorey vegetation and infrastructure inside a plot
- 994 (photo credit: Mingkai Jiang), and c) view from below of the canopy structure and the crane
- 995 (photo credit: Mingkai Jiang).

997 Extended Data Figure 2. Mean annual temperature (MAT) and mean annual 998 precipitation (MAP) for major forest biomes and a selected list of tree-based elevated CO<sub>2</sub> experiments. Gridded temperature and precipitation data were obtained from the 999 Climate Research Unit (CRU) monthly dataset at 0.5 resolution<sup>75</sup>. Global biome boundaries 1000 1001 and definitions were taken from Ref 76 and were spatially aggregated onto the CRU 1002 resolution, following Ref 77. The major forest biomes are defined as: tropical and subtropical 1003 moist broadleaf forests; tropical and subtropical dry broadleaf forests; tropical and 1004 subtropical coniferous forest; temperate broadleaf and mixed forests; temperate coniferous 1005 forests; boreal forests/taiga; and Mediterranean forests, woodlands, and scrub. The list of 1006 elevated CO<sub>2</sub> experiments includes 7 Free Air CO<sub>2</sub> Enrichment experiments (FACE) and a 1007 Whole-Tree Chamber experiment (WTC), namely: EucFACE, DukeFACE, ORNLFACE, 1008 AspenFACE, PopFACE, WebFACE, BiForFACE, and FlakalidenWTC. The site-specific climate, tree age and net primary production (NPP) under ambient CO2 treatment were 1009 1010 collected from Ref 3, 9, 10, 11, 78 and 79. The top inset figure compares global forest NPP 1011 against standing age using data collected from Ref 80. We included data with forest age <1012 500 years, and the NPP reported in Ref 80 included both overstorey and understorey. The 1013 bottom inset figure compares soil total nitrogen and labile phosphorus across the eCO<sub>2</sub> experiments. Soil total nitrogen was extracted from Ref 81 using spatial coordinates of each 1014 1015 experiment, while soil labile phosphorus was spatially extracted from Ref 82. The two dotted 1016 lines indicates N:P ratios of 20:1 and 100:1, respectively.

1017 Extended Data Figure 3. Estimates of (a and b) gross primary production (GPP) and (c 1018 and d) soil respiration (R<sub>soil</sub>) based on different methods for both (a and c) ambient and 1019 (b and d) elevated CO<sub>2</sub> treatment at EucFACE. For estimates of GPP, we compared the model simulated total GPP of overstorey and understorey (GPPo and GPPu, respectively), 1020 1021 with the sum of data-driven estimates of net primary production (NPP) and autotrophic 1022 respiration ( $R_a$ ), which include NPP of overstorey leaf (NPP<sub>ol</sub>), stem (NPP<sub>stem</sub>), fine root 1023 (NPP<sub>froot</sub>), intermediate root (NPP<sub>iroot</sub>), coarse root (NPP<sub>croot</sub>), twigs, barks and seeds 1024 (NPP<sub>other</sub>), understorey aboveground (NPP<sub>ua</sub>), leaf consumption by insects (NPP<sub>ins</sub>), and 1025 respiratory fluxes of overstorey leaf ( $R_{ol}$ ), stem ( $R_{stem}$ ), root ( $R_{root}$ ), understorey aboveground 1026 (R<sub>ua</sub>), growth (R<sub>grow</sub>), and volatile carbon emission (VC). For estimates of R<sub>soil</sub>, we compared 1027 direct estimates of R<sub>soil</sub> scaled up from soil chamber measurements, with the sum of litterfall 1028 and independent estimates of root respiration (Litter + R<sub>root</sub>), assuming no net change in soil 1029 carbon stock over time. Here litterfall was inferred based on NPP of overstorey leaf (NPP<sub>ol</sub>), fine root (NPP<sub>froot</sub>), intermediate root (NPP<sub>iroot</sub>), twigs, barks and seeds (NPP<sub>other</sub>), understorey 1030 1031 aboveground (NPP<sub>ua</sub>), and frass production (Frass). These evaluations provide independent 1032 mass balance checks of the estimated ecosystem carbon budget. Each color represents a flux 1033 variable. Dotted point and vertical line represent treatment mean and standard deviation based on plot-level estimates of the aggregated flux (n=3). Values were normalized by a 1034 1035 linear mixed-model with pre-treatment leaf area index as a covariate to account for predifferences. 1036 existing

1037 Extended Data Figure 4. The fate of additional carbon fixed under elevated CO<sub>2</sub> (eCO<sub>2</sub>) 1038 in a mature forest ecosystem (non-normalized analysis case). a) Column "GPP" 1039 represents the total eCO<sub>2</sub> induced increase in overstorey and understorey gross primary production (GPP<sub>o</sub> and GPP<sub>u</sub>, respectively), column "NPP + R<sub>a</sub>" represents the sum of net 1040 primary production and autotrophic respiration eCO<sub>2</sub> response, and column "R +  $\Delta C_{\text{pools}}$ " 1041 1042 represents the sum of ecosystem respiration and carbon storage  $eCO_2$  response. b) The 1043 relative contributions of individual NPP fluxes to the aggregated NPP response to eCO<sub>2</sub>, 1044 including overstorey leaf (NPPol), stem (NPPstem), fine root (NPPfroot) and understorey 1045 above ground ( $NPP_{ua}$ ). c) The relative contributions of individual respiratory fluxes to the 1046 aggregated R response to eCO<sub>2</sub>, including overstorey leaf (R<sub>ol</sub>), stem (R<sub>stem</sub>), root (R<sub>root</sub>), 1047 understorey above ground  $(R_{ua})$ , and heterotroph  $(R_{hetero})$ . d) The relative contributions of 1048 individual change in carbon storage to the aggregated  $\Delta C_{\text{pools}}$  response to eCO<sub>2</sub>, including 1049 stem ( $\Delta C_{\text{stem}}$ ), fine root ( $\Delta C_{\text{froot}}$ ), leaflitter ( $\Delta C_{\text{lit}}$ ), understorey aboveground ( $\Delta C_{\text{ua}}$ ), and soil ( $\Delta C_{soil}$ ). Variables with an average CO<sub>2</sub> effect of < 5 g C m<sup>-2</sup> yr<sup>-1</sup> were excluded from the 1050 figure for better visual clarification. Each color represents a flux variable, point indicates the 1051 1052 net sum of all variables for a column, and the grey confidence interval represents plot-level 1053 of column standard deviation (n=3) the estimated sum.

Extended Data Figure 5.  $CO_2$  treatment effect (g C m<sup>-2</sup> vr<sup>-1</sup>) for all ecosystem fluxes at 1055 **EucFACE.** a) The CO<sub>2</sub> response of gross ecosystem carbon uptake, including gross primary 1056 1057 production of overstorey (GPP<sub>0</sub>) and understorey (GPP<sub>u</sub>), and soil methane uptake (CH<sub>4</sub>). **b**) 1058 The eCO<sub>2</sub> response of annual incremental change in carbon pool ( $\Delta C_{\text{pools}}$ ), including overstorey leaf ( $\Delta C_{ol}$ ), stem ( $\Delta C_{stem}$ ), coarse root ( $\Delta C_{croot}$ ), fine root ( $\Delta C_{froot}$ ), intermediate 1059 1060 root ( $\Delta C_{iroot}$ ), understorey aboveground ( $\Delta C_{ua}$ ), leaf litter ( $\Delta C_{lit}$ ), soil ( $\Delta C_{soil}$ ), microbe 1061  $(\Delta C_{\text{micr}})$ , above ground insect  $(\Delta C_{\text{ins}})$ , and mycorrhizae  $(\Delta C_{\text{mvco}})$ . c) The eCO<sub>2</sub> response of net 1062 primary production (NPP), including overstorey leaf (NPP<sub>ol</sub>), stem (NPP<sub>stem</sub>), coarse root 1063 (NPP<sub>croot</sub>), fine root (NPP<sub>froot</sub>), intermediate root (NPP<sub>iroot</sub>), understorey aboveground (NPP<sub>ua</sub>), 1064 twigs, barks and seeds (NPP<sub>other</sub>), and leaf insect consumption (NPP<sub>ins</sub>). d) The eCO<sub>2</sub> 1065 response of ecosystem respiration (R) and other out-going flux, including respiration fluxes 1066 of overstorey leaf (R<sub>ol</sub>), stem (R<sub>stem</sub>), root (R<sub>root</sub>), understorey aboveground (R<sub>ua</sub>), growth 1067 (R<sub>grow</sub>), insect (R<sub>ins</sub>), heterotroph (R<sub>hetero</sub>), and soil (R<sub>soil</sub>), and volatile carbon emission (VC) 1068 and dissolved organic carbon leaching (DOC). Dots and grey bars represent means and standard deviations of the CO<sub>2</sub> treatment difference, predicted by a linear mixed-model with 1069

1070 plot-specific pre-treatment leaf area index as a covariate. Red dots indicate negative means

and blue dots indicate positive means. Dashed lines indicate change of scale along the x-axis.

- 1073 Extended Data Figure 6. Estimates of incremental change in carbon pool averaged over
- 1074 the experimental period under ambient (aCO<sub>2</sub>) and elevated CO<sub>2</sub> (eCO<sub>2</sub>) treatment
- 1075 effect at EucFACE ( $\Delta C_{\text{pools}}$ , g C m<sup>-2</sup> yr<sup>-1</sup>). The  $\Delta C_{\text{pools}}$  variables are overstorey leaf ( $\Delta C_{\text{ol}}$ ),
- 1076 stem ( $\Delta C_{stem}$ ), coarse root ( $\Delta C_{croot}$ ), fine root ( $\Delta C_{froot}$ ), intermediate root ( $\Delta C_{iroot}$ ), understorey
- 1077 aboveground ( $\Delta C_{ua}$ ), leaf litter ( $\Delta C_{lit}$ ), soil ( $\Delta C_{soil}$ ), microbe ( $\Delta C_{micr}$ ), aboveground insect
- 1078 ( $\Delta C_{ins}$ ), and mycorrhizae ( $\Delta C_{myco}$ ). Colored bars and black lines represent means and standard
- 1079 deviations for each treatment, with blue represents  $aCO_2$  and red represents  $eCO_2$  treatment.
- 1080 Dashed lines indicate change of scale along the x-axis.

1081 Extended Data Figure 7. Fitted carbon cycle parameters to trace the fate of the 1082 additional carbon under elevated CO<sub>2</sub> at EucFACE. Parameters were estimated by 1083 Markov Chain Monte Carlo (MCMC) fitting algorithm, assuming a simplified carbon cycle 1084 framework based on data collected from EucFACE. Details of the MCMC approach can be 1085 found in the Methods. Plot-level gross primary production (GPP), autotrophic respiration ( $R_a$ ), 1086 heterotrophic respiration (R<sub>hetero</sub>), carbon pools of leaf (C'<sub>leaf</sub>), wood (C'<sub>wood</sub>), root (C'<sub>root</sub>), 1087 mycorrhizae (C'<sub>myco</sub>), microbe (C'<sub>micr</sub>), and soil (C'<sub>soil</sub>), and the corresponding change in pools were used to constrain the model fitting. Net primary production (NPP) was derived as 1088 1089 the difference of GPP and R<sub>a</sub>. Carbon use efficiency (CUE') was calculated as NPP/GPP; it 1090 differs from the value given in the main text owing to the contribution of NPP allocated to 1091 mycorrhizae (NPP<sub>myco</sub>). We fitted two carbon pools (C'aglit and C'bglit), four allocation 1092 coefficients ( $a_{\text{leaf}}$ ,  $a_{\text{wood}}$ ,  $a_{\text{root}}$ , and  $a_{\text{myco}}$ ), six turnover rates ( $\tau_{\text{leaf}}$ ,  $\tau_{\text{root}}$ ,  $\tau_{\text{myco}}$ ,  $\tau_{\text{bglit}}$ ,  $\tau_{\text{micr}}$ , and  $\tau_{soil}$ ), and three fractional coefficients (f'aglit, f'bglit, and f'micr) using MCMC algorithm. The 1093 1094 fractional coefficients indicate the fraction of carbon leaving one pool that enters the 1095 subsequent with the remainder respired pool, as R<sub>hetero</sub>.

1097 Extended Data Figure 8. Data-model intercomparison of some key carbon cycle parameters, under ambient (aCO<sub>2</sub>) and elevated CO<sub>2</sub> (eCO<sub>2</sub>). Parameters include: a) 1098 1099 allocation coefficients to leaf, wood, root and other, b) turnover rates of leaf, root, aboveground litter (Aglit), belowground litter (Bglit), and c) turnover rate of soil. Models 1100 1101 include: Community Atmosphere Biosphere Land Exchange (CABL), Community Land 1102 Model 4 (CLM4), Community Land Model with a phosphorus component (CLMP), Generic 1103 Decomposition And Yield (GDAY), Lund-Potsdam-Jena General Ecosystem Simulator 1104 (LPJX), Orchidee-C-N (OCNX), and Sheffield Dynamic Global Vegetation Model (SDVM). 1105 The model output was generated as part of the model ensemble predictions made in advance 1106 of the experiment reported in Ref 17 for EucFACE. Data-based uncertainties were estimated 1107 using the Markov Chain Monte Carlo data assimilation algorithm, with error bars indicating 1108 one standard deviation. Allocation to other in the data refers to the allocation to mycorrhizal 1109 production, whereas it refers to the allocation to reproductive carbon pool in some models.



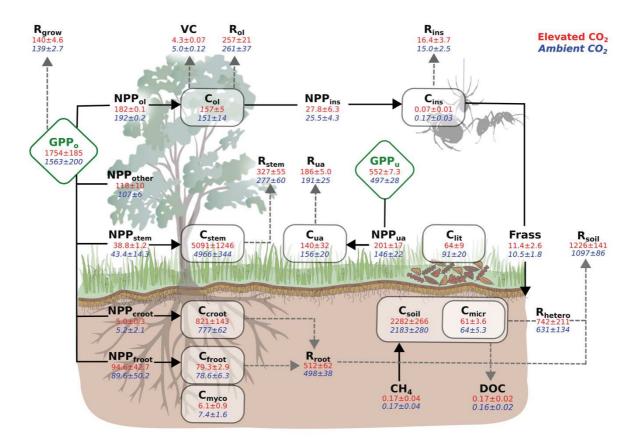
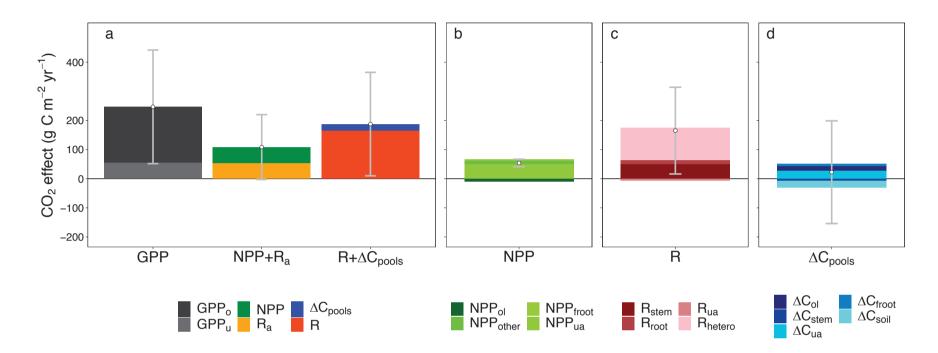


Figure 1. A comprehensive carbon budget under ambient and elevated CO<sub>2</sub> treatment in 764 a mature forest ecosystem. Diamond boxes are gross primary production for overstorey 765  $(GPP_{o})$  and understorey  $(GPP_{u})$ , respectively. Squared boxes are carbon stocks  $(gCm^{-2})$ , 766 767 including overstorey leaf ( $C_{ol}$ ), stem ( $C_{stem}$ ), coarse root ( $C_{croot}$ ), fineroot ( $C_{froot}$ ), understorey aboveground (C<sub>ua</sub>), leaf litter (C<sub>lit</sub>), soil (C<sub>soil</sub>), microbe (C<sub>micr</sub>), aboveground insect (C<sub>ins</sub>), and 768 mycorrhizae (C<sub>mvco</sub>). Unboxed variables are carbon fluxes (gCm<sup>-2</sup>yr<sup>-1</sup>), including net primary 769 770 production of overstorey leaf (NPP<sub>ol</sub>), stem (NPP<sub>stem</sub>), coarse root (NPP<sub>croot</sub>), fineroot (NPP<sub>froot</sub>), and understorey aboveground (NPP<sub>ua</sub>), overstorey leaf consumption by insects (NPP<sub>ins</sub>), 771 respiration fluxes of overstorey leaf (R<sub>ol</sub>), stem (R<sub>stem</sub>), root (R<sub>root</sub>), understorey aboveground 772 (R<sub>ua</sub>), growth (R<sub>grow</sub>), insect (R<sub>ins</sub>), heterotroph (R<sub>hetero</sub>), and soil (R<sub>soil</sub>), and volatile carbon 773 emission (VC), frass production (Frass), dissolved organic carbon (DOC), and soil methane net 774

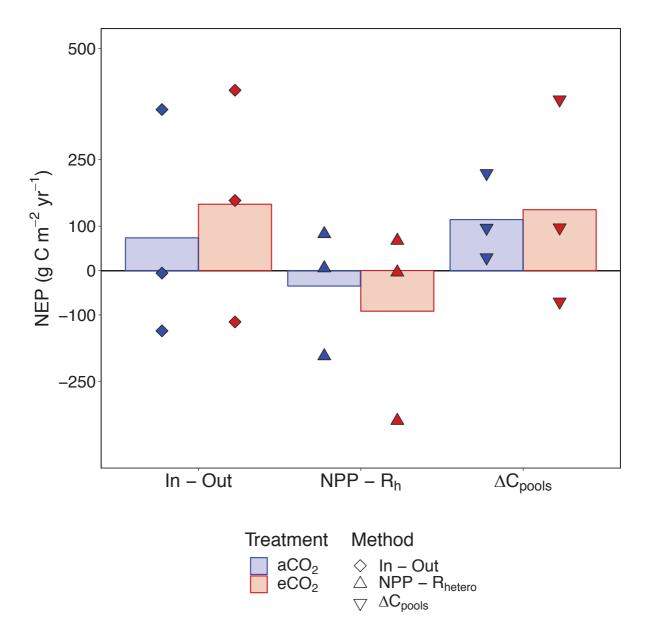
uptake (CH<sub>4</sub>). Solid arrow lines are fluxes entering a pool, dotted arrow lines are fluxes leaving a pool. Blue italic values are means  $\pm$  one standard deviation of the ambient CO<sub>2</sub> treatment (n=3), whereas red values are means  $\pm$  one standard deviation of the elevated CO<sub>2</sub> treatment (n=3). All values are normalized by a linear mixed-model with plot-specific pre-treatment leaf area index as a covariate to account for pre-existing differences. Summary of variable definitions and data availability is provided in Extended Data Table 1.



782

Figure 2. The fate of additional carbon fixed under elevated CO<sub>2</sub> (eCO<sub>2</sub>) in a mature forest ecosystem. a) Column "GPP" represents the total eCO<sub>2</sub>-induced increases in overstorey and understorey gross primary production (GPP<sub>o</sub> and GPP<sub>u</sub>, respectively), "NPP + R<sub>a</sub>" represents the sum of net primary production and autotrophic respiration response, "R +  $\Delta C_{pools}$ " represents the sum of ecosystem respiration and carbon storage response. b) The relative contributions of individual NPP fluxes to the aggregated NPP response to eCO<sub>2</sub>, including NPP responses of overstorey leaf (NPP<sub>ol</sub>), twigs, barks and seeds (NPP<sub>other</sub>), fineroot (NPP<sub>froot</sub>), and understorey aboveground (NPP<sub>ua</sub>); c) The relative contributions of individual respiratory fluxes to the aggregated R response to eCO<sub>2</sub>, including respiration responses of stem (R<sub>stem</sub>), root (R<sub>root</sub>), understorey aboveground

789 ( $R_{ua}$ ), and soil heterotroph ( $R_{hetero}$ ); and **d**) The relative contributions of individual change in carbon storage to the aggregated  $\Delta C_{pools}$  response to eCO<sub>2</sub>, including changes in pool of overstorey leaf ( $\Delta C_{ol}$ ), stem ( $\Delta C_{stem}$ ), understorey aboveground ( $\Delta C_{ua}$ ), fineroot ( $\Delta C_{froot}$ ), and soil ( $\Delta C_{soil}$ ). 790 Variables with an absolute mean  $CO_2$  effect of < 5 gCm<sup>-2</sup>yr<sup>-1</sup> are excluded from the figure for better visual clarification. Individual  $CO_2$  responses 791 are reported in Extended Data Figure 4. Each color represents the CO<sub>2</sub> response of a flux variable, point indicates the net sum of all variables for 792 a column, and the grey error bar represents one standard deviation of the estimated column sum at the plot-level (see Methods). The CO<sub>2</sub> effect is 793 estimated using a linear mixed-model analysis with plot-specific pre-treatment leaf area index as a covariate to account for pre-existing differences 794 (see Methods). The un-normalized response is provided in Extended Data Figure 3, which generally agrees with findings present in this figure, but 795 with less statistical precision. 796



**Figure 3. Estimates of net ecosystem production (NEP) under ambient and elevated CO**<sub>2</sub> **treatment at EucFACE**. Positive values indicate ecosystem net carbon uptake by the ecosystem. "In - Out" calculates NEP based on the difference between total influxes and total outfluxes. "NPP -  $R_{hetero}$ " calculates NEP based on the difference between net primary production (NPP) and heterotrophic respiration ( $R_{hetero}$ ). " $\Delta C_{pools}$ " derives NEP based on incremental changes in all ecosystem carbon pools. Colored bars indicate treatment means based on each method (n=3), with blue representing ambient and red representing elevated CO<sub>2</sub>

treatment. Individual dots are plot-level NEP, derived based on different methods (see
Methods). Values are normalized by a linear mixed-model with plot-specific pre-treatment leaf
area index as a covariate to account for pre-existing differences. Horizontal dotted line indicates
NEP equals zero.

810 Extended Data Table 1. Definition and data availability of variables. Data availability 811 includes start and end year of data included in this study. Time points indicate the number of 812 data collections over the available data period. Within plot sub-replicate indicate the number 813 of replicates within each treatment plot. The detailed methods for estimating each variable is 814 provided in the Method section.

Variable	Data coverage				
Name	Symbol	Start year	End year	Time points	Within plot sub- replicate (plot <sup>-1</sup> )
Specific Leaf Area	SLA	2013	2016	50	3
Leaf Area Index	LAI	2012	2016	303	1
Soil bulk density	ВК	2017	2017	2	3
Diameter at breast height	DBH	2013	2016	4	Individual tree
Overstorey leaf pool	C <sub>ol</sub>	2012	2016	303	1
Understorey aboveground pool	C <sub>ua</sub>	2015	2016	16	4
Overstorey stem C pool	C <sub>stem</sub>	2013	2016	4	Individual tree
Fine root C pool	C <sub>froot</sub>	2014	2016	6	4
Coarse root C pool	C <sub>croot</sub>	2013	2016	4	Individual tree
Forest floor leaf litter C pool	C <sub>lit</sub>	2013	2016	46	-
Microbial C pool	C <sub>micr</sub>	2012	2015	15	4
Soil C pool	C <sub>soil</sub>	2012	2014	11	4

Mycorrhizal C pool	C <sub>myco</sub>	2015	2015	3	-
Insect C pool (aeriel)	C <sub>ins</sub>	2013	2016	43	8
Insect C pool (ground dwelling)	C <sub>ins</sub>	2013	2015	5	4
Overstorey gross primary	GPPo	2013	2016	Annual	1
production					
Understorey gross primary	GPP <sub>u</sub>	2013	2016	Annual	1
production					
Overstorey leaf respiration	R <sub>ol</sub>	2013	2016	Annual	1
Understorey leaf respiration	R <sub>ua</sub>	2013	2016	Annual	1
Stem respiration	R <sub>stem</sub>	2012	2016	Daily	3
Root respiration	R <sub>root</sub>	2012	2015	Daily	-
Methane net flux	CH <sub>4</sub>	2013	2016	35	7
Volatile C emission flux	VC	2013	2016	Daily	1
Insect herbivore respiration	R <sub>ins</sub>	2012	2014	22	-
Dissolved organic C loss flux	DOC	2012	2014	12	4
Soil respiration	R <sub>soil</sub>	2012	2015	Daily	8
Growth respiration	R <sub>grow</sub>	2012	2016	Annual	1
Overstorey leaf net primary	NPP <sub>ol</sub>	2012	2016	49	8
production					
Stem net primary production	NPP <sub>stem</sub>	2012	2016	4	Individual tree

Fine root net primary production	NPP <sub>froot</sub>	2014	2016	5	4
Coarse root net primary production	NPP <sub>croot</sub>	2012	2016	4	Individual tree
Other net primary production (sum	NPP <sub>other</sub>	2012	2016	49	8
of twigs, bark, seeds)					
Twig net primary production	NPP <sub>twig</sub>	2012	2016	49	8
Bark net primary production	NPP <sub>bark</sub>	2012	2016	49	8
Seed net primary production	NPP <sub>seed</sub>	2012	2016	49	8
Understorey aboveground net	NPP <sub>ua</sub>	2015	2016	3	4
primary production					
Frass production	Frass	2012	2014	22	8
Heterotrophic respiration	R <sub>hetero</sub>	2012	2016	Daily	8
Overstorey leaf insect consumption	NPP <sub>ins</sub>	2012	2014	22	-
flux					

# 816 Extended Data Table 2. Carbon (C) fraction used to convert from biomass into C content.

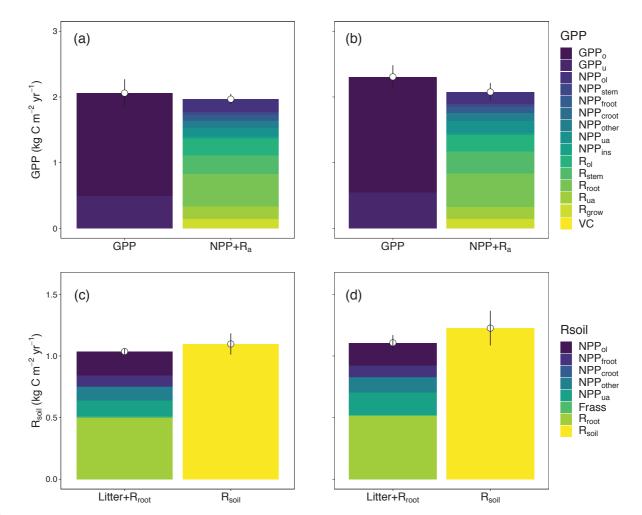
Variable	Symbol	Mean value	Data source
C fraction of overstorey leaf pool	$f_{ol}$	0.5	EucFACE data
C fraction of	f <sub>ua</sub>	0.456	EucFACE data
understorey			
aboveground pool			
C fraction of stem pool	$f_{stem}$	0.445 (ambient plots)	EucFACE data
		0.448 (elevated plots)	
C fraction of coarse	$\mathbf{f}_{croot}$	0.445 (ambient plots)	Assumed the same as
root pool		0.448 (elevated plots)	$\mathbf{f}_{stem}$
C fraction of fine root	$\mathbf{f}_{\mathrm{froot}}$	0.40 (ambient plots)	EucFACE data
pool		0.42 (elevated plots)	
C fraction of	$\mathbf{f}_{\mathrm{lit}}$	0.5	EucFACE data
overstorey leaflitter			
pool			
C fraction of	$f_{ins}$	0.5	Ref 48
aboveground insect			
pool			
C fraction of frass	$\mathrm{f}_{\mathrm{frass}}$	0.53	EucFACE data
production			
C fraction of microbial	$\mathbf{f}_{micr}$	0.534 (ambient plots)	EucFACE data
pool		0.493 (elevated plots)	

C fraction of	$f_{myco}$	0.534 (ambient plots)	Assumed the same as
mycorrhizal pool		0.493 (elevated plots)	$\mathbf{f}_{micr}$
C fraction of soil pool	$f_{soil}$	0.016 (ambient plots) 0.017 (elevated plots)	EucFACE data
C fraction of twigs, barks and seeds production	f <sub>other</sub>	0.5	Assumed



819 Extended Data Figure 1. The *Eucalyptus* free air carbon dioxide enrichment experiment

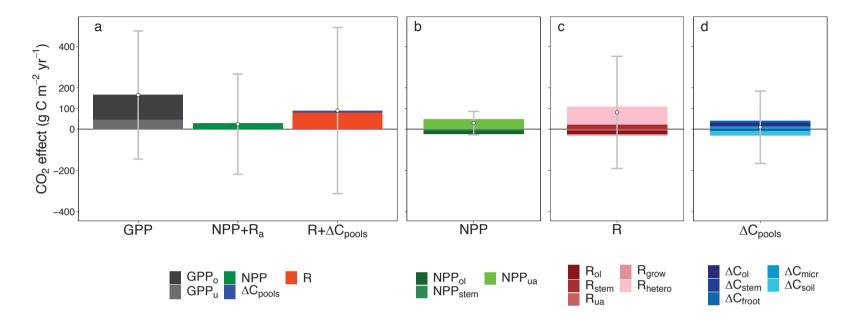
facility (EucFACE). a) A spatial overview of the forest and the facility (photo credit: David
S. Ellsworth), b) an overview of the understorey vegetation and infrastructure inside a plot
(photo credit: Mingkai Jiang), and c) a bottom-up look of the canopy structure and the crane
(photo credit: Mingkai Jiang).



826

827 Extended Data Figure 2. Estimates of (a and b) gross primary production (GPP) and (c and d) soil respiration (R<sub>soil</sub>) based on different methods for both (a and c) ambient and 828 829 (b and d) elevated CO<sub>2</sub> treatment at EucFACE. For estimates of GPP, we compared the 830 model simulated total GPP of overstorey and understorey (GPP<sub>o</sub> and GPP<sub>u</sub>, respectively), with the sum of data-driven estimates of net primary production (NPP) and autotrophic respiration 831 832 (R<sub>a</sub>), which include NPP of overstorey leaf (NPP<sub>ol</sub>), stem (NPP<sub>stem</sub>), fineroot (NPP<sub>froot</sub>), coarse 833 root (NPP<sub>croot</sub>), twigs, barks and seeds (NPP<sub>other</sub>), understorey aboveground (NPP<sub>ua</sub>), leaf consumption by insects (NPP<sub>ins</sub>), and respiratory fluxes of overstorey leaf (R<sub>ol</sub>), stem (R<sub>stem</sub>), 834 835 root (R<sub>root</sub>), understorey aboveground (R<sub>ua</sub>), growth (R<sub>grow</sub>), and volatile carbon emission (VC). For estimates of R<sub>soil</sub>, we compared direct estimates of R<sub>soil</sub> scaled up from soil chamber 836

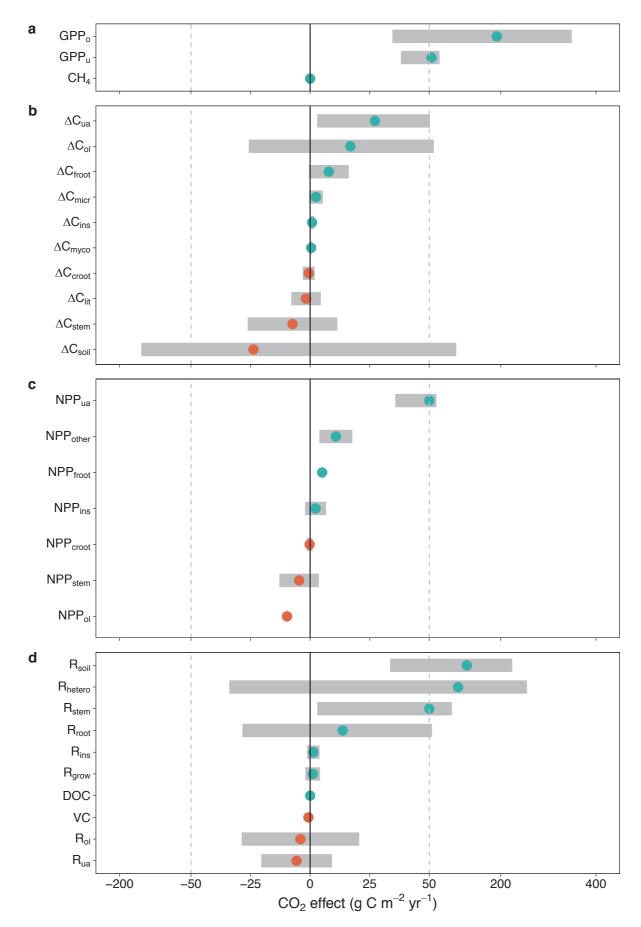
837 measurements, with the sum of litterfall and independent estimates of root respiration (Litter + R<sub>root</sub>), assuming no net change in soil carbon stock over time. Here litterfall was inferred based 838 839 on NPP of overstorey leaf (NPPol), fineroot (NPPfroot), coarse root (NPPcroot), twigs, barks and seeds (NPPother), understorey aboveground (NPPua), and frass production (Frass). These 840 evaluations provide independent mass balance checks of the estimated ecosystem carbon 841 budget. Each color represents a flux variable. Dotted point and vertical line represent treatment 842 843 mean and standard deviation based on plot-level estimates of the aggregated flux (n=3). Values were normalized by a linear mixed-model with pre-treatment leaf area index as a covariate to 844 845 account for pre-existing differences.



846

847 Extended Data Figure 3. The fate of additional carbon fixed under elevated CO<sub>2</sub> (eCO<sub>2</sub>) in a mature forest ecosystem (non-normalized analysis case). a) Column "GPP" represents the total  $eCO_2$  induced increase in overstorey and understorey gross primary production (GPP<sub>0</sub> and 848 GPP<sub>u</sub>, respectively), column "NPP +  $R_a$ " represents the sum of net primary production and autotrophic respiration eCO<sub>2</sub> response, and column "R 849 850 +  $\Delta C_{pools}$ " represents the sum of ecosystem respiration and carbon storage eCO<sub>2</sub> response. b) The relative contributions of individual NPP fluxes to the aggregated NPP response to eCO<sub>2</sub>, including overstorey leaf (NPP<sub>ol</sub>), stem (NPP<sub>stem</sub>), and understorey aboveground (NPP<sub>ua</sub>). c) The relative 851 852 contributions of individual respiratory fluxes to the aggregated R response to eCO<sub>2</sub>, including overstorey leaf (R<sub>ol</sub>), stem (R<sub>stem</sub>), understorey aboveground (R<sub>ua</sub>), growth (R<sub>grow</sub>), and heterotroph (R<sub>hetero</sub>). d) The relative contributions of individual change in carbon storage to the aggregated 853  $\Delta C_{\text{pools}}$  response to eCO<sub>2</sub>, including overstorey leaf ( $\Delta C_{\text{ol}}$ ), stem ( $\Delta C_{\text{stem}}$ ), fineroot ( $\Delta C_{\text{froot}}$ ), microbe ( $\Delta C_{\text{micr}}$ ), and soil ( $\Delta C_{\text{soil}}$ ). Variables with an 854

average CO<sub>2</sub> effect of  $< 5 \text{ gCm}^{-2}\text{yr}^{-1}$  were excluded from the figure for better visual clarification. Each color represents a flux variable, point indicates the net sum of all variables for a column, and the grey confidence interval represents plot-level standard deviation (n=3) of the estimated column sum.



Extended Data Figure 4. CO<sub>2</sub> treatment effect (gCm<sup>-2</sup>yr<sup>-1</sup>) for all ecosystem fluxes at 859 **EucFACE.** a) The CO<sub>2</sub> response of gross ecosystem carbon uptake, including gross primary 860 861 production of overstorey (GPP<sub>o</sub>) and understorey (GPP<sub>u</sub>), and soil methane uptake (CH<sub>4</sub>). **b**) The eCO<sub>2</sub> response of annual incremental change in carbon pool ( $\Delta C_{\text{pools}}$ ), including overstorey 862 863 leaf ( $\Delta C_{ol}$ ), stem ( $\Delta C_{stem}$ ), coarse root ( $\Delta C_{croot}$ ), fineroot ( $\Delta C_{froot}$ ), understorey aboveground 864  $(\Delta C_{ua})$ , leaf litter  $(\Delta C_{lit})$ , soil  $(\Delta C_{soil})$ , microbe  $(\Delta C_{micr})$ , aboveground insect  $(\Delta C_{ins})$ , and 865 mycorrhizae ( $\Delta C_{myco}$ ). c) The eCO<sub>2</sub> response of net primary production (NPP), including 866 overstorey leaf (NPP<sub>ol</sub>), stem (NPP<sub>stem</sub>), coarse root (NPP<sub>croot</sub>), fineroot (NPP<sub>froot</sub>), understorey 867 aboveground (NPP<sub>ua</sub>), twigs, barks and seeds (NPP<sub>other</sub>), and leaf insect consumption (NPP<sub>ins</sub>). d) The  $eCO_2$  response of ecosystem respiration (R) and other out-going flux, including 868 respiration fluxes of overstorey leaf (Rol), stem (Rstem), root (Rroot), understorey aboveground 869 (R<sub>ua</sub>), growth (R<sub>grow</sub>), insect (R<sub>ins</sub>), heterotroph (R<sub>hetero</sub>), and soil (R<sub>soil</sub>), and volatile carbon 870 emission (VC) and dissolved organic carbon leaching (DOC). Dots and grey bars represent 871 872 means and standard deviations of the CO<sub>2</sub> treatment difference, predicted by a linear mixedmodel with plot-specific pre-treatment leaf area index as a covariate. Orange dots indicate 873 negative means and light green dots indicate positive means. Dashed lines indicate change of 874 875 scale along the x-axis.