

THÈSE Transfrontalière UPPA / UPV

UNIVERSITÉ DE PAU ET DES PAYS DE L'ADOUR
ECOLE DOCTORALE DE SCIENCES EXACTES ET
DE LEURS APPLICATIONS (ED 211)

Elorri AREVALO

EN VUE DE L'OBTENTION DU DOCTORAT
DE L'UNIVERSITE DE PAU ET DES PAYS DE L'ADOUR
Spécialité : Physiologie & Biologie des Organismes – Populations – Interactions

Potential effects of Global Climate Change on survival and growth of brown trout (*Salmo trutta* L.): consequences of temperature and floods on young stages



Soutenu le : 05 décembre 2017

Devant la commission d'examen formée de :

Sandrine Plenet
Emili Garcia-Berthou
Véronique Gouraud
Franck Cattaneo
Miguel Clavero
Agnès Bardonnnet

Maître de conférence (HDR), Université de Lyon 1
Professeur, Université de Gerona
Ingénieur de recherche, EDF
Professeur, Hepia
Chargé de recherche, Station Biologique de Donana
Directeur de recherche, INRA

Rapporteure
Rapporteur
Examinatrice
Examineur
Examineur
Directrice de thèse

Remerciements

Je remercie en premier lieu mes encadrants de thèse, Agnès Bardonnnet et Aitor Larrañaga pour m'avoir offert l'opportunité de réaliser ce travail de thèse. Merci pour vos conseils, votre disponibilité, vos multiples corrections et surtout votre patience ... J'ai beaucoup appris pendant ces trois ans.

Je voudrais remercier Stéphane Pansérat, Margaret Lang, Steve Railsback, Etienne Prévost et Arturo Elozegi d'avoir participé à mes comités de thèse. Les discussions que j'ai pu avoir avec vous ont été extrêmement enrichissantes et ont grandement amélioré mon travail. Merci à Valérie Bolliet et Cédric Tentelier de m'avoir fait découvrir l'enseignement et de m'avoir épaulé tout au long de cette expérience. Je remercie également Iban Seiliez et Jacques Labonne pour leur aide et leurs conseils toujours avisés. Je remercie Jesus Pozo de m'avoir accueillie à l'université de Bilbao et m'avoir permis d'y passer quelques temps.

Je remercie chaleureusement toute l'équipe technique qui m'a grandement supportée (dans tous les sens du terme) pendant mes manips – et en dehors aussi ! Merci à Pascale Coste, Pantxo Guéraud, Stéphane Glise, Jacques Rives et Emmanuel Huchet. Nous avons toujours travaillé dans la bonne humeur et sans vous, pas grand-chose n'aurait été possible ! J'ai beaucoup appris à vos côtés. J'ai enrichi mon vocabulaire et mon panel de blagues, et ça c'est plutôt cool ! Merci à Elisabeth Plagnes-Juan de m'avoir guidé dans le monde de la biologie moléculaire. Merci beaucoup à Pantxika qui m'a aidée dans toutes les tâches administratives (qui sont tellement laborieuses ...). Merci à Philippe de m'avoir réconciliée avec les cours d'anglais. Je remercie tous les stagiaires qui ont également contribué à ce travail : Charlie, Léa, Mikel, Manon, Hélène, Charly et Amandine. Vous avez amené de la bonne humeur dans le labo. Je remercie Moustache de s'être sacrifié pour la bonne cause.

Je remercie encore toutes les personnes avec qui j'ai pu travailler mais également toutes les personnes de l'Aquapôle que j'ai pu côtoyer au quotidien. Merci pour votre bienveillance à mon égard, votre soutien, votre réconfort et vos sourires, en toutes circonstances.

Je remercie tous les jeunes que j'ai croisés à Saint-Pée : Zoé, Jordi, Vivi, Weiwei, Marion, Simon, Laury, Tengfei, Louis, Jingwei, Lorea et Charlotte. On a passé de bons moments tous ensemble et j'en garderai un bon souvenir. Je remercie particulièrement Séverine, Sarah, Colin et Thérèse. Vous avez été de superbes partenaires ! Je suis contente d'avoir fait cette thèse pour avoir pu vous rencontrer, vous avez illuminé mon quotidien. On a partagé nos galères mais on a aussi beaucoup rigolé. Sans vous, toute cette aventure n'aurait pas été la même,

heureusement que vous étiez là ... Vous avez amené tellement de joie dans mon cœur ... Vous êtes géniaux, ne changez pas, ça serait tellement dommage ! <3 Je remercie Maximus et Civ, j'espère que mes successeurs s'occuperont bien de vous !! Et bon courage aux nouveaux arrivants, Lucie, Louise et Clément. J'espère que vous allez perpétuer l'UMT.

Dédicace spéciale à Frédéric. Merci pour ton aide technique mais aussi pour la sortie en canoé (où j'ai cru que j'allais mourir), les chasses aux champignons et les parties de pêche. Merci d'avoir été là. Merci d'être ce que tu es.

J'ai passé trois super années avec vous tous. Le café de 8h, les mardis et vendredis soirs à la pala me manqueront. Ainsi que les barbecues, les mölkky sur la plage et tout le reste ... Merci à Ludo, Vincent V, Vincent G, Jean-Christophe, Mathieu, Aurélie, et toutes les personnes avec qui j'ai passé de bons moments (j'en oublie sûrement, je suis désolée !!). Merci à Sébastien de nous avoir permis de faire de Gantxiki notre QG !

Merci à Paolo Nutini, Pink Floyd, Avantasia, Monstercat (et bien d'autres) d'avoir animé mes journées. La musique, c'est la vie ! Merci à Anne-Marie, ma prof de piano.

Je remercie bien sûr ma famille qui a un peu (beaucoup, beaucoup) subi mon stress et ma mauvaise humeur pendant ces trois années ... Et pourtant, ils m'ont toujours soutenue. Merci à mes parents, mon frère, ma belle-sœur, mes neveux, mon Amatxi ... Et bien entendu, Loki !!!
Merci à ma môman de faire de si bonnes omelettes !!

Et enfin, merci à tous les gens moyens ! (#Piste 3)

Abstract

Global Climate Change will increase precipitations in the temperate and Northern coast of Europe during winter and spring. In riverine ecosystems, precipitations affect strongly the discharge of running waters and, thus, it is predicted that streams will face more severe floods. Additionally, air and water temperature will increase all over the world. These new environmental conditions can alter the phenology of species and predator/prey interactions. Newborns of brown trout (*Salmo trutta* L.) start their exogenous feeding in March/April. This stage is a critical step as individuals undergo huge physiological and behavioral changes. To allow a good development and a high survival rate, preys have to be abundant, particularly during early ontogenesis when fish are most vulnerable to food scarcity and predation. In this thesis, experiments in controlled-environment were conducted to estimate the effect of water velocity on the drift of preferred prey taxa for salmonids and to understand the effect of temperature on the metabolism of alevins facing starvation. Experiments in semi-natural conditions were set up to better understand the effects of floods on invertebrate communities and on survival, behavior and growth of first-feeding alevins. Our data support that floods affect trout differently depending on when they start feeding (early or late spring) and the availability of prey in their environment.

Résumé

Le changement climatique devrait induire une augmentation des précipitations pendant l'hiver et le printemps dans les régions tempérées et la côte nord de l'Europe. Dans les écosystèmes fluviaux, les précipitations affectent fortement le débit des eaux courantes et les rivières subiront des crues plus sévères. En outre, la température de l'air et de l'eau augmenteront à travers le monde. Ces nouvelles conditions environnementales vont avoir des conséquences sur la phénologie des espèces et les interactions prédateurs/proies. Les jeunes truites fario (*Salmo trutta* L.) commencent leur alimentation exogène en mars/avril. Cette étape critique de leur cycle de vie induit d'importants changements aussi bien physiologiques que comportementaux. Pour permettre un bon développement des individus et un taux de survie élevé, les proies doivent être disponibles et abondantes, en particulier à ce moment de l'ontogénèse où les juvéniles sont vulnérables au manque de nourriture et à la prédation. Des expériences en milieux contrôlés ont été menées pour quantifier la sensibilité à la dérive de trois espèces d'invertébrés couramment consommées par les salmonidés en fonction de différentes modalités de vitesses de courant et pour évaluer l'effet de la température sur le métabolisme d'alevins en situation de jeûne. Des expériences en milieu semi-naturel ont été mises en place pour mieux comprendre les effets d'une crue sur la communauté d'invertébrés et sur la survie, le comportement et la croissance des alevins en première alimentation. Il apparaît que la crue impacte différemment les truites en fonction du moment de la saison auquel elles commencent à s'alimenter (au début ou à la fin du printemps) et de la productivité du système.

Resumen

Se espera que el cambio climático aumente las precipitaciones durante el invierno y la primavera en las regiones templadas y en la costa norte de Europa. En los ecosistemas fluviales, las precipitaciones influyen directamente al caudal y, por tanto, se espera que los ríos sufran inundaciones más severas. Además, la temperatura del aire y del agua aumentarán en todo el mundo. Estas nuevas condiciones ambientales afectarán a la fenología de las especies y a las interacciones depredador-presa. Los individuos jóvenes de la trucha común (*Salmo trutta* L.) comienzan su alimentación exógena en marzo/abril. Esta etapa crítica en su ciclo de vida conlleva importantes cambios fisiológicos y de comportamiento. Con el fin de permitir un buen desarrollo de los individuos y una alta tasa de supervivencia las presas deben ser abundantes, especialmente durante las primeras etapas, que son más vulnerables a la escasez de alimentos y la depredación. En esta tesis se llevaron a cabo experimentos controlados para cuantificar la sensibilidad de tres especies de invertebrados comúnmente consumidos por los salmónidos a la velocidad del agua, y, para evaluar el efecto de la temperatura sobre el metabolismo de los alevines en ayuno. Además, con otros experimentos en ambientes semi-naturales se ha tratado de comprender mejor los efectos de las inundaciones en las comunidades de invertebrados y en la supervivencia, comportamiento y crecimiento de los alevines en el comienzo de la alimentación exógena. Parece que las inundaciones afectan a la trucha de forma diferente dependiendo de cuándo llegan a esta fase (a principios o finales de la primavera) y la disponibilidad de presas en su ambiente.

Laburpena

Klima-aldaketaren aurreikuspenen arabera eskualde epeletan eta Europako iparraldeko kostaldean prezipitazioak handitu egingo dira neguan eta udaberrian. Prezipitazioek zuzenean eragiten dute ibaien emarien igoera, eta hala, ibaiek uholde gogorragoak jasango dituztela aurreikusten da. Horrez gain, airearen eta uraren tenperaturak mundu osoan egingo du gora. Ingurumen baldintza berri hauek espezieen fenologia eta harrapari/harrapakinen arteko elkarrekintzetan aldaketak sortuko dituzte. Amuarrain arruntak (*Salmo trutta* L.) martxo/apirilean hasten dira elikadura exogenoa erakusten. Bizitza-zikloaren etapa kritiko honek aldaketa garrantzitsuak dakartza fisiologian eta jokabidean. Aleen garapen egokia eta biziraupen-tasa handiak lortzeko, harrapakinak ugaria izan behar du, ontogeniaren lehen urratsetan batik bat, errekurso eskasia eta predazioaren aurrean zaurgarrienak diren momentua bait da. Tesi honetan salmonidoek kontsumitzen dituzten hiru makroornogabe espezieek ur abiaduradi dioten sentsibilitatea estimatu zen. Bestalde, beste experimentu batean baraualdian zeuden alebinetan tenperaturak metabolismoan zuen eragina neurtu zen. Gainera, ornogabe komunitateetan eta elikatze exogenoan hasi berriak ziren alebiren biziraupenean, portaeran eta hazkundean uholdeek zuten eragina estimatu zen baldintza semi-naturaletan gauzatu ziren experimentuetan. Gure datuen arabera uholdeek eragin ezberdina dute amuarrainetan elikatzen hasten diren garaiaren arabera (udaberri hasieran edo bukaeran) eta inguruneak eskaintzen dien harapakin ugartasunaren arabera.

State of the art	5
General context	6
Growing in running waters.....	6
The Match-Mismatch Hypothesis	7
Global Climate Change threatening trophic links	8
Brown trout (<i>Salmo trutta</i> L.) as a case of study	9
Repartition, ecological requirements and life cycle	9
Emergence as a crucial step.....	10
Diet of <i>Salmo trutta</i>	12
High discharge and the biota.....	14
Synthesis and main objectives of this work	15

Chapter I Drift of invertebrates: effects of taxa, water velocity, gravel bed quality and body size17

Study context.....	18
Comparison of the propensity to drift for three invertebrate taxa: a laboratory study.....	19
ABSTRACT	19
Introduction	19
Material and Methods.....	21
Results	25
Discussion	28
Acknowledgements	31
Supporting Information	32
Further considerations and discussion.....	35

Chapter II Effect of feeding conditions and temperature on growth and metabolism of alevins.....39

Study context.....	40
Effect of food shortage and temperature on 0 ⁺ salmonids: a contribution to predict the effects of Climate Change.....	42
ABSTRACT	42
Introduction	42
Material and methods	43
Results	45
Discussion	50
Acknowledgements	52
Supporting Information	53
Further considerations and discussion.....	62

Chapter III Consequences of Global Climate Change on the community of invertebrates, on the survival, growth and behaviour of first-feeding alevins at high density: flood simulation in a semi-natural environment.....71

Study context.....	72
Assessment of Global Climate Change on the community of invertebrates, on the survival, growth and behaviour of first-feeding alevins: flood simulation in a semi-natural environment.....	72
Introduction	72
Material and Methods.....	74
Experimental design.....	74
Fish processing	75
Benthic invertebrate sampling.....	76

Statistical analyses.....	78
Results	83
Effect of the flood on the community of invertebrates	83
Effect of the flood on alevins	86
Behaviour of alevins.....	90
Discussion	97
Artificial flood and invertebrate abundances	97
Artificial flood and performances of alevins.....	97
Artificial flood and behaviour of alevins	98
Artificial flood and season	99
Conclusion.....	99
Supporting Information of the manuscript in preparation.....	100

Chapter IV Consequences of Global Climate Change on the community of invertebrates, on the survival and growth of first-feeding alevins at low density: flood simulation in a semi-natural environment.....107

Introduction	108
Material and Methods.....	108
The experimental channel of the Lapitxuri and experimental design	108
Fish sampling	109
Benthic invertebrate sampling.....	110
Statistical analyses.....	110
Results	113
Effect of the flood on the community of invertebrates	113
Effect of the flood on alevins	116

Relationship between the community of invertebrate and alevins.....	117
Gut contents.....	121
Discussion	123
Conclusion.....	125
General discussion	127
References.....	138
Article references of Chapter 1	158
Article references of Chapter 2	163
Article references of Chapter 3	166

State of the art

General context

Growing in running waters

Streams are classified according to channel geomorphology (size, width, depth) and volume of water discharged (Horton 1945; Strahler 1954, 1957). The smallest streams, namely first order streams, are located at the steepest parts of the watershed. The union of two first-order streams results in a second-order stream, and so on to sea; a sequence that entails important changes in functioning. Flowing waters have four distinct sources of energy (Allan & Castillo 2007; Giller & Malmqvist 1998). The first source comes from plants that use solar radiation *via* photosynthesis to produce instream (autochthonous) primary production. The other three sources of energy are imported into the running water system from the surrounding valley (allochthonous): coarse particulate organic matter (CPOM – $\phi > 1$ mm), fine particulate organic matter (FPOM – $0.5 \mu\text{m} < \phi < 1$ mm) and dissolved organic matter (DOM – $\phi < 0.5 \mu\text{m}$) coming from the riparian vegetation or banks in the river. Then, autochthonous and allochthonous energy, in the form of organic matter, is transferred to consumers. The contribution of each source of energy varies along the river sections and changes the invertebrate community structure longitudinally, in a sequence that is known as the “River Continuum Concept” (RCC – Vannote et al. 1980). In headwater low order streams, most of the time dense canopies reduce the intensity of light and thus the amount of primary production. Then energy mainly comes from leaf inputs and invertebrate communities are principally composed by detritivores, mainly shredders. In mid-order streams, light conditions are improved and favour algal and macrophyte growth, stimulating the presence of grazers and scrapers. Finally, the scarce benthic light under the turbid waters of high order streams limit primary production again, and so, CPOM coming from the upstream parts are degraded in FPOM and consumed by the very abundant collectors. Then, instream energy availability is closely related to the light availability and to the transport of materials, which regulate primary (instream or in riversides) and secondary production (*i.e.* biomass production of both invertebrates and fish).

The perpetuation of species involves the individuals to grow, complete their developmental cycle and reproduce. They choose their habitat to maximize access to food, shelter or partners. Throughout the development their needs change and they may be forced to move from one habitat to another to optimise fitness. These habitat changes over the life of an organism are called “ontogenetic shifts” (Werner & Gilliam 1984). In nature, growth and survival of fish depend on several factors. As they are ectotherms, water temperature governs many physiological processes such as respiration, excretion and growth. The seasonality, the

abundance and the quality of food, within the constraints of the thermal regime, cause significant variability in fish production over time and space (Benke et al. 1988; Huryn 1996; Thompson & Beauchamp 2016) and appear as one of the main drivers that limit fish growth (Wildhaber & Lamberson 2004). As they grow, mortality risks are reduced. The number of potential predators that can feed on fish diminishes (Quinn & Peterson 1996; Sogard 1997), and they also become better competitors and swimmers (Young 2003), which guarantee their access to food (Ryer & Olla 1996), shelter (Harwood et al. 2002; Orpwood et al. 2003). However, individuals have to face events, which can slow down their growth and threaten their survival.

The Match-Mismatch Hypothesis

During the developmental cycle of species, some stages are more vulnerable to food scarcity. For example, the breeding period of the Atlantic puffin (*Fratercula arctica*) is synchronized with the peak of herring abundance, which are their main food items (Durant et al. 2003, 2005). The synchronization between prey availability and predator requirements increase the survival rate of chicks, while a mismatch would produce a weaker cohort and population. Hjort (1914) firstly suggested that mortality of marine fishes (cod, haddock and Norwegian spring herring) was probably due to food scarcity during larval and young fry stages, which were the most critical periods in fish development. In addition, Cushing (1969) noticed that the spawning timing of marine fishes (herring, plaice and cod) allowed the hatching of the eggs and then, the development of larvae when the zooplankton production was high. Indeed, releasing larvae during the spring or autumn peaks in plankton production resulted in high survival rate for the three species of interest (Cushing 1990).

Following these observations on terrestrial and marine species, the “Match-Mismatch Hypothesis” (MMH) emerged and states that critical periods during which predators need energy occur simultaneously with the peak availability of prey. This way, recruitment of predators is maximised. On the contrary, the higher the mismatch between food requirements and food availability, the lower the growth, survival and recruitment of predators (Durant et al. 2007; Woodward et al. 2010; Bewick et al. 2016). Mortality induced by prey scarcity is expected to be higher for fish larvae than for late stages because (i) larvae are not able to withstand prolonged fasting periods and (ii) smaller larvae are more susceptible to predation (Dou et al. 2005; Yokota et al. 2016). The mismatch between predator requirements and food availability can be due to the occurrence of disturbance, but, as far as we know, the MMH has never been tested in freshwater systems.

Global Climate Change threatening trophic links

In Europe, projections for 2100 from the Intergovernmental Panel on Climate Change (IPCC) and from several studies modelling the consequences of Global Climate Change (GCC) on the precipitation patterns, forecast an increase in the magnitude and frequency of extreme hydrological events (Lehner et al. 2006; Dankers & Feyen 2008; IPCC 2014) as well as a modification of events timing (Blöschl et al. 2017). In southern Europe, average seasonal precipitations is predicted to increase from 5 to 40% during winter (from December to February – Figure 1), whereas the decrease of precipitation will induce low flows the rest of the year. In central and northern Europe, precipitations are expected to drastically increase during winter but also in spring (from December to May). It can also be noted that the mean discharge of rivers are predicted to increase outside periods of extreme flood. On the contrary, the summer season (from June to August) will be more arid and droughts will be more extreme almost everywhere in Europe. Additionally, climate scenarios forecast an air temperature increase of 3°C, which will induce an elevation of the river water temperature between 0.8 and 1.6°C (van Vliet et al. 2013; Bal et al. 2014).

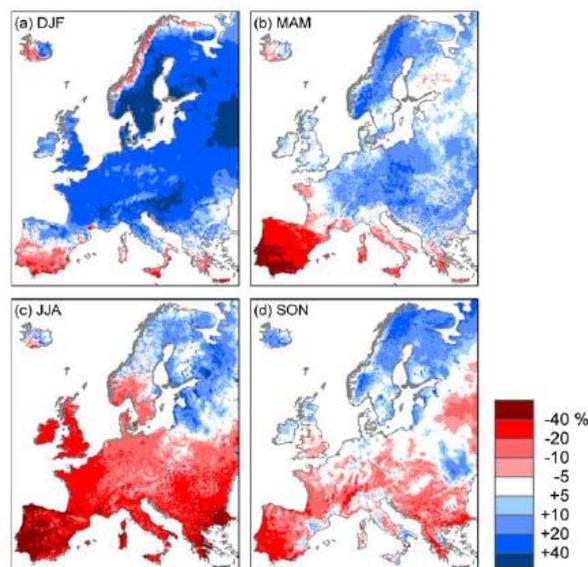


Figure 1 Predictions from Dankers & Feyen (2008) about the consequences of Global Climate Change in the average seasonal precipitation in (a) winter, (b) spring, (c) summer and (d) autumn in Europe.

In rivers, the increase of precipitations will increase the occurrence and the intensity of winter and spring floods. These new flow and temperature patterns can threaten the dynamic equilibrium of species. In particular, links between consumers and resources, which are often the result of a long co-evolution, might be weakened (Woodward 2009; Perkins et al. 2010;

Woodward et al. 2010). Consequently, the increase in stochastic events due to GCC may threaten the synchronization of species phenology and peaks in prey abundance may be shifted (advanced or delayed). Predators may then face higher mortality rates due to starvation during critical periods.

Brown trout (*Salmo trutta* L.) as a case of study

Repartition, ecological requirements and life cycle

Brown trout is indigenous to Europe, North Africa and western Asia (Klemetsen et al. 2003). It is present in many regions of Europe from north of Iceland, Scandinavia and Russia to South of the Mediterranean Sea. After many introductions, brown trout has now reached a world-wide distribution and is present in various biogeographic contexts (Elliott 1994). It is an important economical resource for professional and recreational fishing and it is frequently used as tourist attraction (Aas et al. 2000; Butler et al. 2009).

At the river scale, brown trout prefer upstream sections of rivers (Huet 1949, 1954), with well-oxygenated waters and neutral pH. Flow plays a major role in the distribution of individuals, supply of drifting invertebrates, redd oxygenation (Baglinière & Maisse 1991). They choose complex substrates that provide shelters from water velocity, predators and visual isolation between congeners (Armstrong et al. 2003). Lastly, brown trout is adapted to cool and quite narrow water temperature conditions (Elliott 1994).

Reproduction takes place from November to December, females dig a nest in the gravel, drop their eggs immediately fertilised by males and they cover the clutch with sediment. Eggs grow under the protection of gravel and they hatch around February, at 420 degree-days (*i.e.* the sum of the mean daily water temperature from the egg-laying). New-born alevins stay under the gravel, they feed on their yolk reserves. From March to April, when their yolk sac is almost exhausted (around 730 degree-days) alevins leave the protection of the redd and emerge in the water column to search for prey (Roussel & Bardonnnet 2002). After spending at least one year in the river, juveniles can spend all their life in running waters (“riverine trout”), migrate to lake (“lake trout”) or migrate to sea after physiological adaptations (smoltification; “sea trout” – Figure 2).

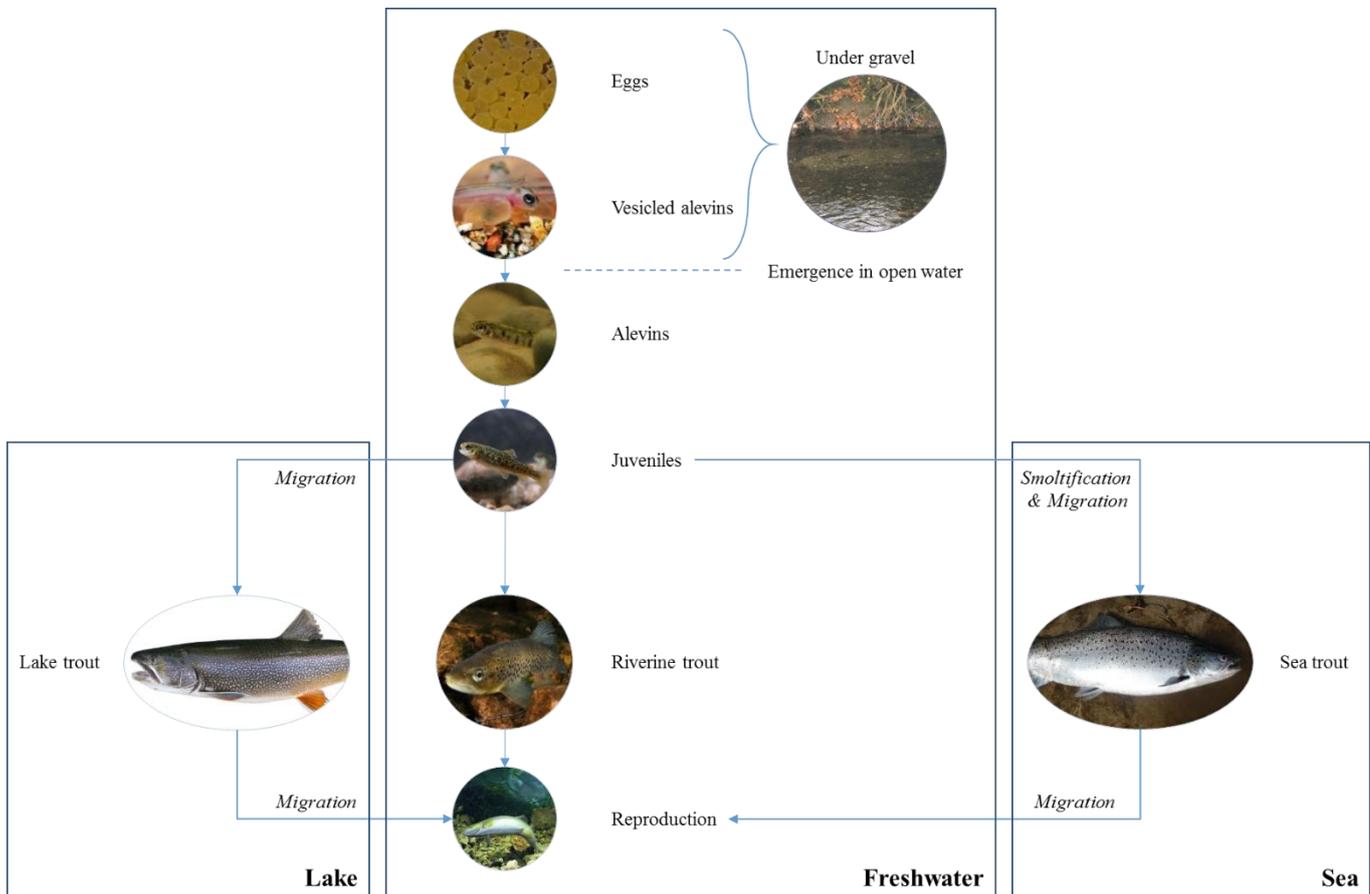


Figure 2 Life cycle of brown trout (*Salmo trutta*) according to life stages and to the different environments that the species is capable to use for growing (lake, freshwater and sea).

Emergence as a crucial step

The move from the under-gravel compartment to water column is called “emergence” and it is considered as a critical time for survival in brown trout with mortality rates as high as 90% (Elliott 1989). Alevins undergo huge behavioural and physiological changes (Einum & Fleming 2000; Skoglund & Barlaup 2006; Kennedy et al. 2008) and they must adapt quickly to a radically different environment. When they emerge, their yolk sac is almost exhausted and they shift to an exogenous feeding, which implies metabolic changes (Mennigen et al. 2013) and the establishment of the processes of digestion, absorption and assimilation (Dabrowski 1984). In addition to physiological changes, emergence involves behavioural modifications. Alevins need to maintain their position in the water column and catch prey that drift in water. Salmonids are territorial animals and a social hierarchy is established the week following emergence (Héland 1999). Alevins compete with congeners (conspecifics from the same clutch and those from surrounding redds) to access to the best hunting spots. They are “sit-and-wait” hunters (Elliott 1967; Cada et al. 1987; Giroux et al. 2000): they defend small territories (0.1–0.2 m² – Grant et al. 1998) close to shelters and near fast-flowing water that provide high quantity of prey. This strategy allows alevins to minimize their energetic expenditures while energy input is maximized by a large supply of food in the drift. These stations are energetically favourable

(Fausch 1984) and are generally chosen by dominant individuals (Jenkins 1969; Fausch 1984; Grant & Kramer 1990). The intensity of the competition depends greatly on the density of alevins. According to Elliott (1989), density exerts a selective pressure during 20-30 days after emergence. Under high density, the scarcity of favourable habitat and/or feeding resource would diminish the growth and then survival of alevins (Figure 3 – Elliott 1989; Milner et al. 2003).

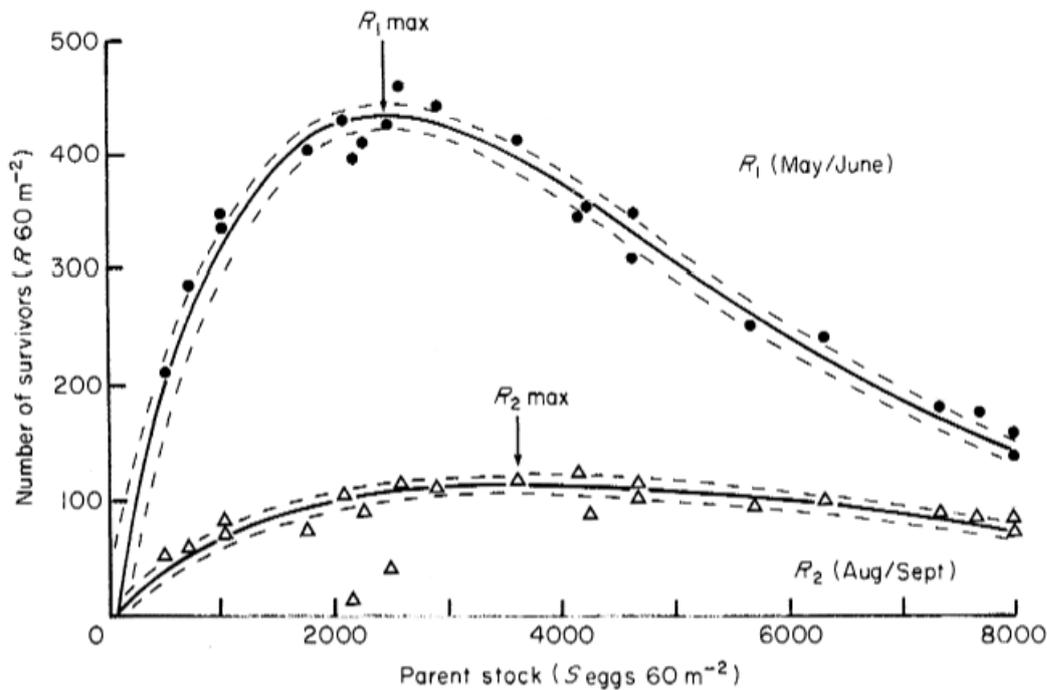


Figure 3 Relationship between parent stock (S eggs per 60 m^2) and number of survivors (R fish per 60 m^2) in May/June (black circles), August/September (empty triangles) – modified from Elliott 1989.

The timing of emergence influences density of alevins and depends on temperature, date of adult spawning and quantity of energy allocated by females to eggs (Armstrong & Nislow 2006; Régnier et al. 2013). Early emergence give access to the best hunting spots but fish are more vulnerable to predation and to hydrological events such as large floods (Einum & Fleming 2000). Late emerging alevins mingle with early emerging survivors and alevin densities may be high. This can decrease the predation risk of alevins by piscivore fish due to a dilution effect (Sogard 1997; Alvarez & Nicieza 2003) but intensify competition (Skoglund et al. 2011). Moreover, prey availability increases throughout spring, which should tend to favour alevins emerging later. First moments of salmonid life are crucial for growth and affect directly life history traits, strength of the cohort and population dynamics (Bacon et al. 2005; Wysujack et

al. 2009; Jonsson & Jonsson 2014). Then, understanding key factors acting on growth and survival of alevins is of main importance to improve our understanding on brown trout population functioning (Elliott 1986).

*Diet of *Salmo trutta**

In open waters, brown trout (alevins and juveniles) feed mainly on “macroinvertebrates”, which are organisms living in/on the top layer of river bed (Tachet et al. 2010) and larger than 0.5 mm at their adult stage. Macroinvertebrates are of main importance in ecosystem processes: they contribute to the processing of allochthonous and autochthonous organic carbon, influence periphyton growth and represent the main feeding resource for many fish and birds living near rivers (Quinn & Hickey 1990; Wallace & Webster 1996; Malmqvist 2002). Stream macroinvertebrates spend their larval stage in freshwaters, while the adult stage takes place in terrestrial systems after a nymphal transformation but some species (such as Mollusca, Crustacea, Oligochaeta and some Coleoptera) spend their entire life cycle in rivers. Macroinvertebrates colonize new habitats by drifting from upstream sources, crawling/swimming from adjacent substrates (including the hyporheic habitat) or adult flying (Mackay 1992). Their instream distribution and abundance depend on a variety environmental factors (Townsend et al. 2003). At the watershed scale, altitude, local climate, topography, geology and catchment vegetation drive the assemblage of invertebrates (Winterbourn 1981; Li et al. 2012). At the river scale, flow directly affects aquatic communities by influencing water quality, food sources, species interactions and the availability of diverse/heterogeneous habitats (Jowett & Richardson 1990; Quinn & Hickey 1990). Water physico-chemistry (temperature, pH, dissolved oxygen concentration) and nutrient availability can also heavily impact the survival, reproduction and growth of invertebrates (Stewart et al. 2000; Rawi et al. 2014).

Brown trout feed mainly on “drifting” aquatic larvae of invertebrates, which are carried by the current. However, brown trout exhibit flexible behaviour (Allen 1951; Ringler 1985; Waters 1988) and can adapt their hunting mode to maximize their energetic gain (Optimal Foraging Theory – OFT; Pyke et al. 1977). They can search prey in the benthos (Ware 1972) or they can pick up terrestrial invertebrates at the water surface (Dahl & Greenberg 1996; Huryn 1996; Nakano et al. 1999). The main factor driving trout prey selection is prey size. Salmonids are gap-limited predators (Sánchez-Hernández et al. 2011) and their access to food increases as the size of their mouth and oesophagus increases. Gut content analyses of newly emerged alevins revealed that prey were selected up to 0.50 mm width even if prey up to 2 mm width could be consumed (Bozek et al. 1994; Domagała et al. 2014) and between 2 and 5 mm length

(Wankowski 1989; Hubert & Rhodes 1992; Keeley & Grant 2001). Another important factor in prey selection is the morphological and behavioural features of invertebrates (Poff et al. 2006). According to these traits, the probability to be captured by a fish differs among taxa. Rader (1997) broadly reviewed traits affecting the availability of invertebrates for drift-feeding fish and all traits related to the propensity of invertebrates to drift are of main importance. Invertebrates intentionally drifting or those easily dislodged by the water current are more likely to be preyed. Moreover, flow exposure depends on the use of the habitat and shelters (Negishi & Richardson 2006; Fuller et al. 2010): invertebrates inhabiting the hyporheic zone are less susceptible to be carried away by the current than invertebrates living at the surface of stones (Ware 1972). The mobility (*i.e.* sessile, attached, crawling or swimming) and the shape (*i.e.* streamlined, cylindrical, dorsoventrally-flattened or spherical) of invertebrates also influence their probability to be washed by the flow as the drag force exerted by the water on individuals depends on their length, width and height (Naman et al. 2016; Schülting et al. 2016). However, the critical factor remains the abundance of the taxon in the system. An abundant taxon, prone to drift, is highly available for fish (Crespin de Billy & Usseglio-Polatera 2002).

Considering the catchability, drift propensity, abundance, energetic profitability of prey and size limitation of salmonid predators, Rader (1997) ranked first the Baetidae family (Order: Ephemeroptera), second the Simuliidae family (Order: Diptera) and third the Chironomidae family (Order: Diptera). Next, the most available prey were from the Ephemeroptera order (*i.e.* Heptageniidae, Ephemerellidae, Leptophlebiidae and Siphonuridae), and Crustacea (Amphipoda). Many researchers confirmed these findings and showed that Baetidae, Simuliidae and Chironomidae larvae were the most important food items in gut contents of brown trout in different geographical areas (McCormack 1962; Elliott 1967; Fahy 1980; Vignes & Heland 1995; Sánchez-Hernández et al. 2011 – Figure 4) and provided over 80% of the energetic inputs of new-borns alevins (Sánchez-Hernández et al. 2012).



Figure 4 Invertebrates most available and consumed by drift-feeding fish, including brown trout alevins. From left to right: Baetidae, Simuliidae and Chironomidae families. Photo from: <http://lifeinfreshwater.net>.

High discharge and the biota

The increase of discharge rises the scouring force of water on the bottom, called shear stress, and displaces the organisms downstream, which in turn alters the composition of instream communities (Leigh et al. 2015). The tight link between the discharge regime and macroinvertebrates has been described by many researchers. The optimal range of water velocities for invertebrates varies between 0.1 m s^{-1} and 0.6 m s^{-1} . Flow velocities higher than 0.7 m s^{-1} are not considered suitable for most macroinvertebrates (Gore et al. 2001; Li et al. 2009; Horta et al. 2009; Shearer et al. 2015).

In rivers, “hydropeaking” refers to frequent discharges pulses generated by hydroelectric power generation (Bratrich et al. 2004; Bretschko & Moog 1990; Bruno et al. 2013) and several studies showed that hydropeaking causes an increase in the number of macroinvertebrate drifting and a reduction of macroinvertebrate biomass and abundance in the benthos (Moog 1993; Céréghino & Lavandier 1998; Céréghino et al. 2002; Bruno et al. 2013; Miller & Judson 2014). Similarly, natural extremely large floods can reduce invertebrate abundances between 15 and 90%, while invertebrate diversity is reduced by 70% or not at all affected (Melo et al. 2003; Argerich et al. 2004; Mesa 2010). McMullen & Lytle (2012) conducted a meta-analysis for the link between flood events and changes in invertebrate communities with 41 studies spread across the world. Despite the differences in river type and regional climate, all these studies concluded that the total abundance of invertebrates as well as the abundance of the major groups of invertebrates significantly decrease immediately after floods. Moderate flow events have also been responsible for the reduction of invertebrate abundance and diversity by 90% and 25%, respectively (Theodoropoulos et al. 2017).

In brown trout, critical water velocities beyond which fish cannot maintain their position are 0.25 m s^{-1} for recently emerged alevins (body length around 3 cm – Heggenes & Traaen 1988; Bardonnnet & Héland 1994) and 0.7 m s^{-1} for adults (around 21 cm – Heggenes 1988). High water velocities can alter recruitment (*i.e.* the juveniles incorporating the population) by

destroying redds, displacing individuals (Seegrist & Gard 1972; Wenger et al. 2011) or diminishing the abundance of suitable microhabitats for feeding and sheltering (Lobón-Cerviá & Mortensen 2005). Temporal variations in water velocity are known to limit survival and growth rates of the youngest juveniles (Lagarrigue et al. 2002; Korman et al. 2011). Lobón-Cerviá (2004; 2005) highlighted the importance of river discharge at the time of emergence and its influence on the population structure over the years. Indeed, year-to-year variation in river discharge at the time of emergence consistently matched recruitment rates, survival rates of mean cohort size and abundance of spawners. This phenomenon has been described for a variety of populations across the European range of brown trout (Elliott et al. 1997; Jensen & Johnsen 1999; Mäki-Petäys et al. 1999; Cattaneo et al. 2002, 2003). Similarly, numerous introduced populations across geographical (climatic) North American regions appear to be constrained by discharge variations (Strange et al. 1992; Nehring & Anderson 1993; Nuhfer et al. 1994; Latterell et al. 1998). Elwood & Waters (1969) suggested that declines in invertebrate populations caused by floods could reduce the food availability for fish causing an apparent decrease in their growth. However, it is difficult to disentangle the relative importance of these factors (*i.e.* diminution in habitat or trophic availability) in the mortality of alevins.

Synthesis and main objectives of this work

In brown trout, alevins emerge in spring, which currently matches with the increase in abundance of invertebrates in the drift (O'Hop & Wallace 1983; Romaniszyn et al. 2007; Leung et al. 2009) after the usually low abundances in temperate areas in winter (Brittain & Eikeland 1988). Besides, literature results seem to underscore a negative effect of hydrological events on both fish survival and growth. Because growth is highly density-dependant in salmonids (Jenkins et al. 1999; Vøllestad et al. 2002) one can suppose that the mechanism acting on 0^+ cohort following spates is related to low food availability (the Match-Mismatch Hypothesis), and/or suitable habitat availability. In this thesis, we carried out experiments which should help assessing to what extent Global Climate Change and more specifically the awaited increase in both spring temperature and precipitations might affect the post-emergence survival and growth of trout in relation to food shortage.

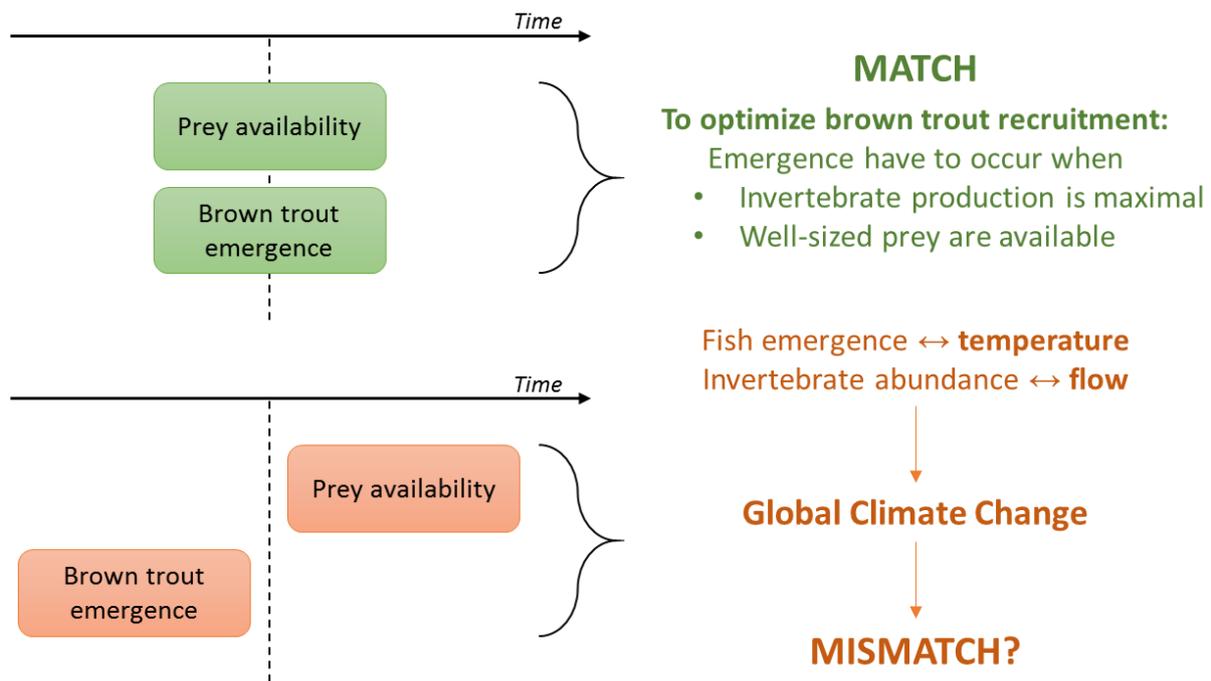


Figure 5 Invertebrates & brown trout: a Match-Mismatch Hypothesis case? Actually, prey availability matches with brown trout emergence (in green) but flow and temperature conditions should be modified by Global Climate Change and delayed prey availability inducing a decrease in brown trout recruitments (in orange).

This manuscript is composed of four chapters. The first chapter, based on an experimental study, investigates the drift propensity of three invertebrate taxa (*Baetis*, *Simulium* and *Chironomus*), which form the bulk of the diet of young salmonids. The second chapter, also conducted in a controlled environment, examines the metabolic pathways of just-emerged alevins facing starvation or a delayed first-feeding at 8 and 11°C. The third chapter relies on an experiment conducted in a semi-natural environment and tries to shed light on the consequences of an artificial flood on the invertebrates and on the survival, growth and behaviour of young trout at high density. The fourth chapter describes the consequences of a simulated flood on the invertebrate community and on the survival, growth and diet of trout alevins at low density. The last part of this manuscript draws a general discussion and proposes some hypotheses and future research pathways about the effect of Global Climate Change on young stages of salmonids.

Chapter I Drift of invertebrates: effects of taxa, water velocity, gravel bed quality and body size

Study context

In research areas focusing on drift-feeding fish, their growth is predicted from foraging and bioenergetic models. The Net Rate of Energy Intake of fish (NREI) is usually based on temperature and food availability (Hayes et al. 2000; Laliberte et al. 2016). Food availability is directly related to the density of drifting invertebrates and to water velocity. The increase in water velocity increases the invertebrate probability of being dislodged from the benthos and then, the number of drifting invertebrates (Brooker & Hemsworth 1978). However, Hayes et al. (2007) and Leung et al. (2009) emphasized that no studies provided an accurate estimation of the entry rate of invertebrates in the drift, which conditions the pertinence of the models. Moreover, Elliott (1971) investigated the distance travelled by invertebrates releasing them in running waters and he highlighted that an increase in water velocity carried the invertebrates further. Although this information is valuable for improving fish bioenergetic models, very few drift studies have attempted to predict invertebrate entry rate. To better understand the effect of floods on the food availability of salmonids, it is necessary to begin by understanding the response of invertebrates and their propensity to drift when facing water at various velocities.

Furthermore, the nature of the substrate is of prime importance. Stability of the gravel bed refers to its resistance to displacement and is generally proportional to particle size (Giller & Malmqvist 1998). Then, large particles increase the stability of the gravel bed, provide safe microenvironments and diminish accidental drift due to the dislodgment of invertebrates (Cobb et al. 1992). Number of shelters available for invertebrates depends on the number of interstices in the gravel bed, their size and the size of invertebrates (St Pierre & Kovalenko 2014).

In this experiment, we aimed to evaluate the drift propensity of three invertebrate taxa chosen because of their significant contribution to the diet of salmonid alevins: *Baetis* sp., *Simulium* sp. and *Chironomus* sp.. The variation of the drifting propensity was estimated in six indoor channels according to two environmental factors (water velocity and gravel bed quality) and a biotic factor (size of individuals).

Comparison of the propensity to drift for three invertebrate taxa: a laboratory study

Elorri Arevalo¹, Aitor Larranaga², Margaret Lang³, Etienne Prevost¹, Agnès Bardonnnet¹

¹*ECOBIOIP, UMR 1224, INRA, Univ Pau & Pays Adour, 64310, Saint-Pée-sur-Nivelle, France*

²*Lab of Stream Ecology, Dept. of Plant Biology and Ecology, Univ. Basque Country, UPV/EHU, 48080 Bilbao, Spain*

³*Environmental Resources Engineering, Humboldt State University, Arcata, CA 95521, USA*

Currently submitted to Hydrobiologia

ABSTRACT

Invertebrate drift is an important ecological process but factors affecting the downstream displacement of invertebrates are difficult to appraise. The influence of water velocity on drift entry rates is still unclear and has rarely been quantified. In this study, we investigated the drift propensity of *Baetis*, *Simulium* and *Chironomus*. In experimental channels, invertebrates were submitted to low, moderate and high water velocities (14 cm s⁻¹, 30 cm s⁻¹ and 40 cm s⁻¹) in either fine or coarse gravel beds. The drift was monitored for 24h to evaluate the effects of water velocity, gravel bed quality and invertebrate size on drift. Gravel bed quality had no effect. Small *Simulium* and *Chironomus* drifted more than larger ones, and drift was only positively related to water velocity for *Chironomus*. Accordingly, we presume that *Baetis* and *Simulium* were not constrained by the experimental conditions and drifted actively while *Chironomus* drifted passively.

Key words: invertebrates; modelling; water velocity; gravel bed quality; head capsule width; season.

Introduction

Invertebrate drift is defined as the downstream displacement of organisms and it is an important ecological process in lotic systems (Waters, 1965; Brittain & Eikeland, 1988). Drift contributes to the dispersal of invertebrate species across the water network, allows connection between habitats and facilitates recolonization processes. James et al. (2008) differentiate two drift categories: passive drift with animals unintentionally entering the water column and active drift with animals intentionally entering the water column.

Drift rate highly depends on abiotic (*i.e.* water chemistry – Hay et al., 2008, quantity of food – Ciborowski, 1983) and biotic (*i.e.* density of organisms – Lehmkuhl & Anderson, 1972; Walton et al., 1977, predation avoidance – Skinner, 1985; Sagar & Glova, 1992) factors and to determine the effect of each is extremely challenging. In addition, taxa respond differently according to their morphology, behaviour and ecology (Poff & Ward, 1991).

Many studies have attempted to investigate the effect of water velocity on invertebrate drift rate. The force exerted by the water on the river bed is called “shear stress” (Giller & Malmqvist, 1998; Vericat et al., 2008) and erodes mineral and biotic particles. The increase in water velocity within the channel increases the shear stress which then increases the risk of dislodgment and the passive transport of organisms downstream. Gibbins et al. (2007) observed the active drift of invertebrates up to a shear stress of 9 dynes cm⁻² (1 dyne cm⁻² = 1 Newton m⁻²) and above this threshold, drift was mainly passive due to the displacement of the gravel. All sheltered invertebrates are inevitably carried away by the current when the gravel bed is scoured. High shear stress mobilises substrate and fine particles and results in a patchwork of both scoured and infilled stream beds (Carling, 1987; Lake, 2000; Matthaei & Townsend, 2000).

High water velocities recorded during extreme flow events increase passive drift and the number of drifting invertebrates (Bruno et al., 2016; Radford & Hartland-Rowe, 1971), with reductions in the abundance of benthic invertebrates ranging from 14 to 95% depending on the magnitude of the event (Matthaei et al., 1997; Nislow et al., 2002; Robinson et al., 2004). However, Theodoropoulos et al. (2017) highlighted the lack of studies that examined the response of freshwater communities to medium water velocities (*i.e.* outside flood periods) and Hayes et al. (2007) go further by pointing out that the entry rate of invertebrates in the drift has yet to be quantified.

Recently, Naman et al. (2016) investigated the response of invertebrates to flow disturbance (from 8 cm s⁻¹ to 28 cm s⁻¹) according to channel architecture (flat vs concave channels) and taxa. The drifting community was comprised of a caddisfly (*Micrasema* sp.) and several taxa of Chironomidae, which for the most part, drifted passively. For the other drifting taxa, drift was primarily active. The shear stress exerted by the water was equal to 5 dynes cm⁻² and not enough to dislodge invertebrates.

The quality of the gravel bed, especially the size of the particles, plays an important role by providing invertebrates with hydraulic refuges to avoid being dislodged. Results from field

studies failed to separate the effect of water velocity, depth and substratum quality on passive drift of invertebrates because to date, assessments have been done at the river scale (Lancaster, 1999, 2000). Most data stemming from studies that examined the role of specific habitat parameters on invertebrate drift were drawn from experiments under controlled conditions and showed a higher drift rate in small gravel compared to cobble for caddisflies, mayflies (Holomuzki & Biggs, 2003) and stoneflies (Long et al., 2011) when submitted to spates. According to these results, the size of the interstices, linked to invertebrate size, could play an important role in the availability of hydraulic refugia.

In the present work we focus on the drift of three invertebrate taxa that have different habitat preferences and locomotor behaviour: *Baetis* sp., *Simulium* sp. and *Chironomus* sp.. They are abundant in riverine ecosystems and they contribute significantly to the diet of drift-feeding fish (Sánchez-Hernández et al., 2011). In experimental channels we manipulated the size of the benthic interstices using different sized substrates. We also considered the size of invertebrates. The interstice size-body interaction has rarely been related to drift propensity at the intra-specific level. Thus, the present study aims to quantify the propensity to drift according to taxa, water velocity, gravel bed quality and invertebrate size. We specifically tested the following hypotheses: 1) free swimming *Baetis* should actively drift more than *Simulium* (attached to the substrate with a suction cup) and *Chironomus* (buried under the substrate), 2) an increase in the water velocity should induce passive drift and increase the drift probability of invertebrates, 3) gravel size should mitigate passive drift with large particles diminishing drift probability of invertebrates, 4) large individuals should passively drift more than small ones due to scarcer shelter opportunities.

Material and Methods

Invertebrate collection

We collected invertebrates from two tributaries of the Nivelles River near Saint-Pée-sur-Nivelles (43°21' N, 1°33' W), on the 9th and the 10th of March and on the 19th and the 20th of October 2015. Baetids (*Baetis* sp.) were sampled from the Lapitxuri Brook, a pristine headwater stream. Simuliids (*Simulium* sp.) were sampled downstream from the spillway of a lake, in a fast-flowing brook characterised by a high percentage of fine sediment. Chironomids (*Chironomus* sp.) were purchased from Grebil (Arry, France). After collection, all invertebrates were kept in containers with aerated water at a constant temperature (*ca.* 13 °C). Rocks with biofilm and conditioned leaves collected in the field were added to give them food and refuge until the start of the experiment, a few days later.

Experimental features

The experiments were conducted in 6 indoor channels (L: 150 cm, W: 10 cm and H: 12 cm) that were continuously fed in parallel by water pumped from a large outdoor reservoir (3000 m³) filled with Nivelles River water. The water from the reservoir passed through a sand filter in order to remove most invertebrates, though we still recovered some small cyclops, oligochaetes and chironomids during the experiment. At this point, the only risk of confusion between the “experimental invertebrates” and the “natural invertebrates” concerned chironomids, but the larger size and the red colour of the reared taxon (*Chironomus* sp.) made it easy to separate wild from experimental animals. After the sand filter, water was sent to a 25 m³ reservoir and pumped to a small reservoir (0.5 m³) that fed by gravity the six experimental channels. The water level in the small reservoir was kept constant through an overflow. Water temperature followed natural changes, since water came from the outdoor reservoir and was registered every 15 min using a temperature logger (mini logger IIT, Vemco). Mean water temperature was 12.2°C (± 0.37, SD) in March and 14.7°C (± 0.50) in October. Light was controlled to mimic natural day and night rhythms. The lit period was chosen to match the natural photoperiod, and 30 min of progressive change in light levels were programmed to produce a 30 min period of dusk at 19:00 in March and 19:30 in October, and dawn at 6:30 in March and 7:30 in October. This results in quite similar L:D durations (12:11 in March and 11.5:11.5 in October).

Two types of gravel beds were created: a “fine” gravel bed (2-4 mm: 33%; 4-8 mm: 34%; 8-16 mm: 33%) and a “coarse” gravel bed (2-4 mm: 25%; 8-16mm: 50%; 20-31.5mm: 25%). Channels were filled to a depth of 3 cm for one gravel type, resulting in 3 fine and 3 coarse channels. Velocities were measured with a mini-velocimeter laid directly on the substratum (Schiltknecht, MiniWater®20 – Table 1, measure depth: 11 mm above the substratum). Three different flow levels were applied to both fine and coarse gravel channels: “low flow” (0.2 l s⁻¹ ~ 14.3 cm s⁻¹), “medium flow” (4 times increase, 0.8 l s⁻¹ ~ 29.8 cm s⁻¹), “high flow” (8 times increase, 1.6 l s⁻¹ ~ 39.6 cm s⁻¹).

Experiments began at 9:00 and lasted for 27 hours. Between 9:00 and 10:00, 50 individuals from each taxon were added to each channel (no flow). A stepwise increase in non-scouring flow was carried out for one hour to avoid an abrupt and high amount of drift (Imbert & Perry, 2000). At 10:00, flow was opened at the low flow level for all channels, and remained at this level for two hours. From 12:00 to 13:00, the flow was increased every 15 min to mimic a

gradual increase in water velocity, up to the high flow in channels 1 and 2, and up to medium flow in channels 3 and 4 (Table 1). Channels 5 and 6 were kept at low flow.

Table 1 Water velocity (mean \pm SD) for the 6 channels at the two trials.

Flow	Gravel bed	Water velocity in March (cm s ⁻¹)	Water velocity in October (cm s ⁻¹)
High	Coarse	36.71 \pm 4.61	40.00 \pm 2.38
High	Fine	43.86 \pm 4.60	37.86 \pm 2.27
Medium	Coarse	29.14 \pm 4.85	29.86 \pm 6.54
Medium	Fine	29.57 \pm 6.11	30.57 \pm 5.91
Low	Coarse	12.86 \pm 4.56	17.71 \pm 1.70
Low	Fine	14.29 \pm 1.80	12.43 \pm 2.23

From 10:00 onward, total hourly drift of invertebrates was gathered at the output of each channel in buckets equipped with two 10 x 10 cm openings closed by a 100 μ m mesh net. During the period of increasing flow (12:15, 12:30 and 12:45) and for each non-sampled twilight time-frame (19:30 and 6:30 in March, and 19:30 and 7:30 in October), additional samples were taken. The last sample was taken on the second day at 12:00. A total of 33 drift samples were collected per channel and, at the end of the experiment, all invertebrates still present in the channels were recovered manually from the substratum (Table S1). Water velocity was also measured at this time, just prior to the recovery of the animals, longitudinally along each channel at 20 cm intervals.

All animals were stored in 70% ethanol and photographed under a binocular microscope (Olympus SZX16). Both Head Capsule Widths (HCW) and Body Lengths (BL – from the top of the head to the end of the abdomen) were measured with the software ImageJ (Abràmoff et al., 2004).

Statistical analyses

We did not account for missing individuals (equivalent to 12% of the individuals, including those not in the drift and those not recovered at the end). From the 1589 individuals that were collected and measured, we discarded animals collected before the beginning of the flow increase, *i.e.* during the settling period. Thus, 1396 individuals were used in the analyses and we assumed that the propensity/decision to drift of an individual was independent from that of the others. Raw data are shown in Figure 1.

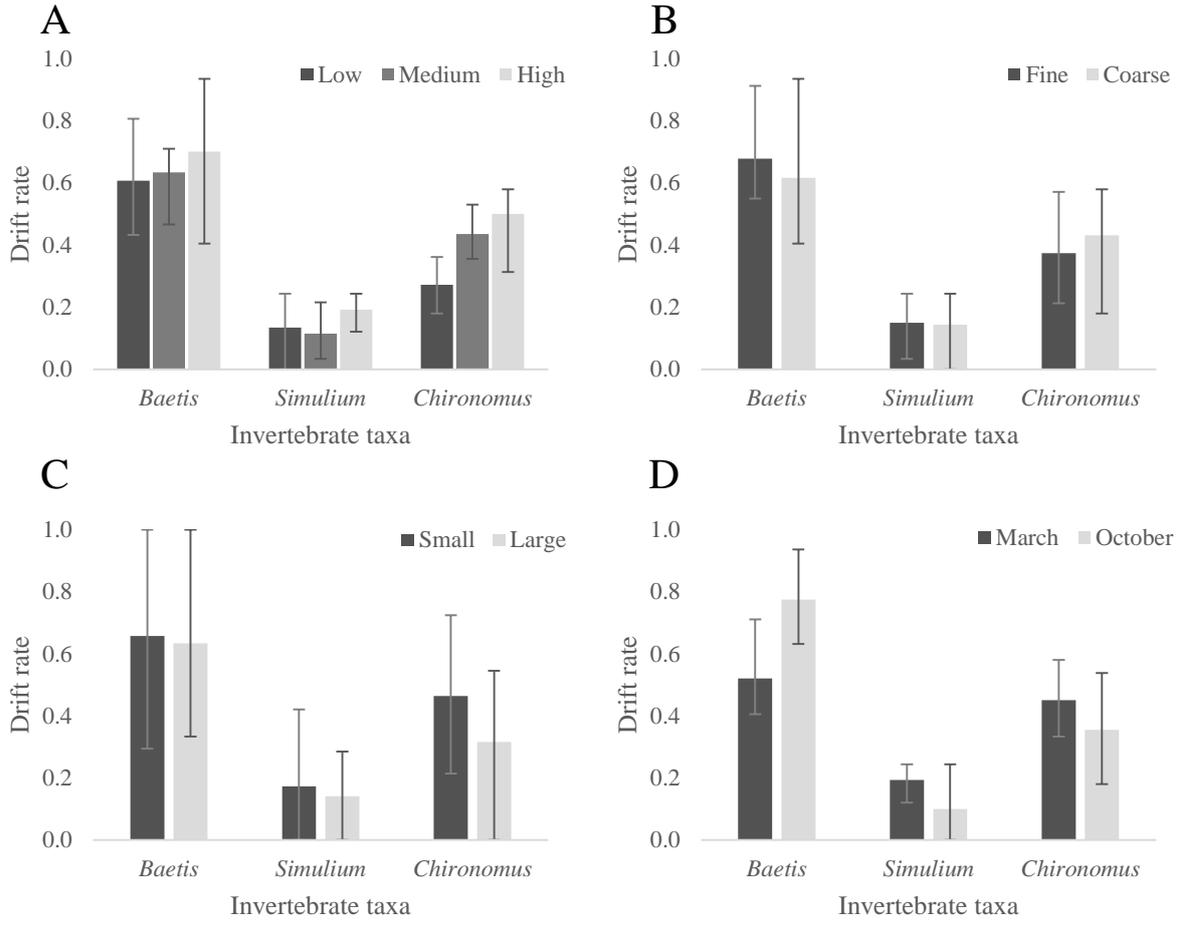


Figure 1 Average drift rate of the three invertebrate taxa according to water velocity (A: low flow in dark, medium flow in grey and high flow in light), gravel bed quality (B: fine gravel in dark and coarse gravel in light), head capsule width (C: small individuals in dark and large individuals in light) and season (D: March in dark and October in light). Bars indicate minimum and maximum drift rate values.

Data were then analysed using a modelling approach computed with OpenBUGS®. For each of the j taxon, if $p.D_i$ is the probability of drifting of the i^{th} individual, we assume:

$$D_i \sim \text{Bernoulli}(p.D_i)$$

$$\text{Logit}(p.D_i) = \mu_{j[i]} + \alpha_{j[i]} V_i + \beta_{j[i]} H_i + \gamma_{j[i]} HCW_i + \eta_{j[i]} S_i + \delta_{l[i]}$$

Where $\mu_{j[i]}$ was the average drifting probability and $\alpha_{j[i]}$, $\beta_{j[i]}$, $\gamma_{j[i]}$ and $\eta_{j[i]}$ were fixed-effect parameters for water velocity (V_i), gravel bed quality (H_i), head capsule widths (HCW_i) or body lengths (BL_i) and season (S_i) respectively. A random effect (δ) was drawn from a common distribution:

$$\delta_{l[i]} \sim \text{Normal}(0, \sigma_\delta^2)$$

Parameters were given independent “weakly informative” priors (*i.e.* sampled in a normal distribution $N(0, 100)$ and for σ_δ in a truncated half-Cauchy $t(0, 1, 1)$ distributions – Gelman & Hill, 2006). Models were tested (Table 2) and the model with the lowest deviance information criterion (DIC) was selected (Spiegelhalter et al., 2002).

We used three independent chains, the first 10 000 iterations were discarded as an initial burn-in period. Then, 10 000 further iterations (resulting from one every ten runs) were performed. The convergence of the chains to their ergodic distribution was tested *via* the Gelman-Rubin (GR) diagnostics integrated in OpenBUGS®.

The significance of the parameters was tested with the step function implemented in OpenBUGS®. At each iteration for a variable X , $\text{step}(X)$ equaled 1 if $X \geq 0$ and equaled 0 if $X < 0$. At the end of the run, if $P(X > 0)$ was lower than 0.1 or higher than 0.9, the parameter X was considered to be different from 0.

Results

The best model with the lowest DIC included the effect of water velocity (α), HCW (γ – instead of BL) and season (η), while the habitat (β) was excluded (DIC = 1573 – Table 2). Under our conditions, the gravel bed did not affect the drift probability for any taxa.

Table 2 Model selection based on minimum Deviance Information Criterion (DIC). Selected model is indicated by bold characters.

Model	Deviance	DIC	Estimated parameter number
$\text{Logit}(p.D_i) = \mu$	1645	1651	3.006
$\text{Logit}(p.D_i) = \mu + \alpha_{j[i]}V_i + \gamma_{j[i]}HCW_i + \delta_{l[i]}$	1517	1578	30.46
$\text{Logit}(p.D_i) = \mu + \alpha_{j[i]}V_i + \beta_{j[i]}H_i + \gamma_{j[i]}HCW_i + \delta_{l[i]}$	1515	1579	32.06
$\text{Logit}(p.D_i) = \mu + \alpha_{j[i]}V_i + \beta_{j[i]}H_i + \gamma_{j[i]}HCW_i + \eta_{j[i]}S_i + \delta_{l[i]}$	1520	1576	27.88
$\text{Logit}(p.D_i) = \mu + \alpha_{j[i]}V_i + \gamma_{j[i]}HCW_i + \eta_{j[i]}S_i + \delta_{l[i]}$	1521	1573	26
$\text{Logit}(p.D_i) = \mu + \alpha_{j[i]}V_i + \gamma_{j[i]}BL_i + \delta_{l[i]} + \eta_{j[i]}S_i$	1525	1578	26.34

When submitted to low, medium and high water velocities, a taxa-specific drifting behaviour was observed (Figure 2). Although all three taxa followed the same pattern and increased their drift probability with water velocity, only *Chironomus* showed a significant relationship ($P(\alpha_C > 0) = 0.997$; $P(\alpha_B > 0) = 0.889$ and $P(\alpha_S > 0) = 0.841$ – Table 3). At low water velocity (10 cm s^{-1}), 44% of *Baetis*, 16% of *Simulium* and 26% of *Chironomus* drifted. At medium water velocity (30 cm s^{-1}), 53% of *Baetis*, 21% of *Simulium* and 45% of *Chironomus* drifted. At high velocity (40 cm s^{-1}), 58% of *Baetis*, 25% of *Simulium* and 55% of *Chironomus* drifted.

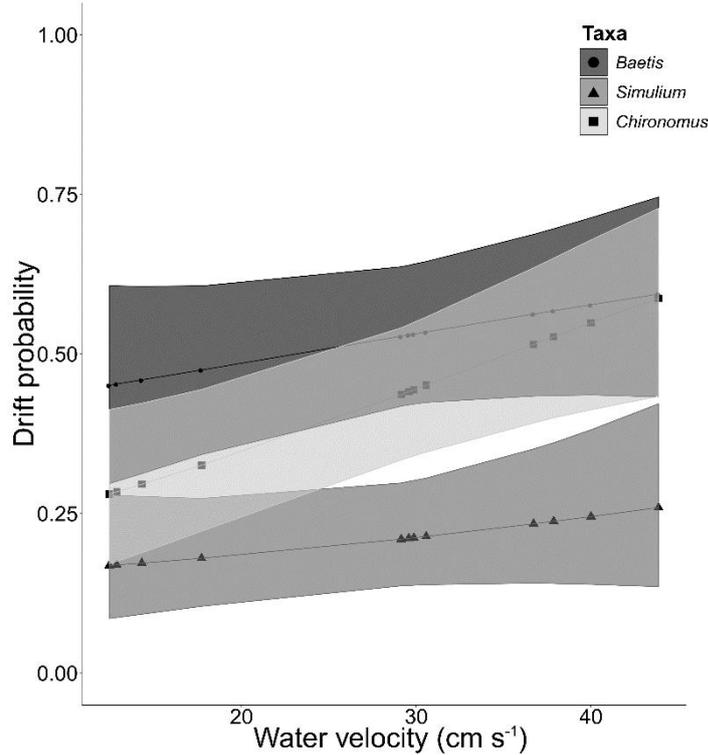


Figure 2 Model estimates of the drift probabilities of the three invertebrate taxa according to water velocity (cm s^{-1}). Shaded areas are the 95% probability intervals of posterior distributions.

Table 3 Main statistics of the posterior probability distribution functions of the free parameters. Parameters are considered significant when $P(X > 0)$ is less than 0.10 or above 0.90 (in bold).

Parameters	Mean	SD	2.5%	Median	97.5%	$P(X > 0)$	
Water velocity	α_B <i>Baetis</i>	0.019	0.016	-0.012	0.019	0.051	0.889
	α_S <i>Simulium</i>	0.018	0.018	-0.017	0.017	0.054	0.841
	α_C <i>Chironomus</i>	0.042	0.014	0.014	0.042	0.071	0.997
HCW	γ_B <i>Baetis</i>	0.183	0.655	-1.101	0.181	1.469	0.609
	γ_S <i>Simulium</i>	-6.512	1.804	-10.130	-6.493	-3.008	0.000
	γ_C <i>Chironomus</i>	-4.979	2.215	-9.316	-4.987	-0.656	0.012
Season	η_B <i>Baetis</i>	1.208	0.341	0.552	1.203	1.900	0.999
	η_S <i>Simulium</i>	-1.216	0.405	-2.031	-1.209	-0.443	0.001
	η_C <i>Chironomus</i>	-0.253	0.309	-0.866	-0.253	0.356	0.200
SD of the random effects	σ_δ	0.396	0.129	0.158	0.390	0.667	1.000

Head capsule width was not related to the propensity to drift in *Baetis* ($P(\gamma_B > 0) = 0.609$). *Simulium* showed a significant relationship between HCW and drift ($P(\gamma_S > 0) = 0.000$), similar to *Chironomus* ($P(\gamma_C > 0) = 0.012$), with small individuals drifting more than large ones in both cases (Figure 3).

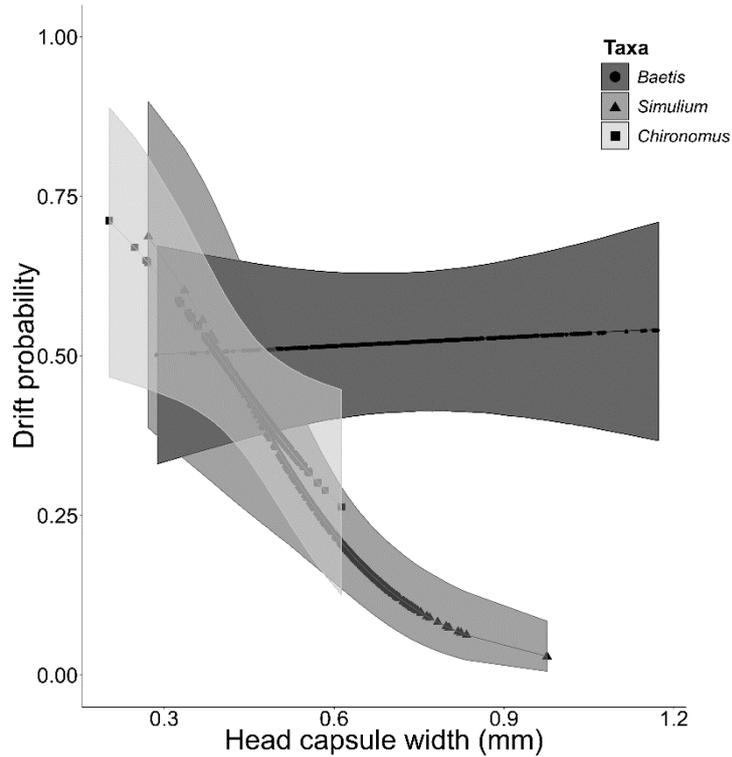


Figure 3 Model estimates of the drift probabilities of the three invertebrate taxa according to head capsule width (mm). Shaded areas are the 95% probability intervals of posterior distributions.

The effect of season on the drifting rates (η) was significant for *Baetis* and *Simulium* ($P(\eta_B > 0) = 0.999$; $P(\eta_C > 0) = 0.200$ and $P(\eta_S > 0) = 0.001$). The drift probability of *Baetis* was higher in October than in March (0.522 in March and 0.782 in October – Figure 4) while the drift probability of *Simulium* was lower (0.206 in March and 0.073 in October). No pattern appeared in the estimations of δ and few values were different from 0 (Table S2), which indicates that no interactions were detected.

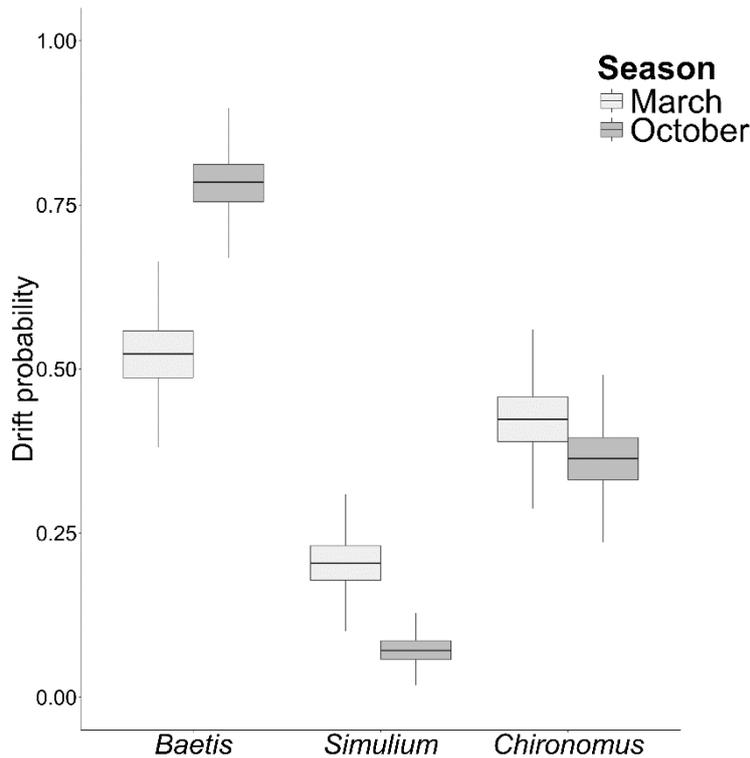


Figure 4 Model estimates of drift probabilities of the three invertebrate taxa according to the season for mean width individual and mean water velocity. Boxplots indicate the 1, 25, 50, 75 and 99 percentiles of posterior distributions.

Discussion

This experimental study provides a quantification of the drift rate for three taxa of invertebrates selected because of their abundance in riverine ecosystems and their potential contribution to drift-feeding fish (Sánchez-Hernández et al., 2011). The drift of *Chironomus* was positively related to water velocity while a positive, but non-significant, trend was observed for *Baetis* and *Simulium*. There was no effect of the substrate size (fine/coarse gravel bed) on any taxa and small *Simulium* and *Chironomus* drifted more than large ones. A seasonal effect was also detected, with *Baetis* drifting more in fall and *Simulium* drifting more in spring.

Under normal flow conditions, invertebrates are distributed along the river depending on their habitat preferences, their ability to cope with water velocity and their trophic requirements (Fjellheim, 1996; Rempel et al., 2000). *Baetis* is a rheophilic taxon and is commonly associated with medium flow habitat, between 25 and 50 cm s⁻¹ (Tachet et al., 2010). They prefer stones that are exposed to water current, as this exposure allows them to feed on benthic microscopic algae and to benefit from the high dissolved oxygen levels of swift currents. Their location above the gravel bed and exposure to the current make them very prone to drift (Kohler, 1983, 1985). Accordingly, *Baetis* demonstrated the highest drift propensity among the three taxa

tested. Gibbins et al. (2005) highlighted that 50% of *Baetis* mayflies drifted with a fixed-bed sediment and that drift rate increased only when water current was strong enough to initiate bed-load transport. In the present experiment, flow was not high enough to mobilise the substrate to induce the passive drift of *Baetis*. As such, and in accordance with literature (Fonseca & Hart, 1996), we assume that in the present experiment the high drift rate of *Baetis* was largely active.

Simulium is the most rheophilic of the 3 taxa, as they demonstrate a preference for water velocities above 20 cm s^{-1} (Tachet et al., 2010), are able to withstand high flows around 90 cm s^{-1} (Finelli et al., 2002), and can be found in harsh conditions with water velocities around 300 cm s^{-1} (Phillipson, 1957; Wotton, 1985). Attached to rocks, they feed by catching fine particles in the water with their filter mandibles. Adapted to high water velocities, they drift more in slow than in fast waters (Fenoglio et al., 2013). In our experiment, they displayed the lowest drift rate and there was no significant relationship between their drift propensity and the tested water velocity gradient, which led us to presume that they drift actively. Indeed, exposed to experimental floods, Fingerut et al. (2015) showed that larval benthic densities of *Simulium* declined in microhabitats with near-bed velocities above 100 cm s^{-1} , much higher than velocities recorded under our conditions.

Chironomus is quite ubiquitous along the water velocity gradient (Tachet et al., 2010) and in the present experiment few individuals drifted under the lowest flow conditions. In running waters, they usually live buried in the substrate or within cocoons feeding on fine particulate organic matter (Berg, 1995). Without any morphological adaptations to cope with the drag force of the current, *Chironomus* was the only taxon that drifted significantly more when the water velocity increased and then, exhibited passive drift. In the field, many authors reported that a high proportion of the drifting community is composed of *Chironomidae* (Imbert & Perry, 2000; Kennedy et al., 2014; Robinson et al., 2004), which under flood conditions, respond as soon as the flow begins to increase. Despite this high sensitivity to flow, the analysis of long-term data (13-years) showed that chironomids are among the most resilient taxa of the community following catastrophic floods, in relation to their ecological traits (*i.e.* their abundance and their short life cycle – Woodward et al., 2015).

Taniguchi & Tokeshi (2004) emphasised the role of shelter availability in invertebrate size distribution, with larger proportions of small individuals in complex habitats. Some works have highlighted the importance of gravel bed quality in limiting the impact of disturbances such as floods and flow variations (Holomuzki & Biggs, 2003; Long et al., 2011). We did not find any

evidence of gravel bed quality impact on drift rate. As *Baetis* and *Simulium* drifted actively, habitat could not mitigate the effects of water velocity on the passive drift of these two taxa. However, considering that *Chironomus* are passive drifters, the lack of relation between drift rates and gravel bed quality could indicate that gravel bed quality was not different enough to induce a distinction between sheltering opportunities. Perhaps this result is due to the fact that we worked with small larvae (Figure S1 and S2) and that the size of the interstices provided by the fine gravel bed was too large to effectively limit sheltering opportunities.

No significant relationship between drift and head capsule width (HCW) was observed for *Baetis*, while *Simulium* and *Chironomus* HCW data showed a clear trend for larvae with small HCW to drift more. This could possibly be linked to a reduction in the ability of young individuals to access shelter due to lower locomotor abilities, at least in *Chironomus* larvae. Concerning *Simulium*, we emphasized that water velocities underwent in the present experiment were much lower than their preferred water velocity (Fingerut et al. 2015). Presumably, their drift was then active. Environmental conditions were good but low organic matter in the experimental channels may have motivated voluntary drift. It may have concerned in priority small *Simulium* since they need higher organic concentration than big ones to optimize their growth (Charpentier & Morin, 1994). *Baetis* are good swimmers, especially compared to the two Diptera used in this experiment. The lack of significant relationship between HCW and drift appears consistent with their swimming abilities, which may have given *Baetis* similar opportunities to reach shelters regardless of their developmental stage. When considering BL, results showed similar patterns to HCW, except for *Chironomus* (see Figure S1). Head capsule width is highly correlated with size in *Baetis* and *Simulium* but not in *Chironomus*. In *Chironomus*, the relationship between larval stages and body length is weak (Richardi et al., 2013). Thus, ontogenesis could be more relevant than larval body size in favouring drift for Diptera larvae. As far as we know, there is no literature linking invertebrate size to drift, except for daylight/night drifting differences. Drift rate of small individuals is higher during daylight while larger individuals drift more at night (Stewart & Szczytko, 1983; Allan, 1984; Bowles & Short, 1988), suggesting that small individuals are more likely to experience passive drift than large ones. In the present experiment, daylight and night-time drift levels were quite similar, but they were not interpreted because of the progressive depletion of invertebrates through time.

Unexpectedly, a seasonal effect on the drift of *Baetis* and *Simulium* was detected. *Baetis* drifted more in fall than in spring, while the opposite was observed for *Simulium*. The mean water

temperature was 2°C warmer in fall, and because invertebrates are ectotherms, this resulted in an increase in metabolism and activity. In addition, Kohler (1983) emphasised an increase in the feeding intensity in fall with *Baetis* more frequently observed on the surface of stones and more prone to drift. For *Simulium*, little information is available on their propensity to drift according to season. Stoneburner & Smock (1979) examined the composition of the drift over the course of a year in a California stream, and noted two peaks for Simuliid densities in drift. Two species of Simuliid were identified with one drifting preferentially in early spring and the second in fall, and these differences were assumed to be driven by their emergence dates. As Simuliids collected for our experiment were larger in March than in October (Figure S2), their higher drift rate in March might possibly be related to a time proximity with their emergence date at that time.

The present experiment was conducted in a closed system. This allowed natural processes such as immigration or the hatching of invertebrates to be avoided, which would have prevented the estimation of the drift entry rate. Ranging from 12 to 43 cm s⁻¹, water velocities obtained in this experiment are representative of conditions in the natural environment. *Baetis* and *Simulium* drifted actively and conditions were not stringent enough to induce significant passive drift. Ranging from 0.36 to 7.34 dynes cm⁻², shear stress experienced by invertebrates in our channels was lower than the 9 dynes cm⁻² threshold needed to dislodge the invertebrates and to induce passive drift (Gibbins et al., 2007). Small *Simulium* demonstrated an abundance of drift, probably through active drift with low velocities driving the movement of individuals. In contrast, *Chironomus*, and specifically young stages, drifted passively.

This work provides the first accurate estimates of invertebrate drift entry rate. The accuracy of these rates are important because they allow the impacts of disturbances on invertebrate communities to be predicted, and they improve models of trophic interactions, such as those for drift-feeding fishes (Hayes et al., 2007).

Acknowledgements

The authors declare no conflicts of interest. E. Arevalo benefits from a cross-border grant (Univ Pau & Pays Adour/UPV) and from Mérimée, a program funded by the French Embassy. Experiments were carried out thanks to the IE ECP facilities and financially supported by CG64 and INRA Ecoserv Metaprogram. We wish to thank F. Gueraud, P. Coste, E. Huchet and J. Rives for their help with the experimental set-up and invertebrate sampling, J. Labonne for his comments on the manuscript and J. Almany for correcting the English manuscript.

Supporting Information

Table S1 Step by step experimental procedure with the sample number and if it is included (Y) or not (N) in the model.

N° sample	Included in the model	12 th of March 2015	Time	22 nd of October 2015	N° sample	Included in the model
	N	Addition of invertebrates	09:00	Addition of invertebrates		N
1	N	Opening of the flow at low level	10:00	Opening of the flow at low level	1	N
2	N		11:00		2	N
3	N		12:00		3	N
4	Y	Gradual elevation of flow	12:15	Gradual elevation of flow	4	Y
5	Y		12:30		5	Y
6	Y		12:45		6	Y
7	Y		13:00		7	Y
8	Y		14:00		8	Y
9	Y		15:00		9	Y
10	Y		16:00		10	Y
11	Y		17:00		11	Y
12	Y		18:00		12	Y
13	Y	Dusk	19:00	Dusk	13	Y
14	Y		19:30		14	Y
15	Y		20:00		15	Y
16	Y		21:00		16	Y
17	Y		22:00		17	Y
18	Y		23:00		18	Y
19	Y		00:00		19	Y
20	Y		01:00		20	Y
21	Y		02:00		21	Y
22	Y		03:00		22	Y
23	Y		04:00		23	Y
24	Y		05:00		24	Y
25	Y		06:00		25	Y
26	Y	Dawn	06:30	Dawn		
27	Y		07:00		26	Y
			07:30		27	Y
28	Y		08:00		28	Y
29	Y		09:00		29	Y
30	Y		10:00		30	Y
31	Y		11:00		31	Y
32	Y	End of the experiment	12:00	End of the experiment	32	Y
33	Y	Recovery of the invertebrates settled in the channels		Recovery of the invertebrates settled in the channels	33	Y

Table S2 Probabilities of the random effects (the residuals) to be different from 0. Parameters are considered significant when $P(\delta[X] > 0) < 0.10$ or $P(\delta[X] > 0) > 0.90$ and are indicated in bold characters.

δ	Taxon	Flow	Season	Gravel bed	$P(\delta[X] > 0)$
1	<i>Baetis</i>	High	March	Coarse	0.121
2	<i>Baetis</i>	High	March	Fine	0.381
3	<i>Baetis</i>	Medium	March	Coarse	0.323
4	<i>Baetis</i>	Medium	March	Fine	0.924
5	<i>Baetis</i>	Low	March	Coarse	0.442
6	<i>Baetis</i>	Low	March	Fine	0.753
7	<i>Baetis</i>	High	October	Coarse	0.842
8	<i>Baetis</i>	High	October	Fine	0.749
9	<i>Baetis</i>	Medium	October	Coarse	0.097
10	<i>Baetis</i>	Medium	October	Fine	0.297
11	<i>Baetis</i>	Low	October	Coarse	0.676
12	<i>Baetis</i>	Low	October	Fine	0.310
13	<i>Simulium</i>	High	March	Coarse	0.385
14	<i>Simulium</i>	High	March	Fine	0.233
15	<i>Simulium</i>	Medium	March	Coarse	0.505
16	<i>Simulium</i>	Medium	March	Fine	0.470
17	<i>Simulium</i>	Low	March	Coarse	0.765
18	<i>Simulium</i>	Low	March	Fine	0.665
19	<i>Simulium</i>	High	October	Coarse	0.891
20	<i>Simulium</i>	High	October	Fine	0.829
21	<i>Simulium</i>	Medium	October	Coarse	0.137
22	<i>Simulium</i>	Medium	October	Fine	0.391
23	<i>Simulium</i>	Low	October	Coarse	0.275
24	<i>Simulium</i>	Low	October	Fine	0.441
25	<i>Chironomus</i>	High	March	Coarse	0.681
26	<i>Chironomus</i>	High	March	Fine	0.370
27	<i>Chironomus</i>	Medium	March	Coarse	0.430
28	<i>Chironomus</i>	Medium	March	Fine	0.371
29	<i>Chironomus</i>	Low	March	Coarse	0.538
30	<i>Chironomus</i>	Low	March	Fine	0.634
31	<i>Chironomus</i>	High	October	Coarse	0.750
32	<i>Chironomus</i>	High	October	Fine	0.099
33	<i>Chironomus</i>	Medium	October	Coarse	0.936
34	<i>Chironomus</i>	Medium	October	Fine	0.504
35	<i>Chironomus</i>	Low	October	Coarse	0.209
36	<i>Chironomus</i>	Low	October	Fine	0.494

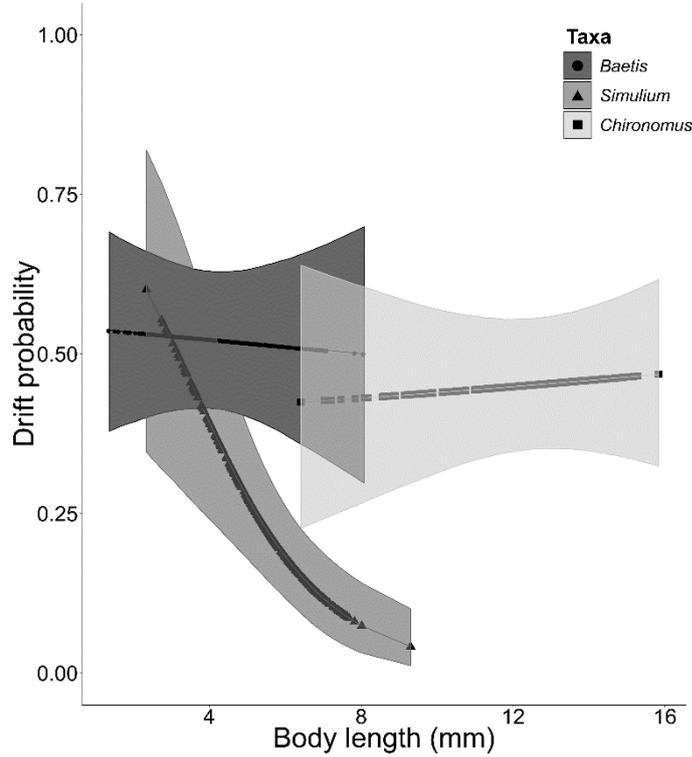


Figure S1 Estimates of the drift probabilities of the three invertebrate taxa according to body length (mm). Shaded areas are the 95% probability intervals of posterior distributions.

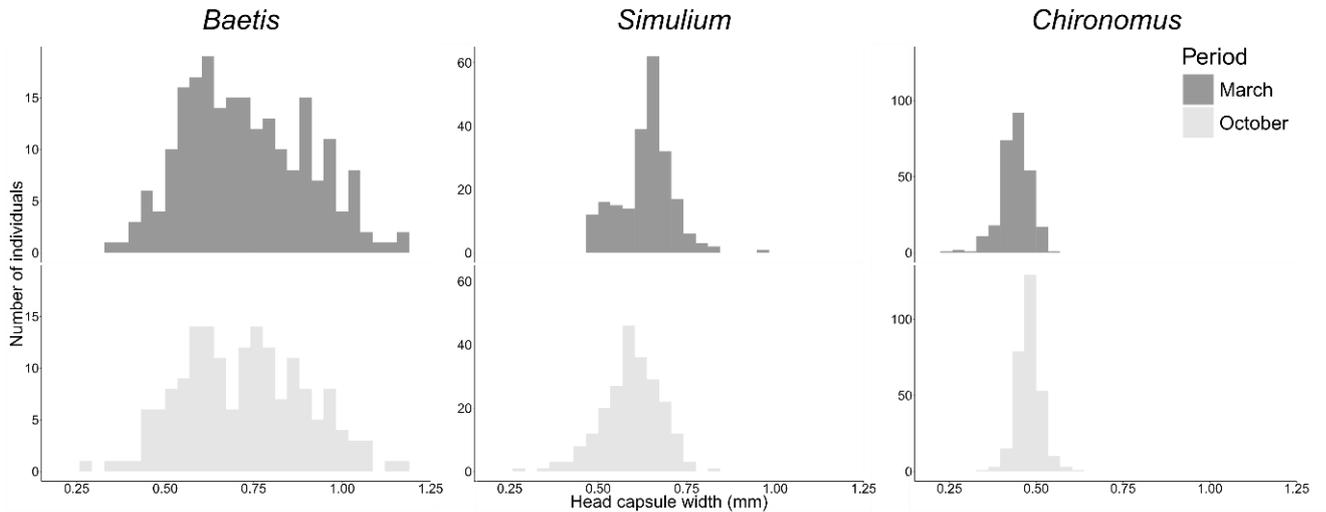


Figure S2 Distribution of the head capsule width (in mm) for the three taxa and the two trials. Differences in HCW between March and October were not significant for Baetis ($P(\text{mean}(\text{HCW}_{\text{March}}) > \text{mean}(\text{HCW}_{\text{October}})) = 0.671$). Simulium were larger in March ($P(\text{mean}(\text{HCW}_{\text{March}}) > \text{mean}(\text{HCW}_{\text{October}})) = 1.000$) and Chironomus were larger in October ($P(\text{mean}(\text{HCW}_{\text{March}}) > \text{mean}(\text{HCW}_{\text{October}})) = 0.000$).

Table 1.1 Synthesis of the main results on the drift propensity of *Baetis*, *Simulium* and *Chironomus* according to water velocity, gravel bed, individual size and season.

Taxa	Mean drift probability	Water velocity 14, 30, 40 cm.s ⁻¹	Gravel bed Fine vs. Coarse	Individual size Head capsule width	Season Spring vs. Autumn
<i>Baetis</i>	0.684	-	-	-	↗ Drift in Autumn
<i>Simulium</i>	0.128	-	-	Small individuals drift more <i>Active: not enough organic matter</i>	↗ Drift in Spring
<i>Chironomus</i>	0.363	↗ Water velocity ↗ Drift rate	-	Small individuals drift more <i>Passive: carried by the current</i>	-

Further considerations and discussion

Results included in the MS showed that head capsule width interfered with water velocity in *Chironomus* and *Simulium* larvae propensity to drift. The model presented in the article estimated an effect of the individual head capsule width (HCW) by taxon (γ). To test our hypotheses about the drift propensity of small *Simulium* and *Chironomus* (*i.e.* small *Simulium* leaving actively the channels and small *Chironomus* carried by the current), the model was modified to estimate an effect of the individual size (HCW) by taxon and by water velocity modality.

Concerning *Simulium*, all the estimated parameters were negative (Table 1.2). The effect of HCW at low velocity (γ_{S1}) was significant ($P(\gamma_{S1} > 0) = 0.000$), while the effects at medium (γ_{S2}) and high velocities (γ_{S3}) were around the threshold of significance ($P(\gamma_{S2} > 0) = 0.122$; $P(\gamma_{S3} > 0) = 0.096$). γ_{S1} was significantly different from both γ_{S2} and γ_{S3} with probabilities of 0.022 and 0.030 respectively. γ_{S2} and γ_{S3} were equal ($P(\gamma_{S2} > \gamma_{S3}) = 0.551$). Small *Simulium* drifted more at low water velocities than at medium or high water velocities (Figure 1.1).

Concerning *Chironomus*, all the estimated parameters were negative (Table 1.1) but only γ_{C2} and γ_{C3} were significant ($P(\gamma_{C1} > 0) = 0.468$; $P(\gamma_{C2} > 0) = 0.048$; $P(\gamma_{C3} > 0) = 0.013$). γ_{C2} was not different from γ_{C3} and γ_{C1} ($P(\gamma_{C1} > \gamma_{C2}) = 0.887$; $P(\gamma_{C2} > \gamma_{C3}) = 0.605$) but γ_{C1} was significantly higher than γ_{C3} ($P(\gamma_{C1} > \gamma_{C3}) = 0.939$). There was no relationship between the head capsule width of invertebrate and the drift probability when the water velocity was low. However, small individuals drifted more in medium and high flow conditions (Figure 1.1). Results of this model supported the hypotheses proposed in the MS about the effect of individual head capsule width on the drift of invertebrates.

Table 1.2 Main statistics of the posterior probability distribution functions of the effect of the individual size (γ) by taxon (*Simulium* and *Chironomus*) and by water velocity modality (low, medium and high). Parameters are considered significant when $P(X > 0)$ is less than 0.10 or above 0.90 (in bold).

HCW parameters	Taxon	Water velocity modality	Mean	$P(X > 0)$
γ_{S1}	<i>Simulium</i>	Low	-12.520	0.000
γ_{S2}		Medium	-3.400	0.122
γ_{S3}		High	-3.896	0.096
γ_{C1}	<i>Chironomus</i>	Low	-0.241	0.468
γ_{C2}		Medium	-6.517	0.048
γ_{C3}		High	-7.899	0.013

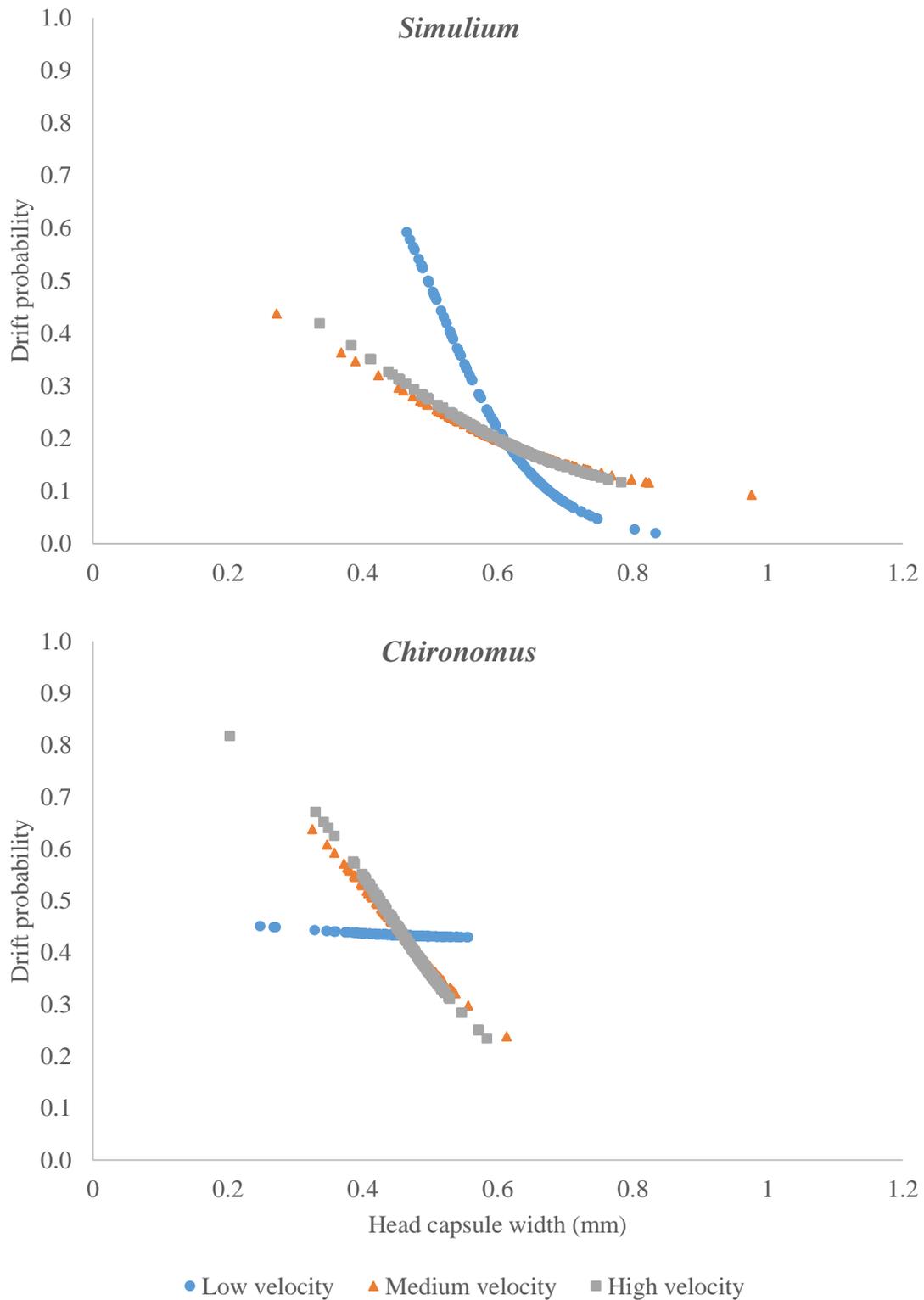


Figure 1.1 Model estimates of the drift probabilities of *Simulium* and *Chironomus* according to head capsule width (mm) at low (blue circles), medium (orange triangles) and high (grey squares) water velocities.

According to the present experiment, moderate water velocity induces passive drift of invertebrates even if bed load transport is not initiated. Then, besides the effect of catastrophic hydrological events that are known to deplete the benthos, an increase in the frequency of moderate floods may also affect their abundance.

For fish, the increase in water velocity increases the energetic costs to maintain hunting positions. They would possibly take advantage of an increase of food (especially *Chironomus*) up to a threshold (25 cm s^{-1} according to Heggenes & Traaen 1988 for alevins). However, it remains to be calculated if the increase in swimming expenditure is balanced by both the increase of energy intake and the decrease of the hunt efficiency. In anyway, at higher water velocities, they will try to shelter the time of the flood, without benefiting from the drifting invertebrates.

Then if the increase in floods frequency forecasted by Global Climate Change exacerbates the risk of depletion of benthic invertebrates it should lead to a decrease in food availability for carnivorous fish. This may especially affect 0^+ young salmonids for at least 4 reasons: *i/* they start exogenous feeding in spring when the risk of flood events is still high (and this risk will increase with GCC), *ii/* they are constrained by the small size of their oesophagus to small sized prey, *iii/* they have much less reserve than large fish to face starvation periods, *iv/* their swimming ability is limited at high water velocity.

Chapter II Effect of feeding conditions and temperature on growth and metabolism of alevins

Study context

The previous chapter highlighted that the increase in water velocity and flood frequency due to Global Climate Change should increase invertebrate drift (and especially of *Chironomus*). This might result in the depletion of the benthic invertebrate community, and on the drifting one as drifting invertebrates come from the benthos. For drift feeding fish, this means an increased risk to face periods of food scarcity.

When salmonid alevins emerge, they have almost exhausted the reserves of their yolk sac and they need to start feeding quickly (Johnson et al. 2013; Ladago et al. 2016). For many fish species, starvation during larval stage induces high mortality rates (Hunter 1981; Jonas & Wahl 1998) and it is an important driving force for 0⁺ salmon (*Salmo salar*) survival (Kennedy et al. 2008) and growth (Ward et al. 2009). In the wild, most species regularly face periods of food deprivation and besides behavioural responses, they may adapt to the quantity of resources available by lowering their metabolism (McCue 2010). The basal metabolism of an organism, its environment (temperature) and the amount of its body resources (usually lipids), are important factors acting on its ability to face long period of starvation.

Whatever the organism, at the beginning of starvation, pancreas increases the secretion of glucagon and diminish the secretion of insulin, which trigger catabolism in the liver and mobilisation of reserves (Sundby et al. 1991). First, liver degrades glycogen in glucose *via* glycogenolysis pathway. Carbohydrates are then used by the brain and carbohydrate-dependent cells (*e.g.* red blood cells). However, salmonids are carnivorous fish and they have adapted their anatomy, physiology and metabolism to their natural diet, which contains few or no carbohydrates (Kamalam et al. 2017). They mainly synthesise glucose from non-carbohydrate precursors *via* gluconeogenesis (*i.e.* lactate, pyruvate and amino acids – NRC 2011). When stocks of carbohydrates are depleted, lipids are catabolised *via* the lipolysis pathway. Triglycerides contained in body fat release fatty acids in the blood, which are caught by the liver and oxidized in acetyl-Coenzyme A (acetyl-CoA) *via* the β -oxidation pathway. Acetyl-CoA is used for the synthesis of ketone bodies to feed muscles and the brain after transformation, or for energy synthesis. Finally, when carbohydrates and lipids are depleted, muscular proteins are mobilised. Muscles constitute the main protein mass and a reserve of amino acids. Muscle proteolysis releases amino acids in the blood that are catabolized by the liver *via* amino acid catabolism. Amino acid nitrogen components are eliminated in urea, while carbon skeletons are converted either into glucose *via* gluconeogenesis or into acetyl-CoA for the synthesis of energy. Acetyl-CoA obtained from the β -oxidation or from the catabolism of

amino acids is invested first in Krebs cycle and then in the respiratory chain for the synthesis of ATP (Figure 2.1 – Simpkins et al. 2003; Hecketsweiler & Hecketsweiler 2004). Synthesis of adenosine triphosphate (ATP) provides the energy needed for all the chemical reactions of metabolism. Fatty acid catabolism and amino acid catabolism can be activated sequentially but some organisms (and particularly larvae, as found in European plaice by Ehrlich 1974) can use them in conjunction to preserve the lipid reserve (and dispose of it longer) as it is the most cost-effective way to produce energy (Bar 2014).

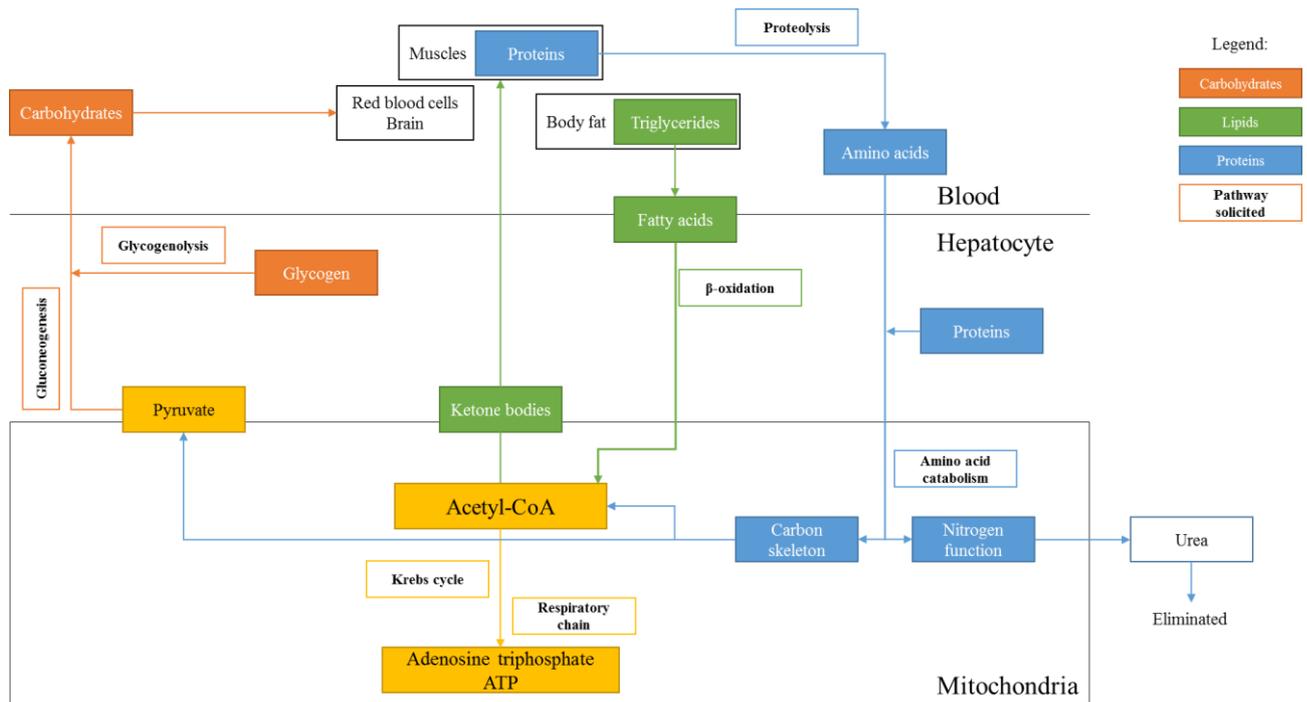


Figure 2.1 Metabolic flows for all living organisms (including salmonids) in fasting situation (modified from Hecketsweiler & Hecketsweiler 2004).

Temperature is an important factor that regulates the metabolism of organisms, especially in poikilotherms, which do not regulate their internal temperature. Warmer temperatures increase their metabolism and so the reserve consumption rate. Therefore, Global Climate Change might affect young trout growth and survival through both a reduction in prey availability and an increase in metabolic loss. Thus in that chapter, we conducted an experiment in order to better understand the interactions of these two factors (prey availability and temperature) on young trout growth and metabolic pathways.

Effect of food shortage and temperature on 0⁺ salmonids: a contribution to predict the effects
of Climate Change

Arevalo Elorri¹, Panserat Stéphane², Seiliez Iban², Larrañaga Aitor³, Bardonnnet Agnès¹

¹ ECOBIOP, UMR 1224, INRA, Univ Pau & Pays Adour, 64310, Saint-Pée-sur-Nivelle, France

² NUMEA, UMR 1419, INRA, Univ Pau & Pays Adour, 64310, Saint-Pée-sur-Nivelle, France

³ Laboratory of Stream Ecology, University of the Basque Country, Bilbao, Spain

Accepted in Journal of Fish Biology

ABSTRACT

In the present experiment, brown trout alevins were maintained at 8°C and 11°C at 3 conditions over a 9 day period from yolk sac exhaustion: fed *ad libitum*, starved or fed *ad libitum* after starvation. Whole body gene expressions for proteins involved in energy metabolism and the two primary proteolytic pathways were assessed and showed that proteasome and autophagy-related genes were over-expressed during and after starvation, particularly at 11°C. Our results suggest that higher temperature will intensify stress induced by starvation.

Key words: *Temperature, Salmo trutta, fry, starvation, prey, metabolism.*

Introduction

Climate Change (CC) will alter precipitation patterns (IPCC 2013) and will lead to an increase in winter/spring rainfall in most coastal areas of the north Atlantic. In freshwater ecosystems, rains are the primary source of running water discharge in rivers (Giller & Malmqvist, 1998) and with heavier and more frequent rainfall predicted, flooding events will become more frequent and intense in riverine systems. Further, CC will increase temperatures globally (IPCC 2013), which will affect metabolic rates, vital activities and growth of ectotherm organisms (Allan & Castillo, 2007).

Brown trout (*Salmo trutta* Linnaeus) alevins start their exogenous feeding in spring, which usually coincides with high prey availability (Romaniszyn et al., 2007). When the young trout emerge from the gravel, they have a limited amount of reserves and thus need to start feeding quickly to avoid mortality. Emergence is described as a critical period (Elliott, 1994), where high mortality rates occur (Elliott, 1986). More recent studies demonstrated that starvation during the critical period was an important driving force for 0⁺ salmon (*Salmo salar*) survival

rates (Kennedy et al., 2008) and growth (Ward et al., 2009). The effect of temperature on metabolism is another factor affecting the survival of emerged salmonids. Régnier et al. (2013) showed that metabolic rate increased by 1.4-1.6 in response to a 4°C temperature increase for hatched brown trout. Thus, CC might affect young trout growth and survival through both a reduction in prey availability, as flood frequency and magnitude have the potential to drastically reduce the invertebrate community (Robinson et al., 2004), as well as through an increase in metabolic loss driven by rising temperatures.

While salmonid growth and survival at the swim-up stage have been relatively well studied (Bilton & Robins, 1973; Twongo & MacCrimmon, 1976; Koss & Bromage, 1990; Edsall et al., 2003), empirical data to assess the interaction between temperature and first-feeding are sparse. In addition, metabolic pathways activated at a molecular level by the starvation of early life stages have not yet been described.

The aims of the present study were to better understand young brown trout response to CC by (1) describing the consequences of starvation on their nutritional status, (2) testing their ability to recover from late first feeding and (3) analysing the effect of temperature on these phenomena (starving/recovering) by measuring growth and mRNA levels of genes involved in fatty acid/amino acid catabolism, as well as proteasomal and autophagy pathways.

Material and methods

Experimental features

Experimentation was conducted in the INRA experimental facilities (UMR Ecobiop, Saint-Pée-sur-Nivelle, France) authorized for animal experimentation (A640141). The experiments were in strict accordance with EU legal frameworks related to the protection of animals used for scientific research (Directive 2010/63/EU) and according to the National Guidelines for Animal Care of the French Ministry of Research.

Forty-two tanks (LxWxH: 50x25x30 cm), each equipped with an individual pump and a Perlon filter, were filled to 20 cm height with filtered water from the Nivelle River. Three pebbles (\varnothing : 4-5 cm) were placed in each tank to serve as a place for the fish to shelter. Twenty one tanks were settled in each of two thermo-regulated rooms that were at 8°C (\pm 0.1 in the water) and 11°C (\pm 0.4 in the water). Natural nyctemeral light-dark regime was simulated, with a 6:30 to 19:00 lit period and two 30 min periods of gradual light intensity change mimicking dawn and dusk.

Fish sampling

On 30/12/2014, eggs were obtained through the artificial fertilization of gametes of wild brown trout caught in the Nivelles watershed (43°21' N, 1°33' W). Eggs and alevins were reared at 9.36°C (\pm 1.18) until complete yolk sac depletion (790 degree-days). Survival was high (up to 82%) and on 30/03/2015, 126 alevins were individually weighed and photographed under binocular (x 10), allowing for individual identification thanks to melanophore distribution patterns (Garcia de Leaniz et al., 1994). Each of the 42 tanks was allocated with a random batch of 3 fish. Ten additional alevins were settled in a bucket for 24h at each temperature in aerated water and killed on Day 0 (D₀). Live Chironomid larvae (Grebil, Arry, France) were distributed *ad libitum* from D₀ in 9 tanks at each temperature. Every morning, leftover food was removed and counted, and a known number of new larvae were then added. All tanks (fed F and starved S) received the same pipette disturbance from the feeding. On D₅, fish from 3 tanks from the fed (F₅) and starved conditions (S₅) at each temperature were measured, weighed, photographed and subjected to lethal anesthesia before being frozen in liquid nitrogen and stored at -80°C until analysis. Fish from another 3 tanks from the starved condition received food *ad libitum* (Delayed-feeding, DF). From D₅ to D₉, 6 fed tanks (F₉), 6 unfed tanks (S₉) and 3 delayed-feeding tanks (DF₉) remained. As some mortality occurred at D₉, the experiment was stopped.

Relative quantification of mRNAs levels for catabolic genes

mRNA levels were determined by quantitative real-time RT-PCR. The extraction of total RNA from whole alevins was performed using the Trizol reagent (Invitrogen, Carlsbad, CA, USA). One microgram of total RNA was used for cDNA synthesis. The SuperScript III RNaseH-Reverse Transcriptase Kit (Invitrogen) with random primers (Promega, Charbonnières, France) were used. Primer sequences to amplify specific trout target genes (Seiliez et al., 2008, 2012; Panserat et al., 2017) are shown in the Supporting information (Table S1). Each PCR assay (Lightcycler 480, Roche Diagnostics, Neuilly-sur-Seine, France) included replicate samples (duplicates of reverse transcription and PCR amplification) and 2 negative controls (one with RNA but without RT enzyme and the other without RNA). For the analysis of mRNA levels, relative quantification of target gene expression was performed using the Δ CT method (Pfaffl, 2001). The relative gene expression of Luciferase was used for the normalization (Marandel et al., 2016).

Statistical analyses

Total length was calculated as the average of measurements from 3 different operators to the nearest 0.1 mm using ImageJ (Abràmoff et al., 2004). Weight gain (Wg) and length gain (Lg) were then calculated (eq.1):

$$Gain = \frac{Final - Initial}{Initial}$$

Wg and Lg were analysed using nonparametric Wilcoxon-Man-Whitney rank sum test.

Molecular analyses were performed on 72 individuals, 6 fish per condition and per temperature on D₀, on D₅ (F₅ and S₅) and on D₉ (F₉, S₉, and DF₉). mRNA levels were analysed using a Bayesian modelling approach with OpenBUGS®. Data were analysed separately according to temperature. If X_i is the expression of the i^{th} individual, we assume:

$$\log(X_i) \sim Normal(\mu_i, \sigma)$$

$$\mu_i = \mu + \alpha_{j[i]}$$

Where the mean of the log expression μ_i was modelled according to a global average (μ) and $\alpha_{j[i]}$ was a fixed-effect parameter for rearing conditions.

Bayesian computation

Parameters were given independent ‘non informative’ priors (*i.e.*, sampled in a normal distribution $N(0, 100)$ and for σ in a half-Cauchy $t(0, 1, 1)$ distributions – Gelman & Hill 2006). The convergence of three Markov Chain Monte Carlo (MCMC) chains to their ergodic distribution was tested for the model parameters *via* the Gelman-Rubin (GR) diagnostics. The first 10 000 iterations were discarded and then, 10 000 further iterations (thin = 10) were performed.

Results

Survival

No death was recorded in the fed group, and the same was true for all groups up to D₈. On D₉, 12 fish died: 4 at 8°C (S₉), 8 at 11°C (7 S₉ + 1 DF₉). The experiment was stopped and dead fish were not considered in further biometric and molecular analyses.

Biometric data

As expected, fed fish gained weight and starved fish lost weight (Fig. 1a). If weight gain on D₉ was higher than on D₅ for fed fish, no difference occurred among starved fish between the two

dates. Temperature did not change weight gain in fed fish. On the contrary, for starved fish weight loss was significantly increased by temperature increase (Wilcoxon test, $p\text{-value}(S_5)$ and $(S_9) < 0.01$). The same tendency was observed for delayed fed fish ($p\text{-value}(DF_9) = 0.059$). DF_9 fish exhibited intermediate values between S_9 and F_9 .

At the start of the experiment, fish measured 2.584 cm (± 0.037) and weighed 0.106 g (± 0.003), (box-plots, Fig. 1b). On average, F_9 fish were longer and heavier than S_9 ones, and DF_9 were in between, similar to results for weight gain (Fig. 1a). Weight/length relationships did not differ much among rearing conditions (Fig. 1b). However, it was noteworthy that at 11°C fish were usually shorter than at 8°C for a similar weight (Fig. 1b). Length gain values confirmed this point with higher gains at low temperature (8°C vs 11°C) for F_9 and DF_9 conditions (Wilcoxon test, $p\text{-value}(F_9) = 0.01$; $p\text{-value}(DF_9) = 0.046$; $p\text{-value}(S_9) = 1$).

During the feeding period, DF_9 fish ate as much as the fed fish (F_5 and F_9), but they grew less, especially at 11°C (Fig. 1c and 1d). For F_5 fish, the mean number of *Chironomus* eaten daily almost doubled at 11°C (8.08 on average vs 4.67 on average at 8°C) but it was not correlated with a better performance in growth. Results for F_9 showed the same trend (4.67 prey on average at 8°C and 8.08 at 11°C for a similar growth).

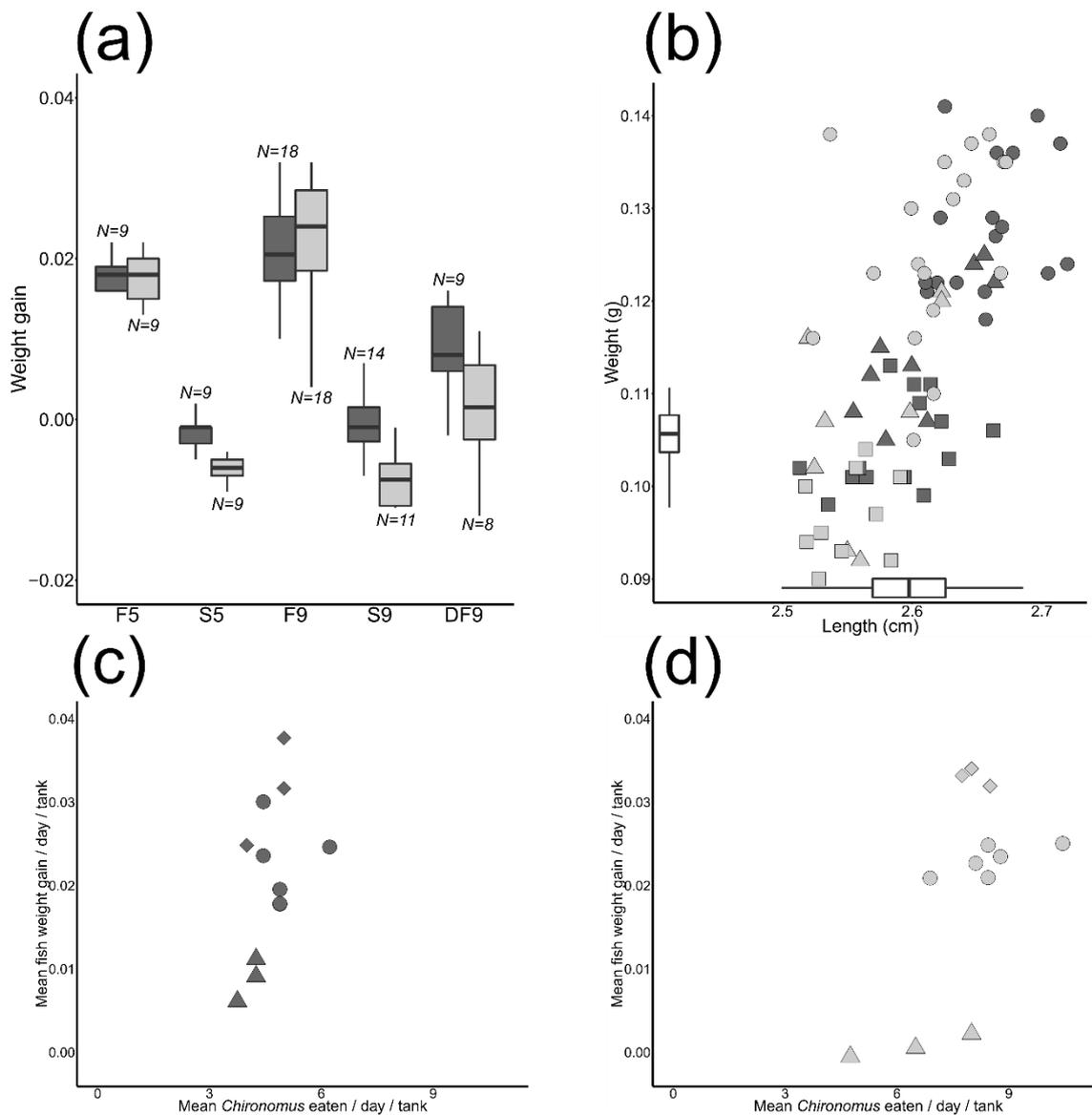


Figure 1 (a): Weight gain ($WG = (final\ weight - initial\ weight) / initial\ weight$) of emerged alevins according to rearing conditions. Boxplots indicate the 1, 25, 50, 75 and 99 percentiles. (b): Length (in cm) and weight (in g) of alevins after 9 days of growth. Boxplots indicate the 1, 25, 50, 75 and 99 percentiles of the initial measures of fish. The daily mean fish WG by tank for the fed conditions (F_9 , DF_9 and F_5) according to the mean number of prey (*Chironomus*) eaten daily by tank at 8°C (c) and 11°C (d). Dark items represent alevins reared at 8°C and light items represent alevins reared at 11°C. Fed fish F_5 are represented in diamond, fed fish F_9 are in circles, delayed-fed fish DF_9 are in triangle and starved fish S_9 are in square.

mRNA levels for catabolic genes

All mRNA levels were usually above 0.5, except in some cases for genes involved in fatty acid catabolism at 11°C, and in proteasome and autophagy in fed fish (Table S2). Many of the differences between feeding conditions were found for genes involved in autophagy and proteasome pathways (Table 1, FC1 to FC8). On D5 and at 8°C, starvation is associated with significantly higher mRNA levels for 4 of the 5 tested genes involved in proteasome and for 1

of the 5 tested involved in autophagy (Fold change, $FC1 < 1$, Table 1). At 11°C, mRNA levels for all the monitored proteasome/autophagy-related genes were significantly higher in S_5 than in F_5 (Table 1, $FC2 < 1$). On D_9 , mRNA levels for autophagy and proteasome genes in S_9 were higher than in F_9 regardless of the temperature ($FC3$ - $FC4$). For fish that were starved for 5 days before feeding (DF_9), mRNA levels for genes coding for autophagy and proteasome were in-between, *i.e.* they were more expressed than those in F_9 ($FC5$ and $FC6$) but less than those in S_9 ($FC7$ and $FC8$). In a comparison of gene expression at different temperatures, FC values were significant ($0.9 < P < 1$) in half of cases, and in all except one, mRNA levels were higher at 11°C ($FC9$ to $FC13$).

Concerning fatty acid catabolism (HOAD and CPT1 genes), FC values in mRNA levels between starved and fed fish were inconsistent between D_5 and D_9 (Table 1, $FC1C2/C3C4$). On D_5 , mRNA levels were higher in starved fish when compared to fed fish (significant $FC < 1$), while lower on D_9 (significant $FC > 1$). Results for amino acid catabolism genes (GDH, ASAT, ALAT genes) were more congruent, indicating higher mRNA levels for these genes in fed fish compared to starved fish on both D_5 and D_9 . For FC values in delayed-feeding (DF) fish and fed (F) fish ($FC5$ - 6), no significant values were reached for genes involved in fatty acid catabolism. Looking at amino acid catabolism, significant FC values were above 1 at 8°C, while they were less than 1 at 11°C, suggesting that mRNA levels for amino acid catabolic genes were higher in fed fish than in delayed fed fish at low temperature, while the opposite was observed at 11°C.

Comparing DF_9 fish with S_9 fish (Table 1, $FC7$ - 8) at 11°C, almost all of the mRNA levels for genes involved in catabolism (8 among 9 tested) were higher in DF_9 . At 8°C only 2 differences for amino acid catabolism genes were observed, also in favour of S_9 .

Concerning the impact of temperature, significant FC values ($0.9 < P < 1$) were observed in a quarter of cases, and all were > 1 , suggesting that genes were expressed at a higher level at 8°C than at 11°C.

Table 1 Comparison of the gene expressions among rearing conditions. Genes involved in fatty acid and amino acid catabolism, proteasome and autophagy were studied (pathways and genes were displayed in the first columns). Fold Changes (FC) were calculated by dividing the mean expression of the condition 1 by the mean expression of the condition 2. Significant differences are in bold and * means that $P(\text{Condition 1} > \text{Condition 2})$ is between 0.90 and 0.95 and ** means that $P(\text{Condition 1} > \text{Condition 2})$ is between 0.95 and 1.00.

FC values		FC1	FC2	FC3	FC4	FC5	FC6	FC7	FC8	FC9	FC10	FC11	FC12	FC13
Pathways	Genes	F ₅ / S ₅ 8°C	F ₅ / S ₅ 11°C	F ₉ / S ₉ 8°C	F ₉ / S ₉ 11°C	F ₉ / DF ₉ 8°C	F ₉ / DF ₉ 11°C	S ₉ / DF ₉ 8°C	S ₉ / DF ₉ 11°C	F ₅ / F ₅ 8/11°C	S ₅ / S ₅ 8/11°C	F ₉ / F ₉ 8/11°C	S ₉ / S ₉ 8/11°C	DF ₉ / DF ₉ 8/11°C
Proteasome	Fbx32	0.10**	0.14**	0.20**	0.11**	0.55*	0.19**	2.82**	1.64**	0.55**	0.78	1.20	0.70*	0.41**
	MuRF1	0.10**	0.16**	0.10**	0.18**	0.33**	0.26**	3.17**	1.49*	0.33**	0.52**	0.67*	1.14	0.54**
	MuRF2	0.20**	0.14**	0.12**	0.06**	0.48**	0.12**	3.97**	1.88**	1.08	0.73	1.17	0.61*	0.29**
	MuRF3	0.30**	0.23**	0.22**	0.13**	0.48**	0.23**	2.24**	1.74**	0.72**	0.56**	1.21	0.75*	0.59**
	Znf216	0.84	0.65**	0.32**	0.32**	0.57**	0.60**	1.76**	1.89**	1.05	0.82	0.69	0.67*	0.72
Autophagy	atg4b	0.38**	0.18**	0.10**	0.08**	0.28**	0.12**	2.73**	1.63**	1.45*	0.70	0.78	0.56*	0.34**
	atg12l	0.95	0.75*	0.49**	0.51**	0.67**	0.66**	1.37*	1.30	1.15	0.92	0.64*	0.66**	0.62**
	SQSTM1	0.65	0.70*	0.33**	0.21**	0.43**	0.52*	1.30	2.48**	0.78*	0.84	0.57	0.36	0.69*
	Mul1	0.98	0.81*	0.49**	0.68**	0.47**	0.77	0.96	1.13	1.16	0.95	0.58*	0.80	0.95
	Bnip3	0.42	0.28**	0.20**	0.19**	0.55**	0.33**	2.71**	1.75**	1.03	0.67*	0.82	0.76*	0.49**
Fatty acid catabolism	HOAD	1.21	0.98	1.32*	1.49**	1.31	0.93	0.99	0.63**	1.13*	0.92	1.25**	1.41**	0.89
	CPT1A	0.62**	0.44**	1.02	1.76*	1.50	1.26	1.47	0.72	2.26**	1.59*	1.19	2.04**	1.00
	CPT1B	0.53**	0.46**	0.91	3.11**	1.02	1.08	1.12	0.35**	1.31	1.13	1.56*	5.32**	1.66**
Amino acid catabolism	GDH1	1.17	0.96	1.10	1.08	1.33**	0.80*	1.21	0.74**	1.03	0.84	1.13*	1.12*	0.68
	GDH2	1.04	0.87	1.05	0.97	1.28*	0.75**	1.22	0.78*	1.02	0.85	1.10*	1.01	0.65
	GDH3	1.65**	1.30*	1.39**	1.82**	1.44**	1.16	1.04	0.64**	1.11	0.88	1.02	1.33**	0.82
	ASAT1	1.54**	1.42**	1.17	1.75**	1.31**	1.21	1.12	0.69**	0.97	0.90	0.93	1.38**	0.85
	ASAT2	1.23*	1.05	0.92	1.26	1.20	0.97	1.31**	0.77*	0.97	0.83	0.93	1.27	0.75
	ALAT	1.16	0.82	1.13	1.37**	1.57**	0.96	1.38**	0.70**	1.26	0.89	1.33	1.61**	0.81

Discussion

This experimental study is the first to describe the influence of temperature and food depletion on mRNA levels of genes involved in the nutritional status of brown trout at the time of the first exogenous feeding. The transition from yolk reserves to exogenous feeding is a critical period for many fish, and may lead to high mortalities (Kennedy et al., 2008). Our data have clearly shown that in case of starvation, genes involved in autophagy and proteasome were highly expressed and it was boosted at 11°C. Surprisingly, the expression of genes coding for fatty acid and amino acid catabolism was not higher during starvation at 11°C (Table S2). These results are counter-intuitive and suggest that the regulation of these catabolic pathways may occur at different levels (*e.g.* post-transcriptional regulation; Salem et al. 2007), or that some other mechanisms such as hypometabolism are at work. This study has also demonstrated that young brown trout were able to feed and grow after 5 days of starvation. However, after this short period of starvation, mortalities were recorded at 11°C but not at 8°C, suggesting that in the wild, recovery would likely be uncertain at this temperature, even if food availability was restored. Finally, results demonstrated that at 8°C, food was more efficiently converted to tissue than at 11°C.

Energetic stress indicators: autophagy and proteasomal pathways at a molecular level

During nutrient restriction, metabolism changes to provide cellular energy *via* catabolic processes. Autophagy and proteasome are induced to mobilise energy and most often occur when essential nutrients are limited (Wing et al., 1995; Mizushima, 2007). To provide metabolic fuel, autophagy degrades all potential sources of energy (proteins, lipids and glycogen) and proteasome degrades skeletal-muscle proteins. In this regard, an induction of the expression of genes involved in both pathways can indicate energetic stress. The mRNA levels were greater in fish that were starved for 5 days and these differences were exacerbated at 9 days, highlighting the metabolic distress of starving fish. Consequences of starvation were still noticeable in delayed-fed fish although food input drove the majority of them back to a normal state. These results confirm that both pathways (autophagy and proteasome) are regulated by the feeding status (Robinowitz & White, 2010; Seiliez et al., 2010). Moreover, temperature exacerbates stress induced by starvation probably in relation with a higher standard metabolic rate (Régnier et al., 2013). More energy is needed to maintain vital functions and body reserves of alevins are probably rapidly depleted, which would explain a higher induction of these

catabolic pathways at 11°C, while recovery was not boosted at a sufficient rate to compensate when food was available again.

Recovery from delayed first exogenous feeding

Introduced by Blaxter & Hempel (1963), the “point-of-no-return” (PNR) is the threshold after which fish are still alive but too weak to feed. At 8 °C, no delayed-fed fish died and they were in an intermediate state, having gained weight when compared with starved fish, but exhibiting lower performance than fed fish. PNR value is largely dependent on temperature (McGurk, 1984; Dou et al., 2002). At 11°C, starvation over a 5-day period was an intense stress and all fish were not able to recover. At D₉, for delayed-fed survivors, results were mixed. Some gained weight, but some still had the same weight loss levels than starved fish, suggesting that they reached the PNR and would not be able to recover. Fish can exhibit compensatory growth after a period of food deprivation (Nicieza & Metcalfe, 1997), but the duration of the present experiment was probably too short to detect such a phenomenon. However, there were no evidences from prey consumption rate and catabolism that a catch-up growth may arise.

Feed utilisation at different temperatures

Because brown trout are ectotherms, their growth is linked to temperature. The best energy conversion efficiency into growth for *Salmo trutta* is around 9-10°C (Marr, 1966; Blaxter, 1969; Elliott & Hurley, 2001). Our results demonstrated a better food conversion in growth efficiency at 8°C when compared to 11°C (similar weight gain and catabolism, even though feed intake was higher at 11°C). In addition, length gain was higher at 8°C. Similar results for another salmonid were found by Malzahn et al. (2003) who highlighted hyperplasia phenomenon leading to longer coregonid fish in colder water.

In the context of CC, an increase of 3.2°C in air temperature will produce a moderate but sensible increase in stream water temperature from 1 to 2°C (Bal et al., 2014). Present results suggest that this may cause higher mortality rates during the critical period of emergence, especially when in conjunction with food shortage which is not a scarce event in the wild (Kennedy et al. 2008). Besides its effect on water temperature, CC will affect flow through changes in precipitation patterns. In winter and spring, higher flows will likely magnify the effect of temperature by reducing the availability of invertebrates both through habitat reduction (Kennedy et al. op. cit.) and invertebrate washout.

Acknowledgements

No competing interests declared. E. Arevalo benefits from a cross-border grant (Univ Pau & Pays Adour/UPV) and from the French Embassy mobility helping program (Mérimee). Experiments were carried out thanks to the IE ECP facilities and financially supported by CG64 and INRA Ecoserv Metaprogram. F. Gueraud, P. Coste, E. Huchet, S. Glise, F. Lange, E. Plagnes-Juan and J. Rives helped in the experimental setting and the laboratory experiment. We thank J. Almany for correcting the English manuscript.

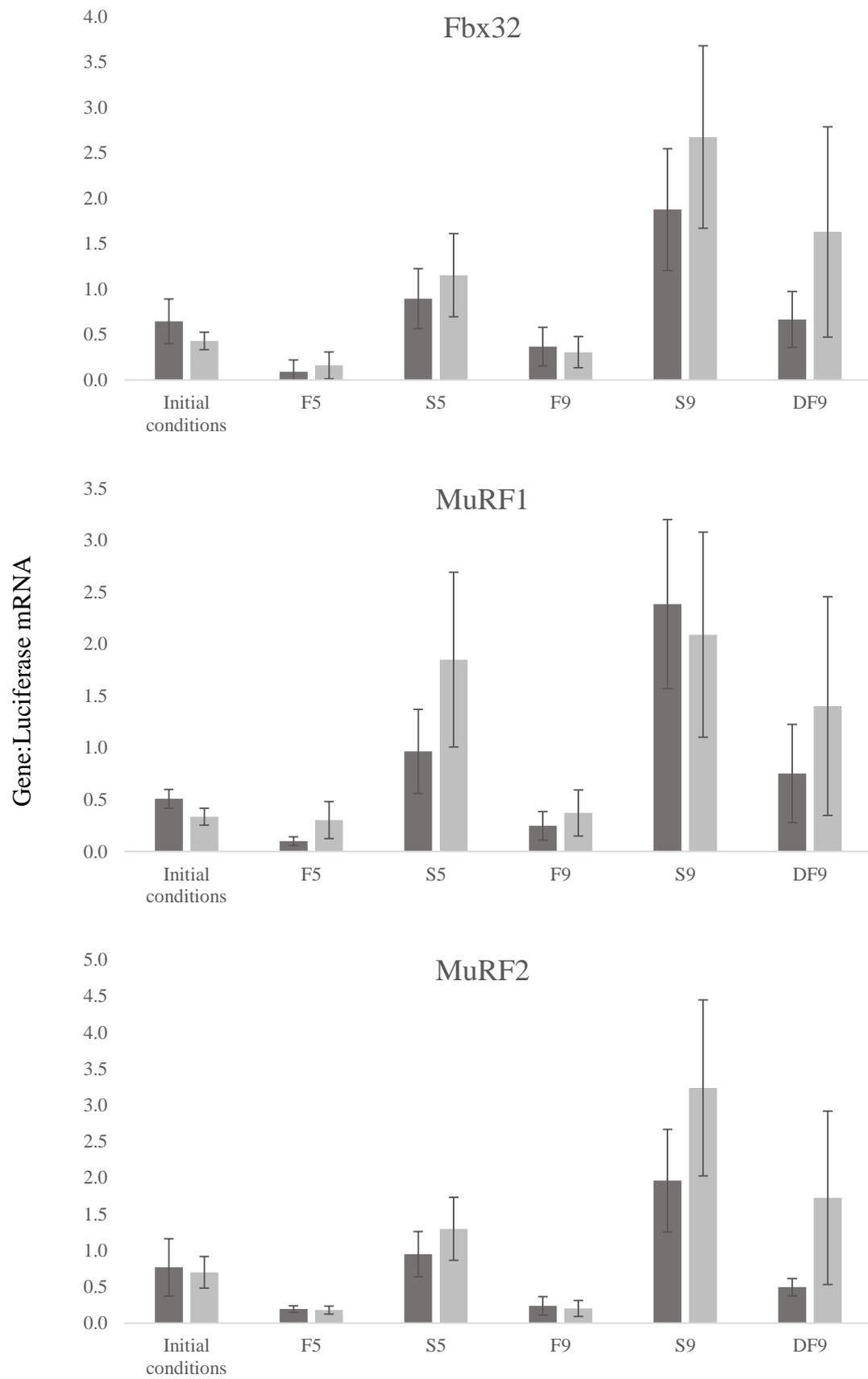
Supporting Information

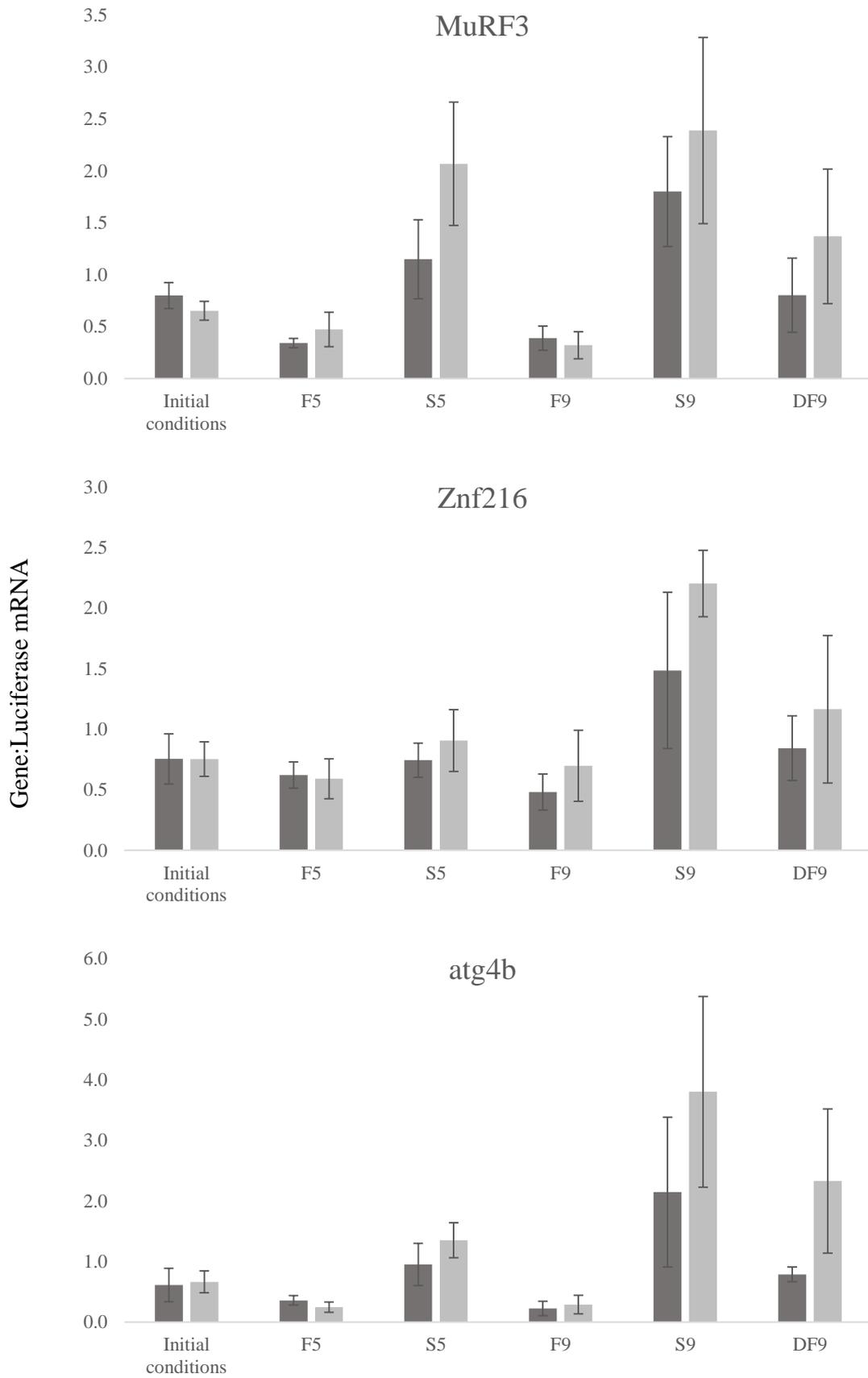
Table S1: Primers used for real time RT-qPCR analysis.

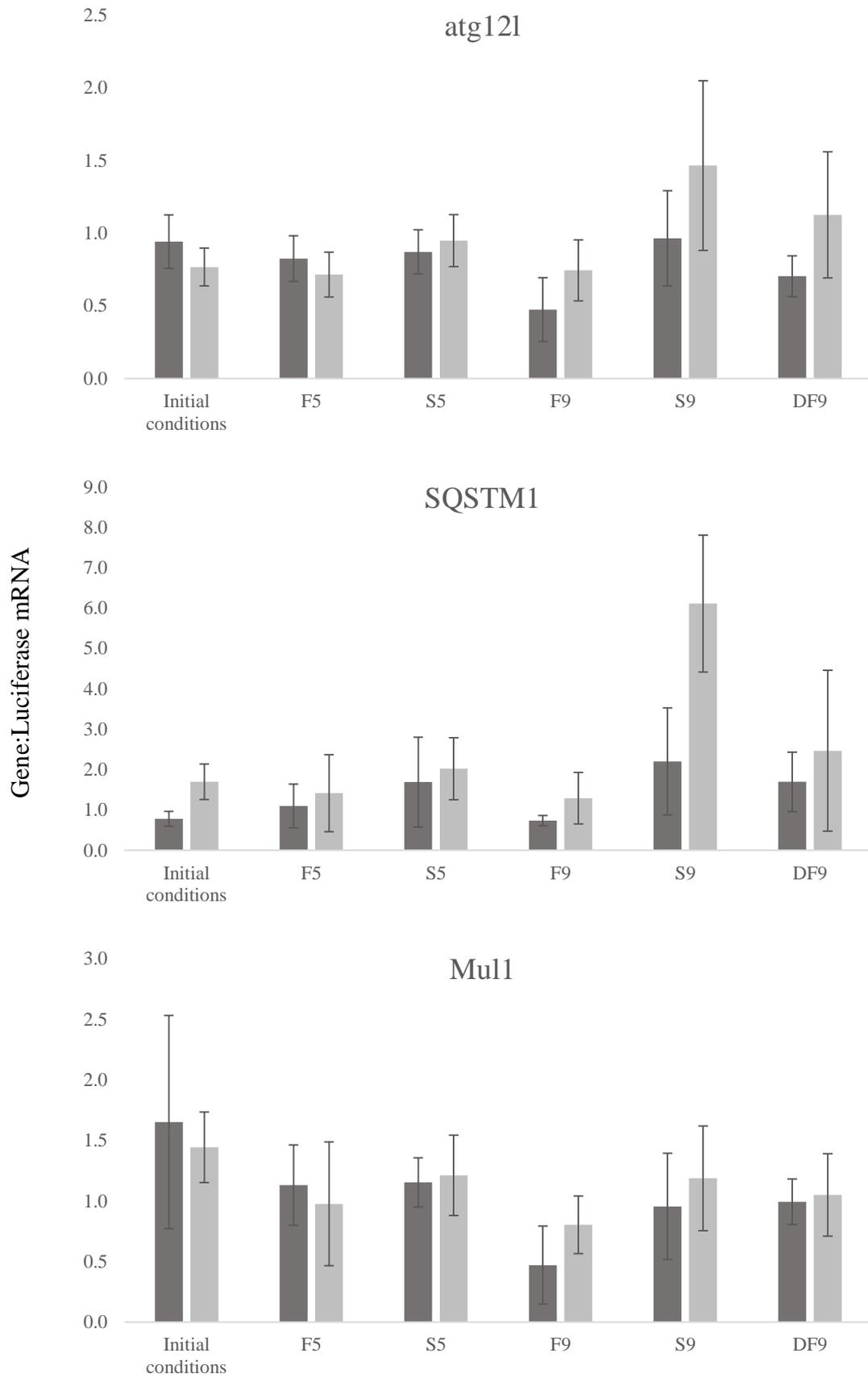
Pathways	Genes	Forward primer	Reverse primer
Proteasome	Fbx32	5'-TGCGATCAAATGGATTCAAA-3'	5'-GATTGCATCATTTCCCCACT-3'
	MuRF1	5'-CTGATTAGTGGCAAGGAGCTG-3'	5'-GTAAGGTGCTCCATGTTCTCG-3'
	MuRF2	5'-TGGAGGAGTCAGAGATGGCTA-3'	5'-TCCAGGTGGGAGATGTTAGTG-3'
	MuRF3	5'-ATGTCCATTGCAGGGACTCTA-3'	5'-AACTGGGGTAAGCCATTGTGT-3'
	Znf216	5'-AAGAGGGTGGGCCTCACAG-3'	5'-GACATCCTTTTGCCACTCGT-3'
Autophagy	atg4b	5'-TATGCGCTCCGAAAGTTGTC-3'	5'-CAGGATCGTTGGGGTTCTGC-3'
	atg12l	5'-GATGGAGGCCAATGAACAGC-3'	5'-GCGTTTGAAGTAAAAGGGCTAA-3'
	SQSTM1	5'-AGCCCACTGGGTATCGATGT-3'	5'-GGTCACGTGAGTCCATTCCT-3'
	Mul1	5'-CCACGAGATGGAGGAGATGT-3'	5'-AGAGCGTTGTGGAAGCAACT-3'
	Bnip3	5'-CCTGTGACAGTCCTCCGAGA-3'	5'-CCACTTCACGTCTCCGTTCT-3'
Fatty acid catabolism	HOAD	5'-GGACAAAGTGGCACCAGCAC-3'	5'-GGGACGGGGTTGAAGAAGTG-3'
	CPT1A	5'-TCGATTTTCAAGGGTCTTCG-3'	5'-CACAACGATCAGCAAAGTGG-3'
	CPT1B	5'-CCCTAAGCAAAAAGGGTCTTCA-3'	5'-CATGATGTCACTCCCGACAG-3'
Amino acid catabolism	GDH1	5'-AACTCCGCAGCGTCTCTTTCCCCAT-3'	5'-TCACCTCATCAACAGACACCTCTTCA-3'
	GDH2	5'-ATCAAGCCCTGCAACCACGTCCT-3'	5'-TCTTCACTGTAACGGATCCCCCTTT-3'
	GDH3	5'-CTGCAACCATATACTGAGTGTATCGTTCC-3'	5'-ATGTCATCAGCGAGGCCAGGGCTTT-3'
	ASAT1	5'-TCAAGAGTGGCAGGAACATCA-3'	5'-AGCGTCTCTGAAGATGGGTGT-3'
	ASAT2	5'-TCTGTGCCAGTCCTTCTC-3'	5'-GGAGGGTTGGACCAGGT-3'
	ALAT	5'-TGGGTGCGTACAGTGCCAGT-3'	5'-GACGCACCCTCACCACACAC-3'

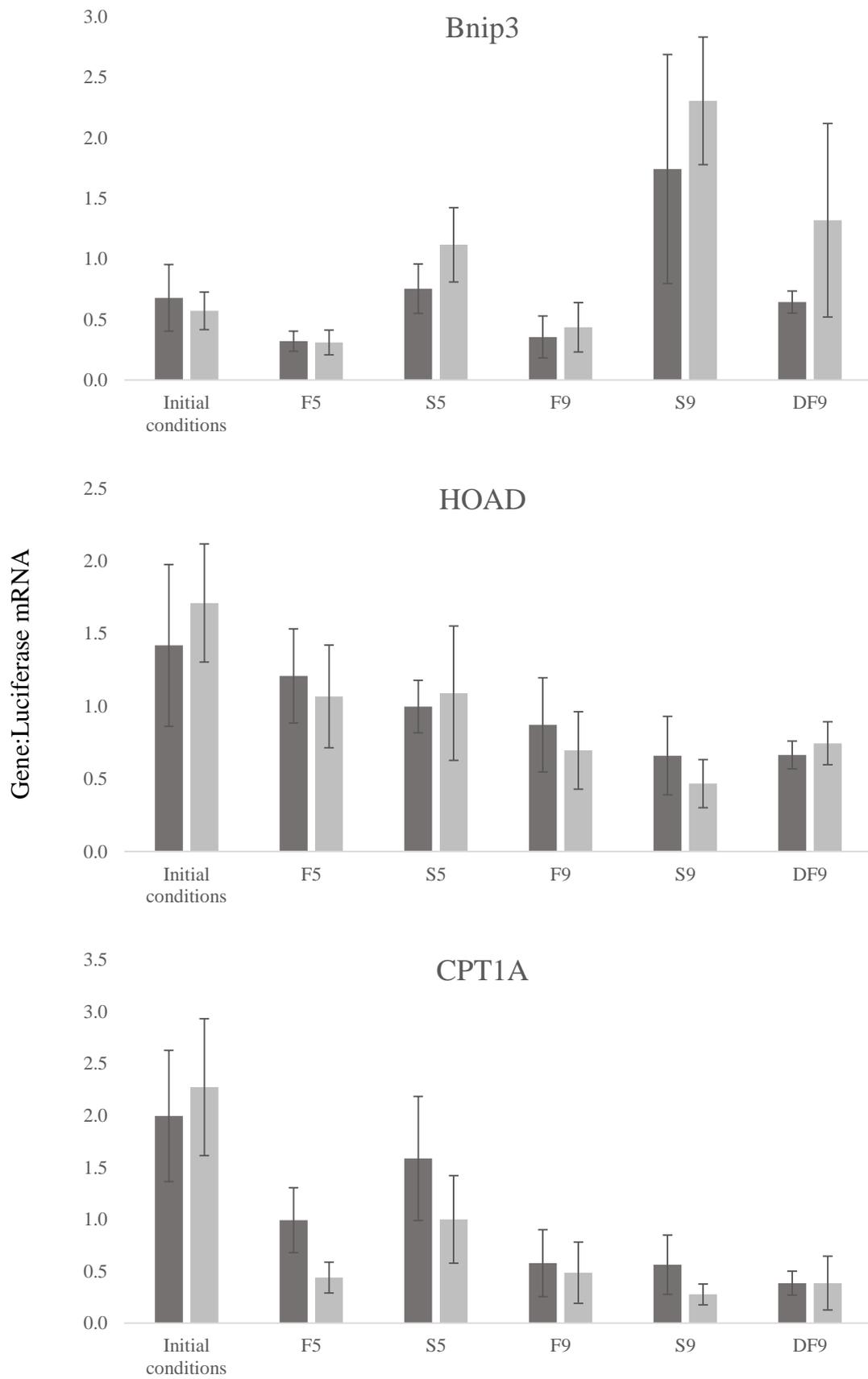
Table S2: Mean and standard deviation of mRNA levels of genes involved in fatty acid/amino-acid catabolism and proteasome/autophagy pathways. From yolk sac exhaustion, 6 fish per condition were sampled at the beginning (D_0), after 5 days (F_5 , S_5) and after 9 days (F_9 , S_9 , DF_9) of growth at 8°C and at 11°C. Gene expression level was normalized by the abundance of exogenous luciferase RNA and has no unit. The mean low expressions are in white, intermediate expressions in light grey and high expression in dark grey.

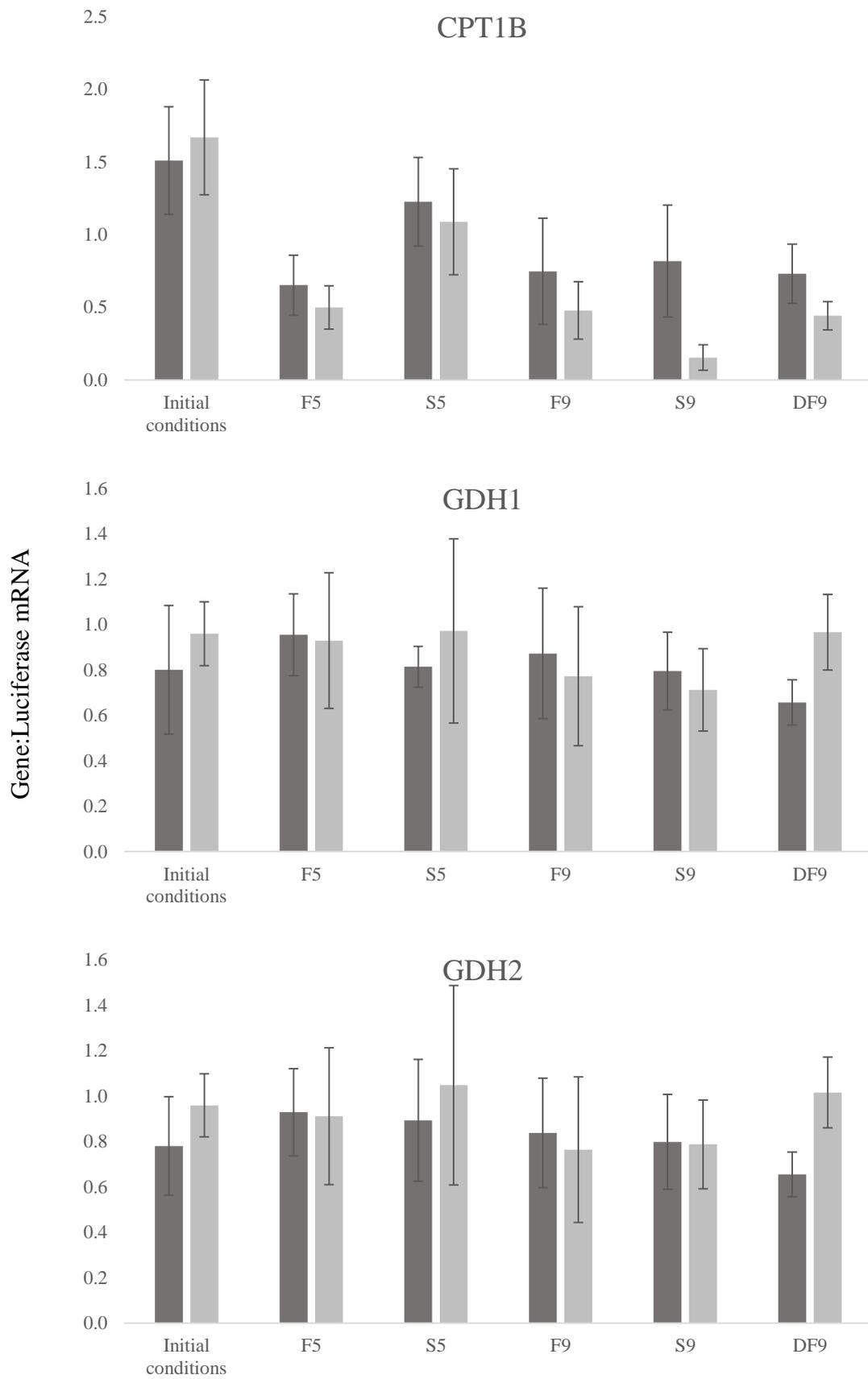
Pathways	Genes	Initial Conditions - D_0				F_5				F_9				S_5				S_9				DF_9			
		8°C		11°C		8°C		11°C		8°C		11°C		8°C		11°C		8°C		11°C		8°C		11°C	
		Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Proteasome	Fbx32	0.646	0.24	0.430	0.10	0.089	0.13	0.160	0.15	0.367	0.21	0.306	0.17	0.896	0.33	1.153	0.46	1.877	0.67	2.675	1.00	0.666	0.31	1.630	1.16
	MuRF1	0.508	0.09	0.335	0.08	0.101	0.04	0.302	0.18	0.247	0.14	0.371	0.22	0.964	0.40	1.848	0.84	2.384	0.81	2.088	0.99	0.752	0.47	1.401	1.05
	MuRF2	0.767	0.39	0.698	0.22	0.192	0.05	0.178	0.06	0.236	0.13	0.202	0.11	0.949	0.31	1.297	0.43	1.960	0.71	3.237	1.21	0.493	0.12	1.725	1.19
	MuRF3	0.799	0.12	0.652	0.09	0.341	0.04	0.472	0.17	0.389	0.12	0.321	0.13	1.149	0.38	2.067	0.59	1.801	0.53	2.388	0.90	0.803	0.36	1.369	0.65
	Znf216	0.754	0.21	0.753	0.14	0.621	0.11	0.591	0.17	0.480	0.15	0.697	0.29	0.744	0.14	0.905	0.25	1.485	0.65	2.203	0.27	0.843	0.27	1.164	0.61
Autophagy	atg4b	0.610	0.28	0.664	0.18	0.357	0.08	0.245	0.09	0.224	0.12	0.286	0.15	0.951	0.35	1.350	0.29	2.146	1.24	3.801	1.58	0.787	0.12	2.328	1.19
	atg12l	0.941	0.18	0.767	0.13	0.825	0.16	0.714	0.15	0.474	0.22	0.744	0.21	0.872	0.15	0.949	0.18	0.965	0.33	1.465	0.58	0.704	0.14	1.126	0.43
	SQSTM1	0.779	0.19	1.698	0.44	1.098	0.54	1.413	0.95	0.735	0.13	1.289	0.64	1.689	1.11	2.021	0.77	2.202	1.33	6.108	1.70	1.694	0.74	2.465	1.99
	Mul1	1.651	0.88	1.443	0.29	1.131	0.33	0.976	0.51	0.470	0.32	0.803	0.24	1.153	0.20	1.211	0.33	0.954	0.44	1.187	0.43	0.992	0.19	1.049	0.34
	Bnip3	0.678	0.27	0.570	0.15	0.320	0.08	0.310	0.10	0.355	0.17	0.435	0.20	0.754	0.20	1.117	0.31	1.742	0.95	2.305	0.53	0.644	0.09	1.319	0.80
Fatty acid catabolism	HOAD	1.418	0.56	1.710	0.41	1.208	0.32	1.067	0.35	0.872	0.32	0.696	0.27	0.997	0.18	1.089	0.46	0.660	0.27	0.467	0.17	0.665	0.10	0.745	0.15
	CPT1A	1.995	0.63	2.273	0.66	0.991	0.31	0.438	0.15	0.576	0.32	0.486	0.29	1.585	0.60	0.999	0.42	0.562	0.28	0.276	0.10	0.384	0.12	0.384	0.26
	CPT1B	1.510	0.37	1.670	0.40	0.652	0.21	0.499	0.15	0.747	0.37	0.478	0.20	1.228	0.30	1.089	0.36	0.818	0.38	0.154	0.09	0.732	0.20	0.442	0.10
Amino acid catabolism	GDH1	0.801	0.28	0.959	0.14	0.955	0.18	0.929	0.30	0.872	0.29	0.772	0.31	0.814	0.09	0.972	0.41	0.794	0.17	0.712	0.18	0.656	0.10	0.966	0.17
	GDH2	0.780	0.22	0.959	0.14	0.929	0.19	0.911	0.30	0.837	0.24	0.764	0.32	0.893	0.27	1.048	0.44	0.798	0.21	0.787	0.20	0.655	0.10	1.015	0.16
	GDH3	1.015	0.22	1.209	0.20	1.318	0.14	1.185	0.29	0.945	0.36	0.928	0.42	0.799	0.09	0.913	0.29	0.678	0.18	0.510	0.20	0.655	0.18	0.799	0.13
	ASAT1	0.921	0.24	1.017	0.13	1.163	0.21	1.199	0.30	0.898	0.35	0.970	0.40	0.757	0.08	0.845	0.32	0.766	0.20	0.554	0.11	0.685	0.12	0.803	0.08
	ASAT2	0.894	0.29	0.934	0.18	0.955	0.23	0.983	0.30	0.754	0.24	0.811	0.27	0.776	0.09	0.938	0.43	0.822	0.24	0.644	0.15	0.626	0.10	0.838	0.12
	ALAT	0.963	0.27	0.892	0.38	1.030	0.12	0.820	0.21	0.871	0.25	0.654	0.23	0.885	0.08	0.996	0.27	0.768	0.10	0.477	0.26	0.555	0.09	0.684	0.08

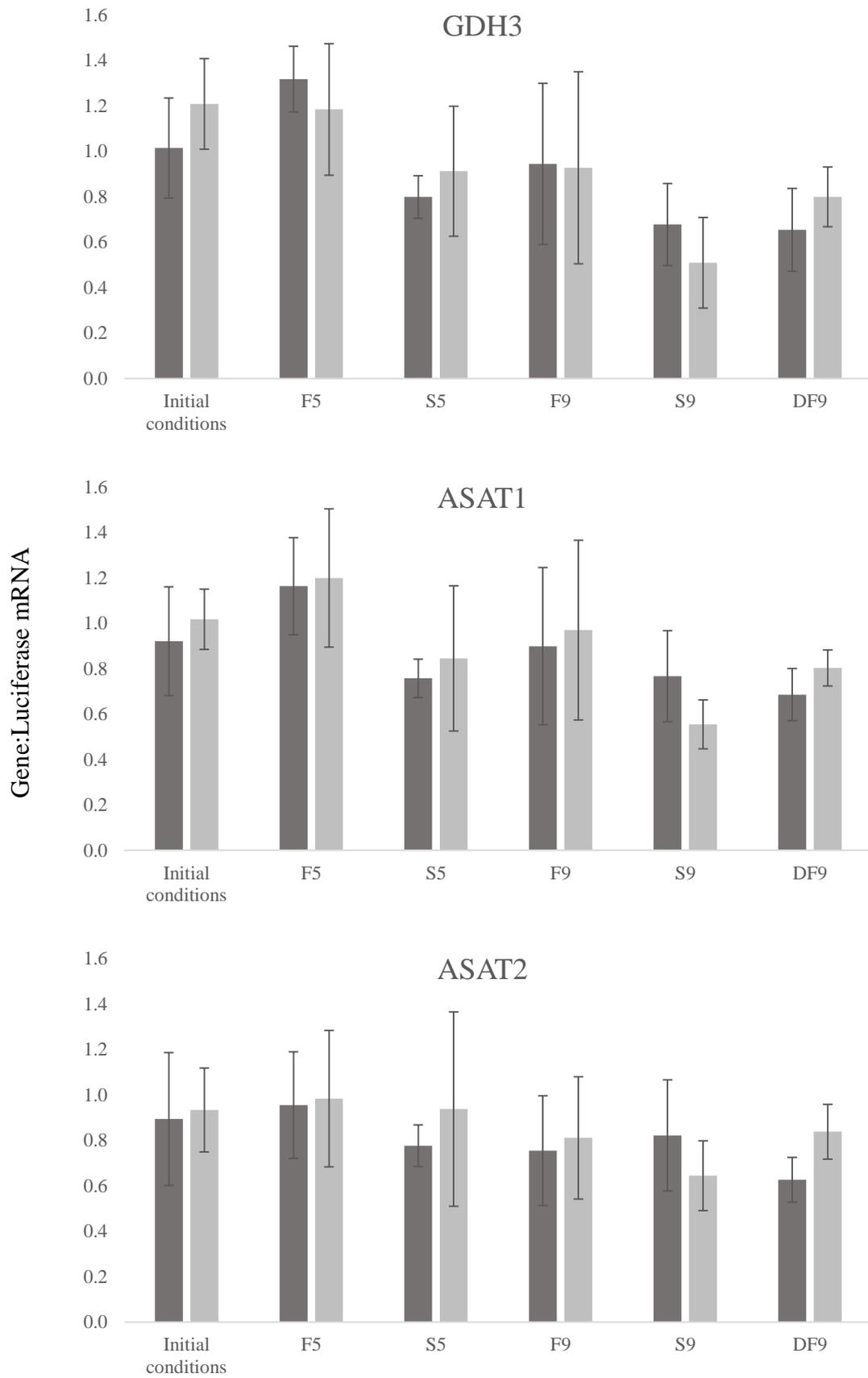












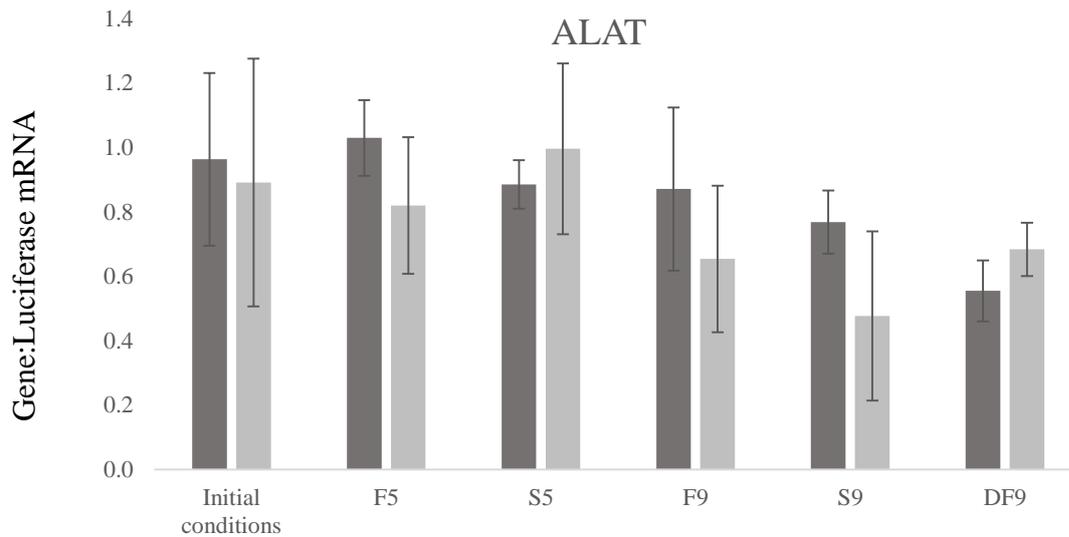


Figure S1 Mean and standard deviation of mRNA levels of genes involved in proteasome (Fbx32, MuRF1, MuRF2, MuRF3, Znf216), autophagy (atg4b, atg12l, SQSTM1, Mul1, Bnip3) pathways, fatty acid (HOAD, CPT1A, CPT1B) and amino-acid (GDH1, GDH2, GDH3, ASAT1, ASAT2, ALAT) catabolism. From yolk sac exhaustion, 6 fish per condition were sampled at the beginning (Initial conditions), after 5 days (F₅, S₅) and after 9 days (F₉, S₉, DF₉) of growth at 8°C (dark grey) and at 11°C (light grey). Gene expression level was normalized by the abundance of exogenous luciferase RNA.

Table 2.1 Synthesis of the main results on the growth and metabolism of starved and delayed first-feeding alevins at 8 and 11°C.

Temperature	Starvation		Delayed First-feeding
8°C	↘ Growth	Biometric data	Recovery
	Similar to F & DF fish <i>Body reserves still available?</i>	Lipid & AA catabolism	Similar to F fish
	+++	Autophagy & Proteasome	+
11°C	↘↘ Growth	Biometric data	Contrasted recovery
	↘↘ Gene expression <i>Reserves depleted?</i> <i>Hypometabolism?</i>	Lipid & AA catabolism	Intermediate state
	++++	Autophagy & Proteasome	++

Further considerations and discussion

In the present paper, analyses were focused on averages of mRNA levels according to conditions, while variance was not commented as an informative factor by itself because of the paper format chosen (brief communication). Because I thought it might brought interesting developments, I include this analysis here. After 9 days of experiment, standard deviation values of mRNA levels of genes coding for fatty acid and amino acid catabolism ranged from 0.09 to 0.42, while standard deviations of genes involved in proteasome and autophagy ranged from 0.11 to 1.99 (Figure 2.2). The important standard deviations of genes involved in proteasome and autophagy indicate that individual response to the treatment varied a lot. Considering weight as a revealer of stress induced by starvation (the individuals suffering most from starving being those losing more weight), we tested if the induction of these genes correlated with the weight. We hypothesised that the alevins suffering most from starvation should be those who had lost more mass and that they should be the individuals in which the expression of genes involved in proteasome and autophagy was maximized.

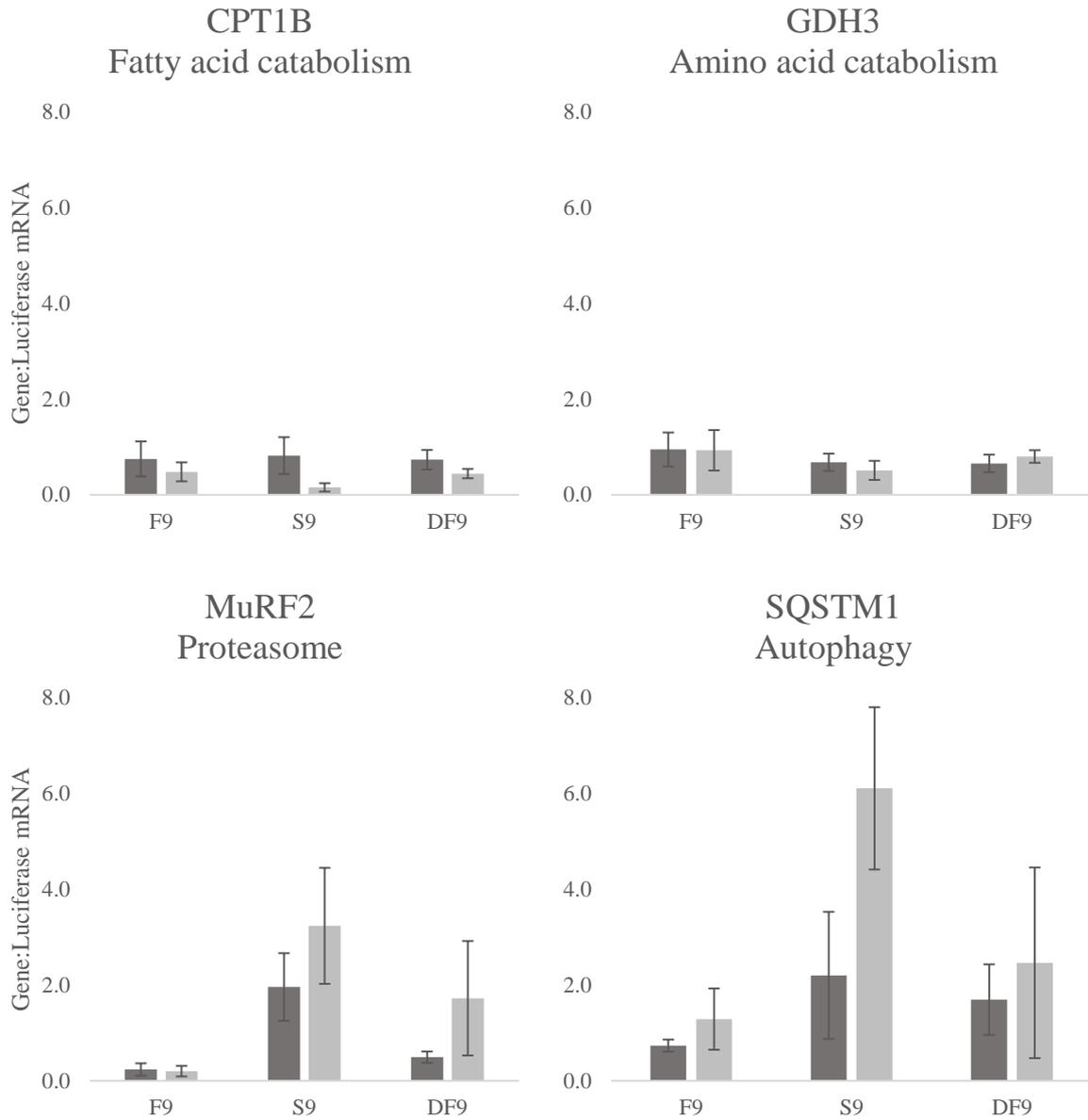


Figure 2.2 Mean mRNA levels of genes involved in fatty acid (*CPT1B*), amino acid catabolism (*GDH3*), proteasome (*MuRF2*) and autophagy (*SQSTM1*) according to the treatment (Fed F_9 , Starved S_9 or Delayed-First feeding DF_9) after 9 days of experiment. mRNA levels were normalized by the abundance of exogenous luciferase RNA. Bars represent standard deviation. Alevins reared at 8°C are represented in dark grey and those reared at 11°C are in light grey.

Considering only data of the 9th day, correlations were tested with OpenBUGS® with estimations of coefficients for each j condition (F_9 , S_9 and DF_9). If X_i is the mRNA level of the i^{th} individual, we assumed:

$$X_i = a_{j[i]} Wg_i + b_{j[i]}$$

With $a_{j[i]}$ the slope coefficient and $b_{j[i]}$ the intercept, both estimated according to j conditions. Individual weight gain (WG_i) was calculated as follows:

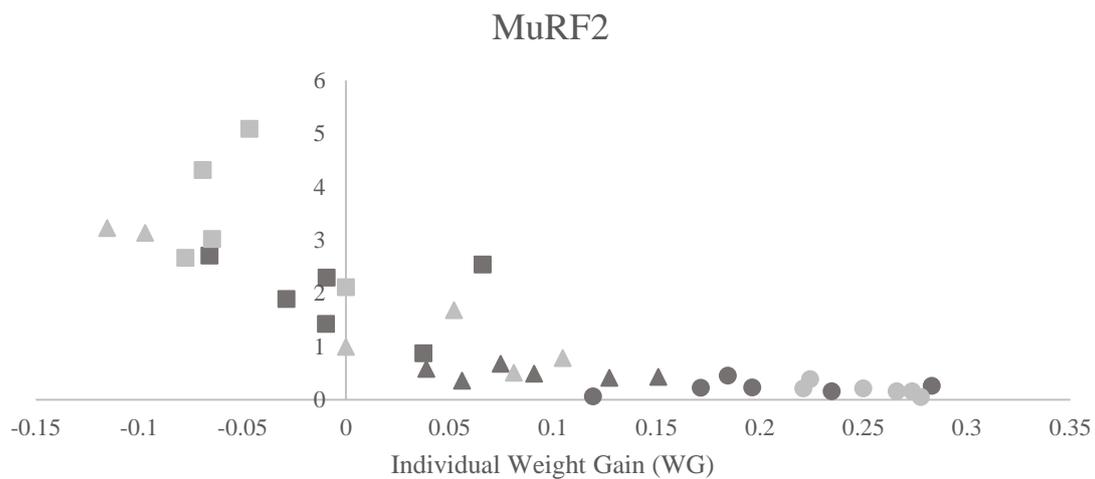
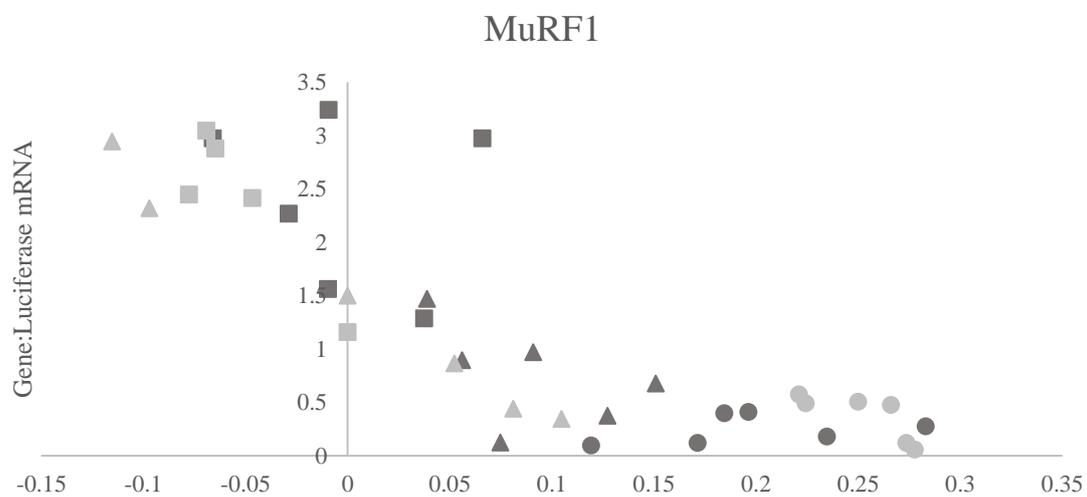
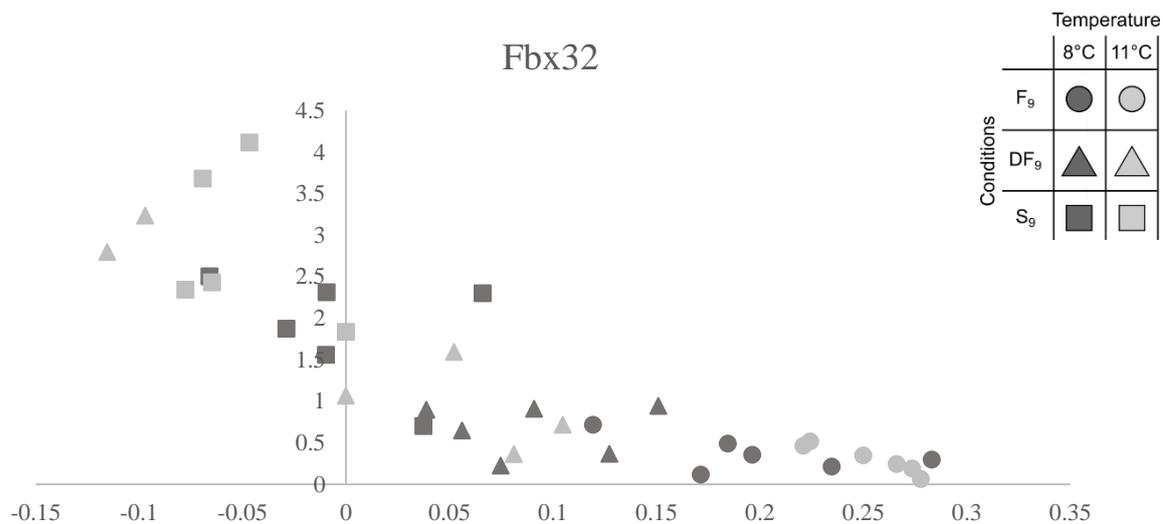
$$WG_i = \frac{Final\ Weight_i - Initial\ Weight_i}{Initial\ Weight_i}$$

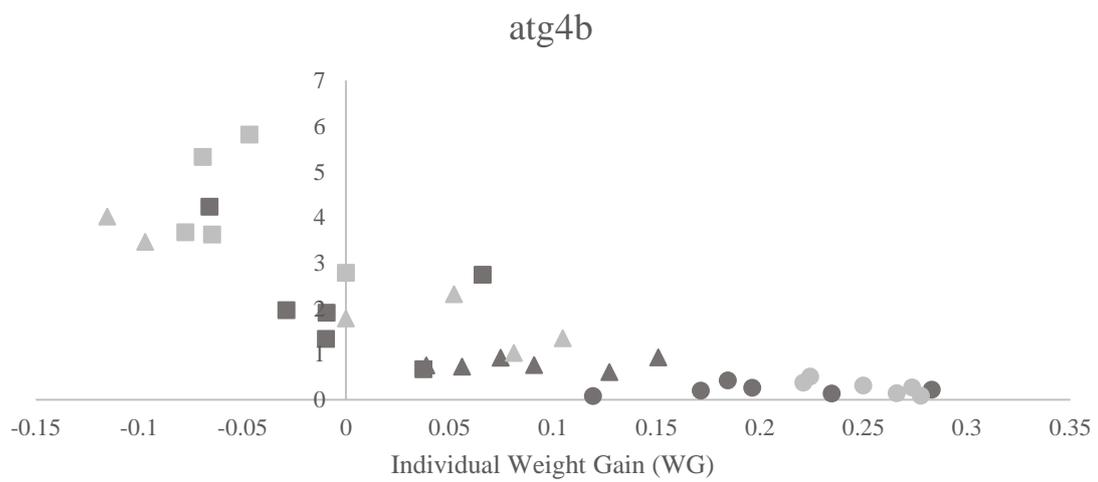
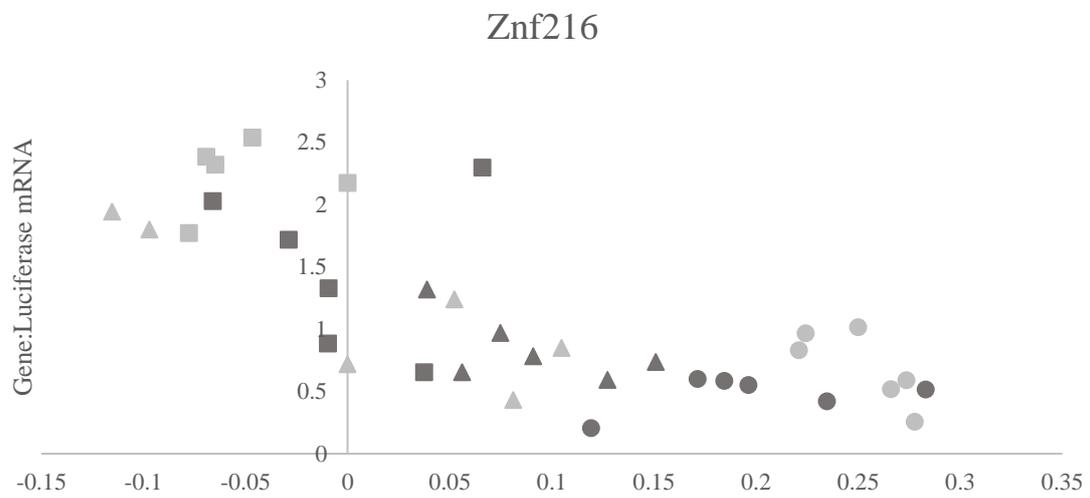
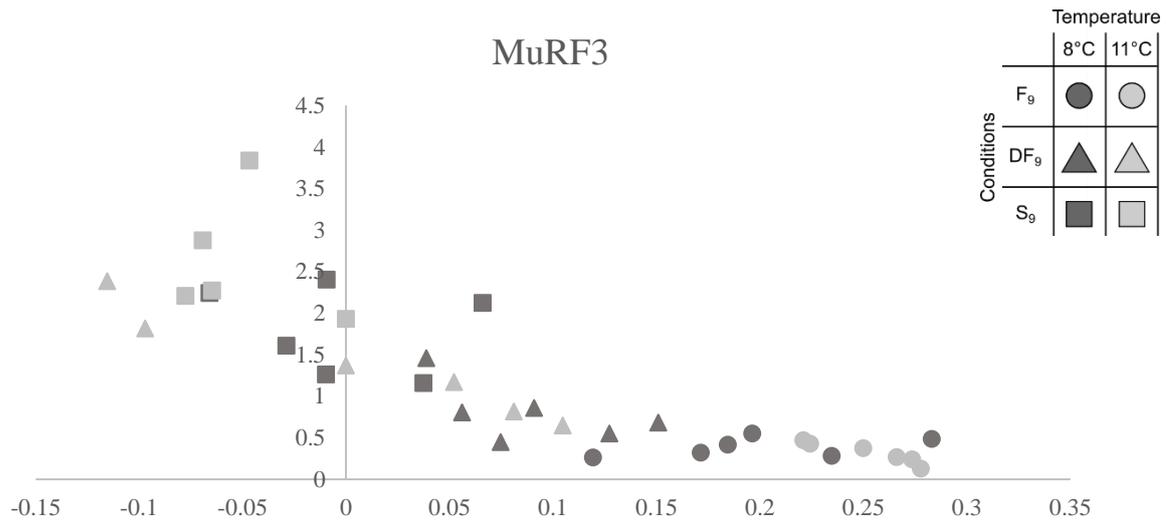
Fit of the correlation was calculated by dividing the variability unexplained by the correlation model with the variability observed in the data set as follows:

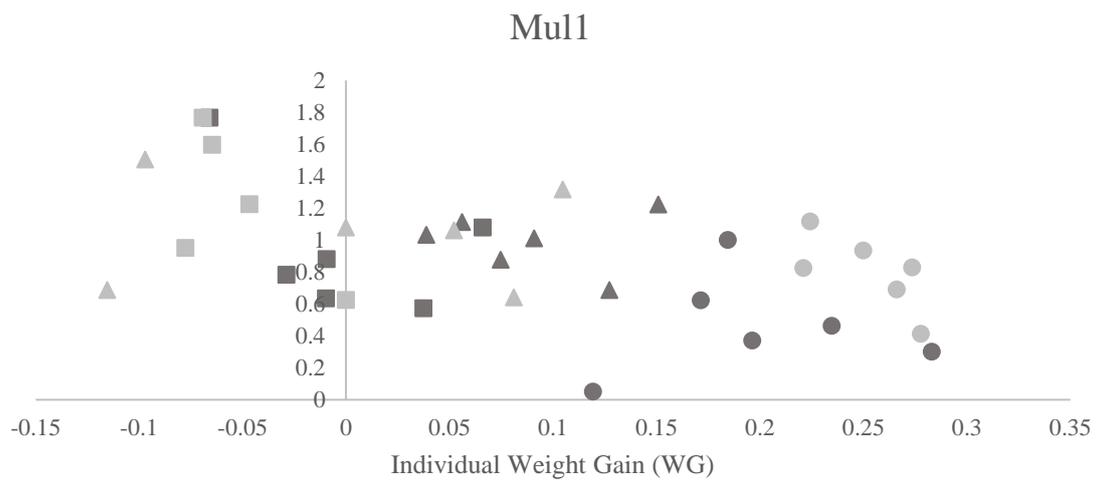
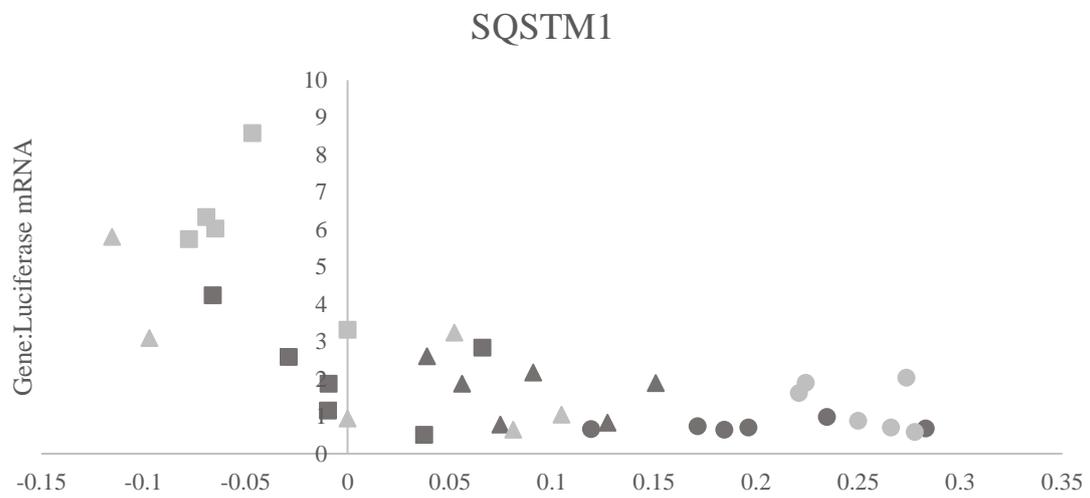
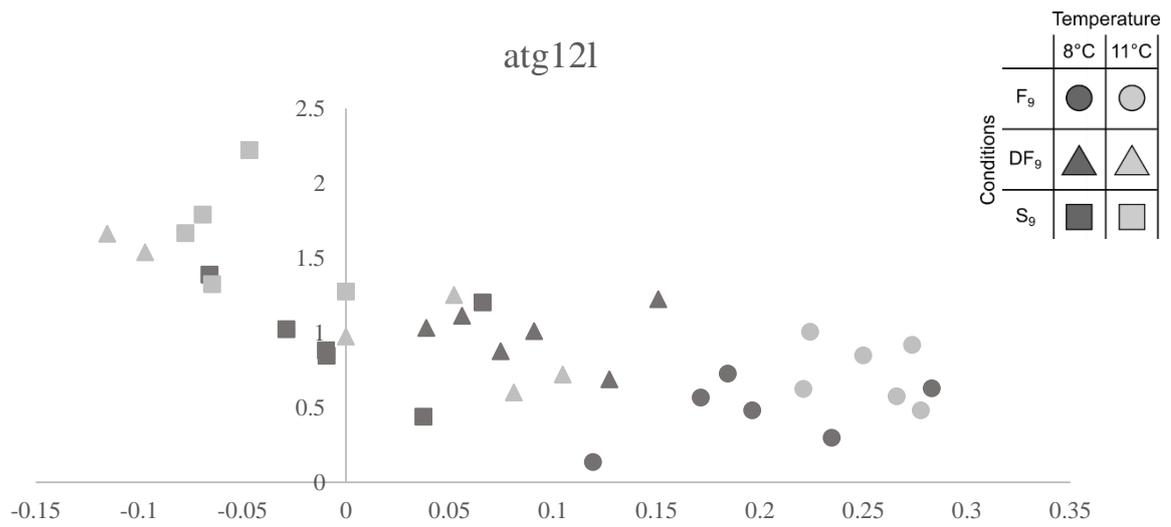
$$Fit = 1 - \frac{Variability\ unexplained\ by\ the\ model}{Variability\ of\ the\ data\ set}$$

When the fit was between 0.5 and 1, the correlation between mRNA levels and WG was good. When the fit was between 0 and 0.5, the correlation was intermediate and when the fit is negative, there was no correlation. For each condition (fed F₉, starved S₉ and delayed first-feeding DF₉), we tried to link individual weight gain with the expression of the autophagy and proteasome related genes. The fit represented the strength of the relationship, then a represented the direction of the relationship and b the intercept. A total of 30 relationships were tested (3 conditions x 10 genes).

For fed fish (F₉), considering 10 correlations between individual weight gain and mRNA levels, 8 fits were negative (Table 2.2). The majority of the parameters (7/10 intercepts and 8/10 slope coefficients) were equal to 0. This means that mRNA levels of genes involved in these pathways were very low and no relationship between weight gain (WG) and mRNA levels was observed. For starved fish (S₉), fits were intermediate, ranging from 0 to 0.30. 7 slope coefficients (a) were significantly lower than 0. For delayed first-feeding fish (DF₉), 9 fits were good, over 0.44 (except Mul1). 9 slope coefficients (a) were significantly lower than 0. It appeared that when fish face periods of food deprivation (S₉ and DF₉), the more they lost weight, the more genes involved in proteasome and autophagy were expressed. Moreover, whatever the gene considered, F₉ and S₉ constituted two distinct groups of individuals, while DF₉ made the link between F₉ and S₉ (Figure 2.3).







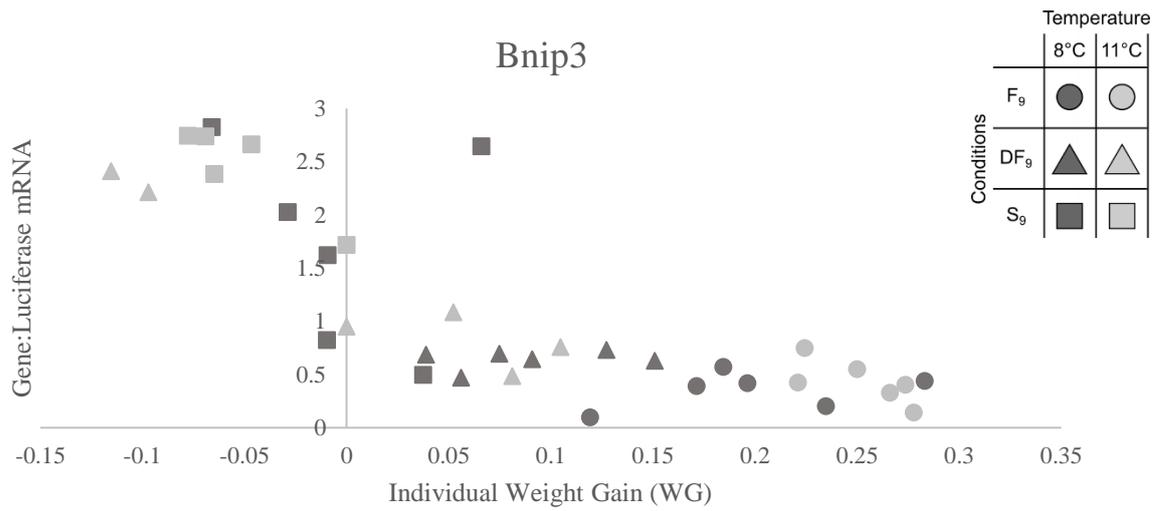


Figure 2.3 mRNA levels according to Individual Weight Gain for genes involved in proteasome (Fbx32, MuRF1, MuRF2, MuRF3 and Znf216) and autophagy (atg4b, atg12l, SQSTM1, Mull, Bnip3) pathways. Dark items represent alevins reared at 8°C and light items represent alevins reared at 11°C. Fed fish F₉ are in circles, delayed-fed fish DF₉ are in triangle and starved fish S₉ are in square.

Table 2.2 Correlations between mRNA levels (Y) and individual weight gain (Wg ; X) of alevins, mean of posterior probability distribution functions of a (the slope coefficient), b (the intercept) and fit of the models. Parameters were considered significant when $P(X > 0)$ is less than 0.10 or above 0.90 (in bold).

Conditions	Pathways	Genes	a	P(a > 0)	b	P(b > 0)	Equations	Fit
F ₉	Proteasome	Fbx32	-2.33	0.02	0.86	1.00	Fbx32 = -2.329 Wg + 0.8606	0.29
		MuRF1	0.40	0.63	0.22	0.77	MuRF1 = 0.402 Wg + 0.2188	-0.15
		MuRF2	-0.22	0.39	0.27	0.92	MuRF2 = -0.2178 Wg + 0.2678	-0.16
		MuRF3	-0.42	0.31	0.45	0.98	MuRF3 = -0.4177 Wg + 0.4493	-0.14
		Znf216	0.86	0.70	0.39	0.85	Znf216 = 0.8641 Wg + 0.3935	-0.12
	Autophagy	atg4b	-0.02	0.49	0.26	0.88	atg4b = -0.01875 Wg + 0.2598	-0.16
		atg12l	2.19	0.91	0.12	0.63	atg12l = 2.187 Wg + 0.1172	0.08
		SQSTM1	2.14	0.75	0.53	0.76	SQSTM1 = 2.142 Wg + 0.5299	-0.08
		Mul1	1.41	0.75	0.32	0.75	Mul1 = 1.406 Wg + 0.3199	-0.09
		Bnip3	0.45	0.64	0.29	0.84	Bnip3 = 0.4489 Wg + 0.2939	-0.14
S ₉	Proteasome	Fbx32	-7.97	0.07	2.14	1.00	Fbx32 = -7.969 Wg + 2.136	0.18
		MuRF1	-4.29	0.18	2.28	1.00	MuRF1 = -4.287 Wg + 2.279	0.00
		MuRF2	-8.63	0.09	2.42	1.00	MuRF2 = -8.627 Wg + 2.422	0.17
		MuRF3	-5.71	0.11	2.03	1.00	MuRF3 = -5.711 Wg + 2.034	0.09
		Znf216	-4.12	0.16	1.73	1.00	Znf216 = -4.124 Wg + 1.725	0.01
	Autophagy	atg4b	-11.75	0.06	2.81	1.00	atg4b = -11.75 Wg + 2.808	0.26
		atg12l	-5.88	0.03	1.14	1.00	atg12l = -5.88 Wg + 1.137	0.27
		SQSTM1	-11.72	0.10	3.63	1.00	SQSTM1 = -11.72 Wg + 3.626	0.21
		Mul1	-5.40	0.03	0.95	1.00	Mul1 = -5.399 Wg + 0.9494	0.26
		Bnip3	-7.21	0.07	1.89	1.00	Bnip3 = -7.207 Wg + 1.888	0.18
DF ₉	Proteasome	Fbx32	-9.52	0.00	1.60	1.00	Fbx32 = -9.517 Wg + 1.596	0.70
		MuRF1	-9.25	0.00	1.51	1.00	MuRF1 = -9.253 Wg + 1.511	0.81
		MuRF2	-10.82	0.00	1.62	1.00	MuRF2 = -10.82 Wg + 1.616	0.77
		MuRF3	-6.45	0.00	1.39	1.00	MuRF3 = -6.451 Wg + 1.389	0.82
		Znf216	-4.78	0.00	1.23	1.00	Znf216 = -4.777 Wg + 1.227	0.64
	Autophagy	atg4b	-11.60	0.00	2.10	1.00	atg4b = -11.6 Wg + 2.102	0.74
		atg12l	-4.09	0.00	1.11	1.00	atg12l = -4.09 Wg + 1.107	0.76
		SQSTM1	-10.89	0.01	2.58	1.00	SQSTM1 = -10.89 Wg + 2.584	0.44
		Mul1	-0.31	0.38	1.04	1.00	Mul1 = -0.3092 Wg + 1.035	-0.15
		Bnip3	-6.89	0.00	1.31	1.00	Bnip3 = -6.891 Wg + 1.306	0.76

The results confirm that mobilisation of protein stored in muscles results in a loss of weight (Sandri 2010). Late access to food (DF₉) would induce a decrease in proteolytic activity for some alevins which would probably come back to a normal metabolic activity and survive, but the DF alevins with still high mRNA levels for these genes on the 9th day would probably die in the following hours/days. Further analyses measuring the expression of these genes on alevins in the wild could help to assess the starving situation of 0⁺ fish. Indeed, if it is possible to use them as markers of metabolic stress, they would be an indicator of the alevin access to trophic resources.

According to the results of the present experiment conducted in controlled environment, alevins can survive only a 9-day period to starvation once the yolk sac is exhausted and warmer temperature clearly intensifies the metabolic distress. Consequently, survival and growth of salmonid alevins would be threatened by Global Climate Change because of the increase in starvation risk. However, these results should be considered with caution and may be difficult to transpose to the wild due to additional factors such as the cost of swimming, the risk of predation and the impact of competition and territoriality that may also increase.

Chapter III Consequences of Global Climate Change on the community of invertebrates, on the survival, growth and behaviour of first-feeding alevins at high density: flood simulation in a semi-natural environment

Study context

Previous chapters investigated factors triggering entry rate in the drift of invertebrates and the response of alevins to periods of food deprivation. Both were carried out under laboratory conditions. This third chapter attempts to connect the response of invertebrates with the response of alevins following a flood. In the field, heterogeneity of the environment makes difficult to separate the effect of floods from others abiotic factors. To overcome this, we used an experimental river fed naturally where some abiotic factors can be controlled (flow rate, water levels, gravel bed and the community of vertebrates).

Manuscript in preparation

Assessment of Global Climate Change on the community of invertebrates, on the survival, growth and behaviour of first-feeding alevins: flood simulation in a semi-natural environment

Introduction

Precipitation projections for 2100 from the Intergovernmental Panel on Climate Change (IPCC) forecast an increase of the average precipitation in Northern and Central Europe from October to March, while no change or a moderate reduction is expected in Southern Europe and Mediterranean (IPCC 2013). Then, Global Climate Change should increase precipitation during winter and spring in the northern distribution range of brown trout and freshwater ecosystems should face more frequent and extreme floods. Such events should lead to a decrease in the abundance of benthic invertebrates (Nislow et al. 2002; McMullen & Lytle 2012). As the number of invertebrates drifting is linked with benthic abundances (*i.e.* the benthic-drift hypothesis; McLay 1968; Hildebrand 1974; Kennedy et al. 2014), food availability for drift-feeding fish could be reduced. At the time of emergence, alevins have a limited amount of reserves and need to feed quickly to avoid mortality (Cushing 1972; Skoglund & Barlaup 2006). A desynchronization of species phenology could occur with a limited quantity of prey when alevins need to feed (*i.e.* the Match-mismatch hypothesis; Cushing 1990). Additionally, behaviour of alevins such as foraging activity, aggressive behaviours and dominance relationships could be modified by the scarcity of prey (Keeley 2000).

During reproduction, females dig nests in the gravel, drop hundreds to thousands of eggs immediately fertilised by males and they cover the clutch with gravel. As the development of the eggs depends largely on temperature, individuals of the same clutch are subjected to the

same conditions and hatch at the same time. Similarly, the emergence is quite synchronous between the individuals of the same clutch but also between the clutches laid simultaneously. Consequently, densities of just-emerging alevins can be very high (Jenkins et al. 1999). As salmonids are territorial species, alevins establish territories progressively (Grant 1997) and defend a feeding position to ensure a positive net energy balance and to optimise the energy gained. Territoriality and aggressive interactions can be noted as early as the first day after emergence (Kalleberg 1958) or 2.5 days after emergence (Titus & Mosegaard 1991). The size of the territory is related to the size of fish and alevins defend small areas (0.1–0.2 m² – Grant & Kramer 1990; Grant et al. 1998). However, the territory size and the intensity of the competition are influenced by other factors including competitor density, resource density and resource distribution in space and time (Brown 1964). The timing of the emergence plays an important role in competition and alevins emerging early can settle in more profitable position than congeners emerging later (Metcalf & Thorpe 1992; Einum & Fleming 1999; Johnsson et al. 1999). The threshold model of feeding territoriality (Carpenter 1987) predicts that animals defend feeding territories according to the food abundance: when the food is too scarce, territorial individuals have a lower fitness compared to non-territorial ones; on the contrary, when food is superabundant, non-territorial individuals gain the same amount of food as territorial ones without paying the cost of defence (Grant et al. 2002; Brännäs et al. 2003; Imre et al. 2004).

The social hierarchy is organized by dominant individuals, subdominant and less aggressive, subordinate fish (Adams et al. 1998; Sloman et al. 2000). Generally, large individuals have the higher growth potential and dominate their congeners and the access to the best stream positions affording maximum potential profits (Chapman 1962; Mason & Chapman 1965; Fausch 1984). Indeed, high metabolic turnover rate is needed to allocate energy both to somatic growth and to territorial defence.

The diminution in prey availability would increase both territory size and foraging activity of alevins to increase encounter rate of prey (Biro et al. 2003) and maintain a constant abundance of food (Toobaie & Grant 2013). This would result in alevins emigration or death and a decrease in fish density when food abundance is low. Aggressive behaviour of young brown trout impacts their growth and survival and influence significantly the population dynamics (Biro et al. 2003) and it is necessary to consider events occurring at the individual level to understand these regulatory mechanisms (Titus 1990). Salmonid abundance is related to the individual territory size, consequently, any environmental factor affecting territoriality should affect

population density (Grant et al. 1998). Moreover, food availability for fish depends on the productivity of the system and it is established that productivity is low during winter and increases in spring when day length, brightness and temperature increase (Sumner & Fischer 1979; Nakano & Murakami 2001).

So, in order to assess the interaction between flood and some components of the ecosystem phenology (emergence timing and invertebrate production) on young salmonid performances, we conducted two experiments at different timing (early and late spring). In a semi-natural stream, we put first-feeding fish in large cages (to assess performances) and in small cages (to monitor behaviour). Then, we compared invertebrate production as well as alevins performances and behaviour according to the occurrence or the absence of a hydraulic disturbance (artificial flood) just before alevins release.

We aimed to test the following hypotheses:

1. Flood should diminish the abundance of invertebrates,
2. Flood should diminish food availability for fish, which should reduce growth and survival of alevins,
3. Flood should lead to an increase in alevins activity to search for prey and a more intense competition between congeners,
4. Ecosystem productivity should increase between early and late spring.

Material and Methods

Experimental design

The Lapitxuri semi-natural stream is a 2.80 m wide channel divided into 13 reaches of 10 m long each. Water is diverted from a tributary of the river Nivelle located near the French-Spanish border in the upstream part of the Nivelle watershed. Lapitxuri tributary is a typical trout brook characterized by a good water quality. We realized two trials of the same experiment, one in early and one in late spring. The experiment took place in two reaches of the artificial stream (30 m² each) located in the middle of its course (60 m downstream the flow entry). Flow was kept constant during the whole experiment (60 L s⁻¹). On 11/02/2016, three large cages (LxWxH: 100x100x50 cm – 1 m²; growth cages) and two small cages (LxWxH: 50x25x50 cm – 0.0125 m²; observation cages) were set up into each reach (Figure 1). The cages were buried 10 cm deep in the substrate. The particle size distribution was the same in all cages,

i.e. cobbles and pebbles ranging from 1 cm to 5 cm in size. The upstream and downstream gates of the cages (facing the water flow) were kept open to let the invertebrates colonise the cages until the beginning of the experiment. When fish were placed, they were closed to prevent the emigration of fish. The mesh size of the cages was 1 mm (even in the buried floor), except on the upstream gate, which was 2 mm to allow entering small invertebrates. Inside the cages water velocity was 5.16 cm s^{-1} (SD = 1.44) and water depth 12.57 cm (SD = 1.45) on average during the experiment. The large cages were used to study alevins growth and survival, while the small cages were used to study alevins behaviour. The observation cages were installed in front of the window of two underwater observation rooms. In the upstream reach, observation cages were located on the left bank at 4 m from the upstream limit of the reach. In the downstream reach, observation cages were located on the right bank at 2 m from the upstream limit of the reach (Figure 1).

Two trials took place, one in March (early spring) and one in April-May (late spring). Reach 1 (upstream) was used as Control Reach, whilst Reach 2 (downstream) was used as Impacted Reach (flood simulation). In the early spring (first trial) two growth cages were used in the “Control” and in the “Flood” Reaches. They were taken out before the second trial. Only one growth cage per reach was used in late spring (second trial).

To simulate the effect of a scouring flood, the out-flow coming from the flexible hose of a motor-pump was directed downward with a 30° angle to the gravel for 10 seconds, disturbing the gravel on a 20 cm wide and 50 cm long area. The flooding operation started at the right bank of the upstream end of the reach 2, then the pump was displaced by 20 cm to the left and used again for 10 seconds. This procedure was repeated over the entire width of the reach (14 times) and such transects were made every 50 cm to cover the entire area of the reach. The use of the motor-pump increased the mean water velocity to 97.74 cm s^{-1} (SD = 29.42) at 50 cm in front of the outflow. These velocities are comparable to velocities measured in the Nivelle river during a Q_{10} flood (*i.e.* a flood that has a 10% chance to occur due to its high intensity – see Figure S1 and Table S1 in Supplementary Information). After the flood simulation, the gravel bed was flattened again.

Fish processing

First-feeding alevins were randomly split in batches: four fish per batch for the observation cages and thirty-two alevins per batch for the growth cages. All of them were individually weighted, measured (total body length, BL), photographed for individual identification using melanophore distribution patterns (Garcia de Leaniz et al. 1994) and released in control and

impacted cage the same day, or the day after the artificial flood. Fish density in both growth and behaviour cages was 32 fish m². At the end of the experiment, all the surviving alevins were manually recovered (after 22/21 days in the growth cages, and 21/15 days in small cages, Table 1) and left fasting 24h. They were then anaesthetised, individually weighted, measured (BL), photographed and subjected to a benzocaine lethal anesthesia.

In order to monitor individual behaviour, fish were marked with Visible Implant Elastomer (VIE) alongside the anal fin (Olsen & Vøllestad 2001). Three days before fish release, a forty fish devoted to the observation cages were anaesthetised and tagged. Three fluorescent VIEs were used (Orange, Green and Blue), together with the non-fluorescent white colour (Figure 1). On the release day, four fish per colour were chosen according to the quality of the mark. They were anaesthetised, weighted, measured and allocated to the four observation cages (each cage received four fish having a different VIE-tag colour). Behavioural surveys were done three times a day, every day during the establishment of the hierarchy (the first week) and every two days during the second week. Surveys were done around 10:00am, 1:00pm and 4:00pm. For each cage, the location of all fish was first noted on a tridimensional map of the cage. Then, focus observations started for each targeted fish for 5 minutes (successively) and the following behaviours were noted: (i) prevailing activity (2 states: resting on the substratum or swimming), (ii) number of catch prey attempts, (iii) distance of capture (3 categories: short < 2 cm, medium = 2-5 cm, long > 5 cm), and (iv) number of attacks given (4 types: intimidation, charge, nip or chase – Adams et al. 1995) or (v) reaction to attack (3 types: no reaction, fleeing, riposte).

Benthic invertebrate sampling

Immediately after the flood simulation (07/03/2016 and 21/04/2016) and at the end of the experiment (30/03/2016 and 12/05/2016), 3 invertebrate samples were randomly collected in growth cages. Sampling was carried out by burying a corer (ø: 13 cm) 10 cm deep in the substrate. Gravels and pebbles were then gently removed and the invertebrates in the pipe pumped, recovered in a 500 µm sieve and preserved in 70% ethanol. This was repeated 3 times per cage, providing 12 invertebrate samples for the early spring trial and 6 for the late spring one. Invertebrates were identified and assigned to different groups: Diptera, Trichoptera, Ephemeroptera, Mollusca, Coleoptera, Plecoptera, Oligochaeta, Crustacea and “Others”.

The origin of the fish as well as the main environmental characteristics during each trial are summarized in Table 1.

Table 1 Date, conditions and events at each step of the experiment for both trials (in early spring and in late spring).

	1 st trial – Early spring	2 nd trial – Late spring
Origin of genitors	Wild Nivelle watershed	Produced by INRA experimental facilities (Lees-Athas)
Rearing temperature (°C)	9.00 (SD = 2.33)	8.58 (SD = 1.37)
Development of alevins (Degree-days) at the time of release	787	740
Flood simulation		
Mean water velocity (cm s ⁻¹)	86.37 (SD = 23.25)	109.37 (SD = 30.50)
Minimum water velocity (cm s ⁻¹)	51	51
Maximum water velocity (cm s ⁻¹)	150	217
Number of benthos samples	24	12
Observation period (days)	21	15
Number of surveys	40	36
Fish growth period (days)	22	21
Mean water temperature (°C)	10.58 (SD = 0.9)	12.33 (SD = 1.4)

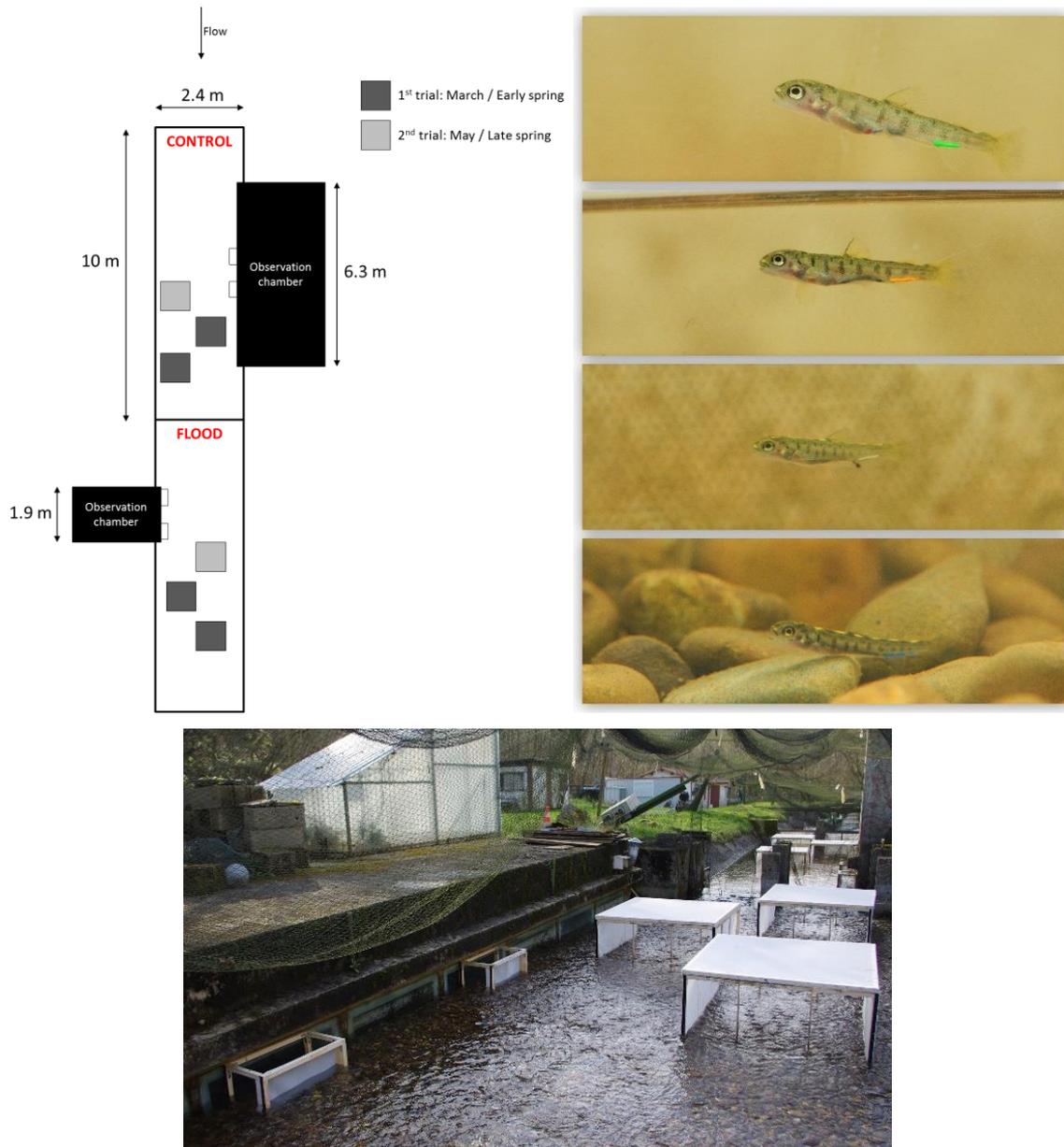


Figure 1 Experimental setup in the semi-natural Lapitxuri channel. The upstream reach was the control, while a flood was simulated in the downstream reach. Four cages were used for the growth of alevins in March/early spring (in dark grey) and two in April-May/late spring (in light grey). Behavioural surveys were conducted in additional small cages (not shaded), close to the observation chambers during the two trials with alevins tagged differently (top right photos). The picture shows an overview of the device.

Statistical analyses

Growth Cages

Abundance of invertebrates (number of individuals by sample) as well as the individual weight gain of alevins and fish productivity (sum of final weights) were analysed using a bayesian modelling approach computed with OpenBUGS®. We assumed that the variables of interest

followed a normal distribution. Then, the mean of the variable (μ .Variable) was modelled according to three parameters (Table 2): a fixed-effect relative to the flood (α), a fixed-effect relative to the season (β) and a fixed-effect relative to the interaction flood*season (γ). The logit of the probability of alevins survival was modelled with these same three effects.

*Table 2 Parameters taken into account according to the season and the condition. μ .Variable is the mean of the variable of interest, α represents the flood effect, β the second trial effect and γ the flood*second trial effect.*

		Season	
		1 st trial (early spring)	2 nd trial (late spring)
Conditions	Control	μ .Variable	μ .Variable + β
	Flood	μ .Variable + α	μ .Variable + α + β + γ

Abundance of invertebrates

If $Abond_i$ is the number of individuals counted in the i^{th} sample, we assumed:

$$\mu.\log(Abond_i + 1)_i = \mu + \alpha Flood_i + \beta Season_i + \gamma Interaction_i$$

Where μ is the mean of the logarithm of the abundance of invertebrates, α is a fixed-effect parameter for the impact of the flood, β is a fixed-effect parameter for the season and γ is a fixed-effect parameter for the interaction flood*season. The same modelling treatment was applied to the total abundance of invertebrates and to the abundance per group (*i.e.* Diptera, Trichoptera, Ephemeroptera, Coleoptera, Plecoptera, Oligochaeta, Crustacea, Mollusca, and “Others”).

Survival probability of alevins

We assumed that the alevins could not escape from the cage and so, not recovered alevins at the end of each trial were considered dead. Then, if $p.S_i$ is the probability of survival of the i^{th} alevin, we assumed:

$$S_i \sim \text{Bernoulli}(p.S_i)$$

$$\text{Logit}(p.S_i) = \mu + \alpha Flood_i + \beta Season_i + \gamma Interaction_i$$

Where μ is the mean of the logit of the survival probability, α is a fixed-effect parameter for the impact of the flood, β is a fixed-effect parameter for the season and γ is a fixed-effect parameter for the interaction flood*season.

Individual weight gain of alevins

All the alevins were weighted at the beginning of each trial ($W.beg_i$) and all the survivors were weighted at the end ($W.end_i$). Photos allowed the individual recognition of alevins and then calculation of the individual Weight Gain (WG_i):

$$WG_i = \frac{W.end_i - W.beg_i}{W.beg_i}$$

Then, we stated that WG followed a normal distribution with $\mu.WG_i$ the mean and $\sigma.WG_j$ the standard deviation. We assumed:

$$\mu.WG_k = \mu + \alpha Flood_i + \beta Season_i + \gamma Interaction_i$$

Where μ is the mean of the individual weight gain of alevins, α is a fixed-effect parameter for the impact of the flood, β is a fixed-effect parameter for the season and γ is a fixed-effect parameter for the interaction flood*season.

Fish productivity

The final weights of alevins ($W.end_i$) were summed to obtain the fish production of each j cage ($Prod.Fish_j$).

$$\mu.Prod.Fish_j = \mu + \alpha Flood_j + \beta Season_j + \gamma Interaction_j$$

Where μ is the mean fish production, α is a fixed-effect parameter for the impact of the flood, β is a fixed-effect parameter for the season and γ is a fixed-effect parameter for the interaction flood*season.

Determination of hierarchy

Only One Side Attacks (OSA) were considered to establish the hierarchy matrix, *i.e.* attacks that induced a fleeing reaction or a gently sink underneath and downstream the aggressor (Katano 1985, 1990; Nakano & Furukawa-Tanaka 1994). Aggression Index (AI) was calculated for each alevin by dividing the number of OSA made (OSA+) by the number of OSA sustained (OSA-). Then, the AI was related to the positioning of the alevin in the cage and alevins were ranked. "A" was associated to the highest AI and an upstream positioning within the cage and referred to the dominant alevin. "B" was associated to the second highest AI and referred to the subdominant alevin. "C" and "D" was associated to the lowest AI and a downstream positioning within the cage and referred to the subordinates.

Behavioural analyses

Following emergence, fish try to establish territories and start to feed. They have to learn how to swim in the water current, how to catch prey, and the hierarchy take a few days to be established. Alevins became progressively active and began to hunt and interact with their congeners over time. Therefore, δ represented the increase in the probability to be active or in the occurrence of discrete events (*i.e.* attempt to catch prey or aggressive interaction), δ_1 being the increase in control cages and δ_2 the increase in impacted cages. Difference between δ_1 and δ_2 was tested. Moreover, in the same reach, the second cage was positioned 2 meters downstream the first one, then a fixed-effect parameter λ relative to downstream cage positioning was also considered (Table 3).

Table 3 Parameters taken into account according to the condition and the position of the behavioural cage. μ .Variable is the mean of the behaviour of interest, δ_1 represents the increase in the control cages, δ_2 the increase in the impacted cages and λ the downstream cage position effect.

		Position of the behavioural cage in the reach	
		Upstream	Downstream
Conditions	Control	μ .Variable + δ_1	μ .Variable + δ_1 + λ
	Flood	μ .Variable + δ_2	μ .Variable + δ_2 + λ

If $p.A_k$ is the probability of being active (swimming in the water column) for an alevins during the k^{th} survey, we assumed:

$$Activity_k \sim Benoulli(p.A_k)$$

$$Logit(p.A_k) = \mu + \delta_{Condition[k]}Time + \lambda Position_k$$

Where μ is the average of the logit of the probability of alevins to be active, δ is a fixed-effect parameter for the increase in activity over time and λ is a fixed-effect parameter for the position of the cage.

If $Hunt_k$ is the number of attempts to catch prey (the feeding activity) of an alevin during the k^{th} survey of five minutes, we assumed:

$$Hunt_k \sim Poisson(\mu.Hunt_k)$$

$$\log(\mu.Hunt_k) = \mu + \delta_{Condition[k]}Time + \lambda Position_k$$

Where μ is the average of the logarithm of the numbers of attempts to catch prey in 5 minutes, δ is a fixed-effect parameter for the increase of the feeding activity over time and λ is a fixed-effect parameter for the position of the cage.

During a 5-minute survey, all the aggressive acts were summed, even if the focused alevins was the victim. An intimidation, a charge, a nip or a chase were equally considered as aggressive acts. Then, if Agg_k is the number of aggressive interactions during the k^{th} survey of five minutes, we assumed:

$$Agg_k \sim Poisson(\mu \cdot Agg_k)$$

$$\log(\mu \cdot Agg_k) = \mu + \delta_{Flood[k]+1} Time + \lambda Position_k$$

Where μ is the average of the logarithm of the numbers of aggressive interactions in 5 minutes, δ is a fixed-effect parameter for the increase of the aggressive activity over time and λ is a fixed-effect parameter for the position of the cage.

Correlations between individual weight gain of alevins and (i) the number of surveys during which each alevin was actively swimming in the water column, (ii) the average number of attempts to catch prey by survey and (iii) the average number of aggressive interactions by survey were tested as previously described. Similarly, the number of surveys during which each alevin was actively swimming was linked to the average number of attempts to catch prey by survey and the average number of aggressive interactions by survey. Correlations were tested with OpenBUGS®. If Y_j was the individual weight gain or the alevin activity, we assumed:

$$Y_j = a Behaviour_j + b$$

Where Y_j is for each j alevin, the individual weight gain (WG_j) or the activity of alevins, a is the slope coefficient, $Behaviour_j$ is the behaviour of interest and b is the intercept. Fit of correlation was calculated by dividing the variability unexplained by the variability observed in the data set, as follows:

$$Fit = 1 - \frac{Variability\ unexplained\ by\ the\ model}{Variability\ of\ the\ data\ set}$$

When the fit is between 0.5 and 1, the correlation is good. When the fit is between 0 and 0.5, the correlation is intermediate and when the fit is negative, there is no correlation.

Bayesian computations

Parameters were given independent “weakly informative” priors. Following recommendations of Gelman & Hill (2007), μ , α , β , γ , a , b , δ_1 , δ_2 and λ were sampled in a normal distribution, $N(0, 100)$. $\sigma_{\log.Aband}$ and $\sigma_{\text{Prod.Fish}}$ were sampled in a half-Cauchy truncated distribution, $t(0, 1, 1)$. Hyperparameters of $\sigma.WG_j$ (B. $\sigma.WG$ and E. $\sigma.WG$) were sampled in a gamma distribution, $\text{gamma}(1, 1)$. We used each time three independent chains, the first 10 000 iterations were discarded as an initial burn-in period. Then, 10 000 further iterations (resulting from one every ten runs) were performed. The convergence of the chains to their ergodic distribution was tested *via* the Gelman-Rubin (GR) diagnostics integrated in OpenBUGS®. The significance of the parameters was tested with the step function implemented in OpenBUGS®. At each iteration for a variable X , $\text{step}(X)$ equaled 1 if $X \geq 0$ and equaled 0 if $X < 0$. At the end of the run, if $P(X > 0)$ was lower than 0.1 or higher than 0.9, the parameter X was considered to be different from 0.

Results

Effect of the flood on the community of invertebrates

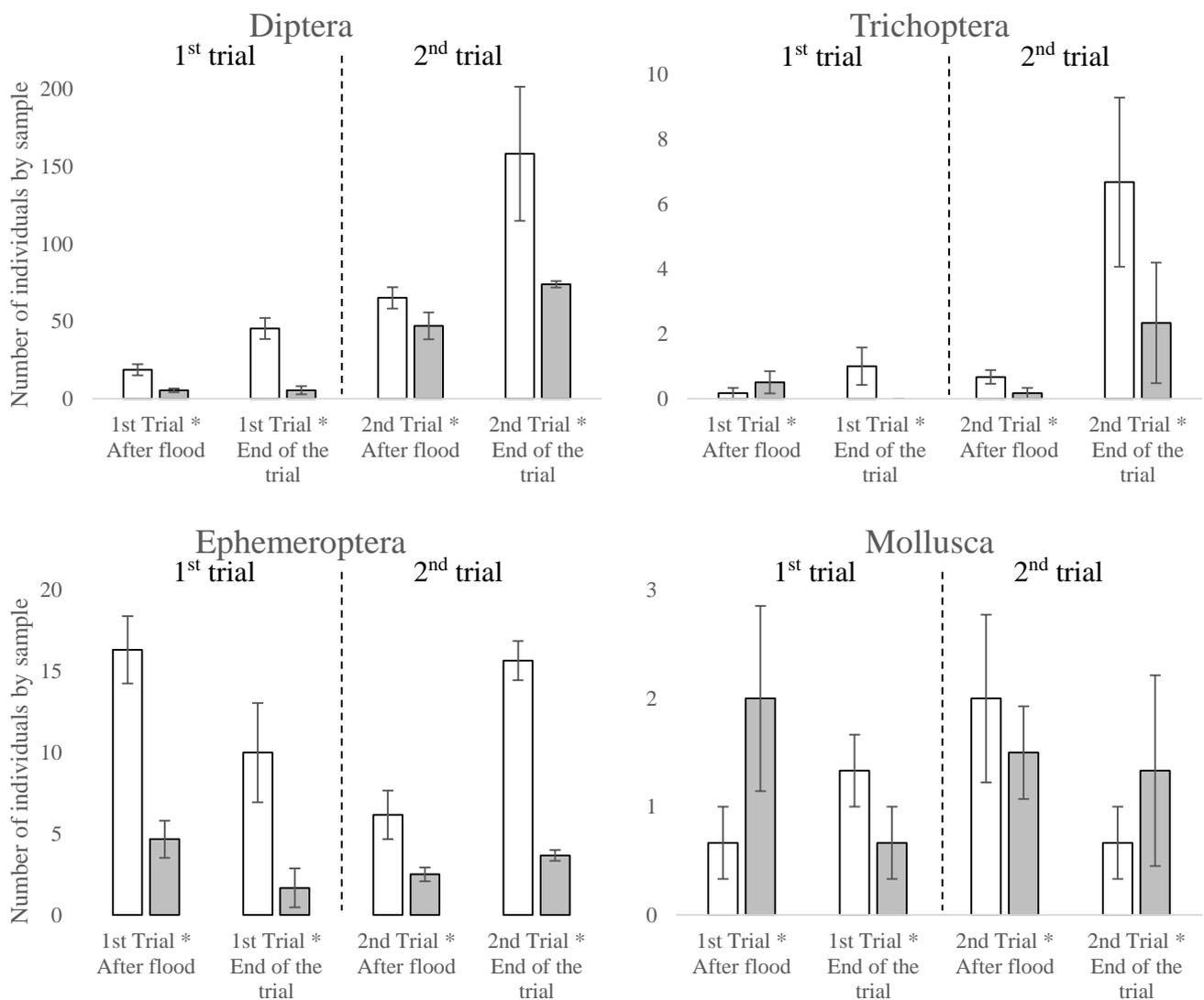
During the first trial (early spring), immediately after the artificial flood, Diptera, Ephemeroptera, Coleoptera, Plecoptera and the “Others” groups were less abundant in the impacted than in the control cages. The abundance of Mollusca and Crustacea increased, while the abundance of Trichoptera, Oligochaeta and the total abundance did not change (Figure 2 and Table 4 – α_1). At the end of the trial, the abundance of Diptera, Ephemeroptera, Coleoptera, Oligochaeta and the total abundance decreased in the impacted cages, while the abundance of Crustacea increased. The abundance of Trichoptera, Mollusca, Plecoptera and the Other invertebrates remained equal between the impacted and the control cages (α_2).

During the second trial (late spring), the artificial flood decreased the abundance of all groups of invertebrates (α_1 and γ_1). At the end of the trial, the abundance of Diptera, Ephemeroptera, Crustacea, Coleoptera, Oligochaeta and the total abundance remained lower in the impacted cages, while the abundances of Plecoptera and Exogenous were higher. The abundance of Mollusca, Trichoptera and the Other invertebrates remained equal between the control and the impacted cages (α_2 and γ_2).

Whatever the conditions, there was a significant seasonal effect comparing the two trials. The abundance of Diptera, Trichoptera, Ephemeroptera, Oligochaeta, Crustacea, Others and the total abundance was higher in late spring than in early spring (β_1 and β_2).

The important information to better understand trophic availability concerns the main prey groups for 0⁺ trout, *i.e.* Diptera and Ephemeroptera. For these groups (but still true for total abundances):

- Just after the flood and at the end of the trials, abundances in control were higher than in impacted cages in both early and late spring and these differences were more pronounced at the end of the trial than just after the flood,
- Abundances were much higher in late than in early spring for Diptera, which were ten times more abundant than Ephemeroptera.



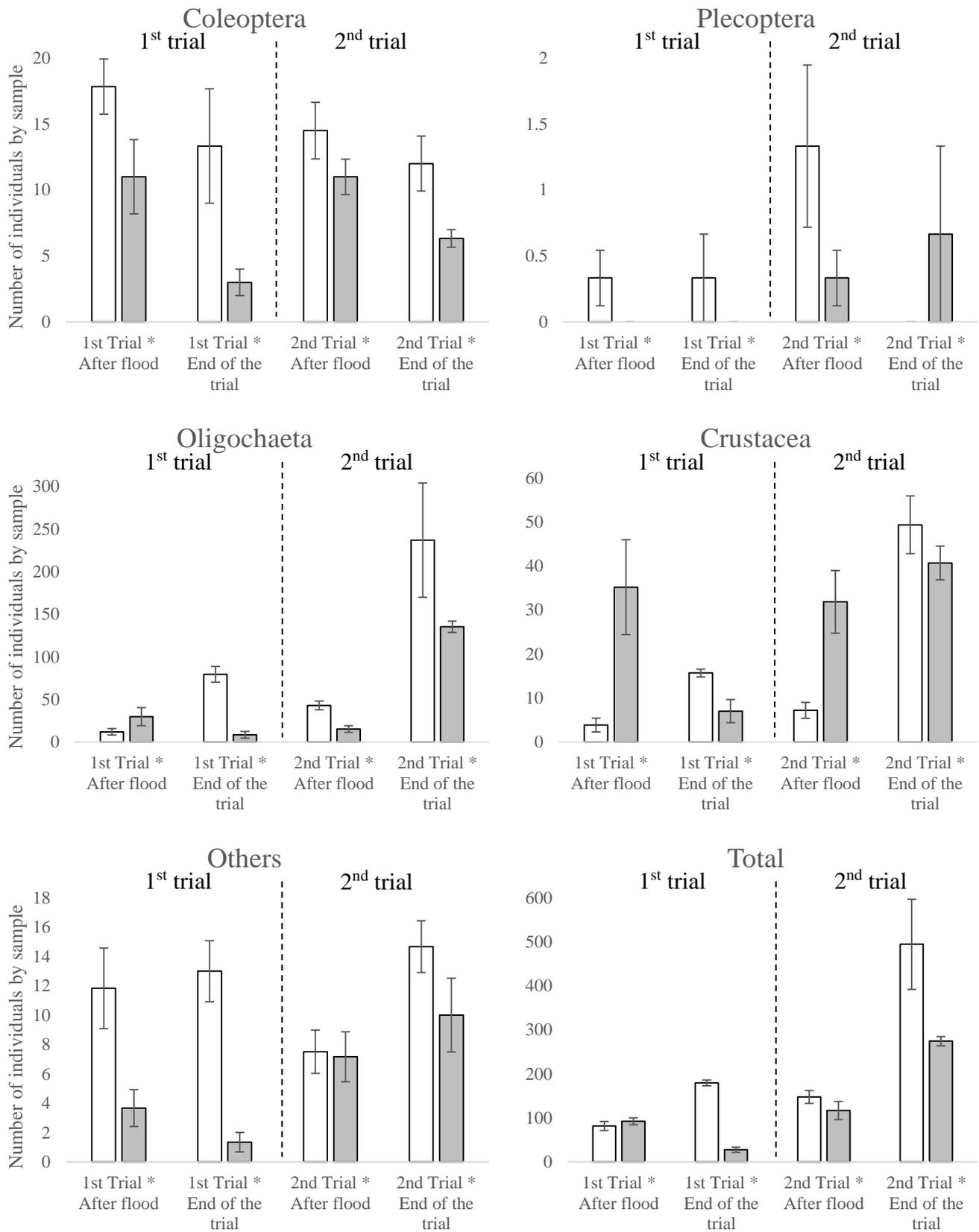


Figure 2 Number of individuals per sample (average \pm standard error) of the groups of invertebrates identified according to treatment (Control in white and Impacted by flood in grey) immediately and at the end of the trial.

Table 4 Mean of the posterior probability distribution functions of the effect of the flood (α), season (β) and the interaction flood*season (γ) on the logarithm of the abundance of invertebrates. Effects were estimated for each group and for the total abundance, immediately after the flood and at the end of the trial. Parameters were considered significant when $P(X > 0)$ is less than 0.10 or above 0.90 (in bold).

Groups	Immediately after the flood						At the end of the trial					
	α_1	$P(\alpha_1 > 0)$	β_1	$P(\beta_1 > 0)$	γ_1	$P(\gamma_1 > 0)$	α_2	$P(\alpha_2 > 0)$	β_2	$P(\beta_2 > 0)$	γ_2	$P(\gamma_2 > 0)$
Diptera	-1.245	0.007	0.926	0.948	-1.099	0.084	-0.388	0.059	0.834	0.994	-0.291	0.238
Trichoptera	0.180	0.773	0.478	0.941	-0.774	0.038	-0.346	0.143	1.423	0.999	-0.659	0.123
Ephemeroptera	-1.259	0.002	-0.478	0.151	-0.299	0.321	-0.642	0.006	0.956	0.997	-0.631	0.066
Mollusca	0.508	0.946	0.415	0.865	-0.874	0.057	-0.065	0.428	-0.429	0.169	0.290	0.683
Coleoptera	-0.591	0.035	-0.345	0.185	-0.637	0.123	-0.240	0.079	-0.160	0.219	-0.315	0.140
Plecoptera	-0.232	0.082	-0.001	0.498	0.000	0.500	-0.412	0.106	-0.640	0.059	0.774	0.913
Oligochaeta	0.635	0.887	2.013	0.998	-2.909	0.002	-1.112	0.000	1.628	1.000	0.651	0.917
Crustacea	2.083	1.000	1.519	0.994	-2.950	0.001	1.435	1.000	1.955	1.000	-1.613	0.005
Others	-1.134	0.005	0.184	0.651	-0.744	0.142	-0.115	0.360	0.690	0.957	-0.275	0.310
Total	0.138	0.795	0.819	0.999	-2.037	0.000	-0.280	0.088	1.176	1.000	-0.255	0.233

Effect of the flood on alevins

The survival probability was significantly higher in the control than in the impacted cages during the first trial (early spring – 0.609 vs 0.376 – $P(\alpha > 0) = 0.004$ – Figure 3 and Table 5). The opposite was observed during the second trial (late spring), even if the ratio between the two survival levels was much lower (0.813 vs 0.937 – $P(\gamma > 0) = 0.995$).

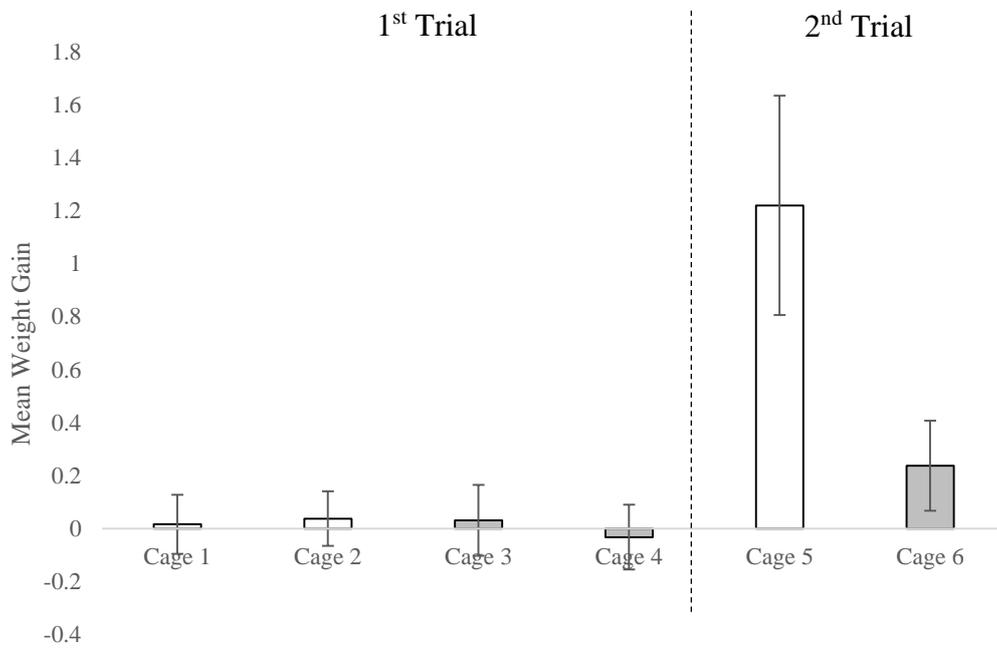
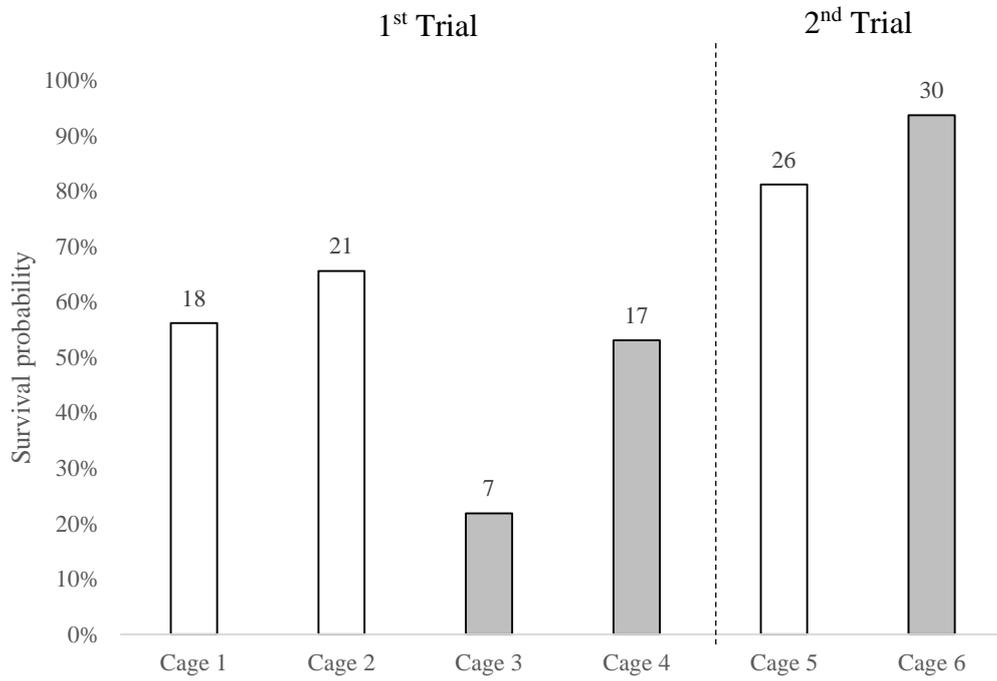
At the beginning of the experiment, alevins weighted on average 0.117 g (SD = 0.007) at the first trial (early spring) and 0.078 g (SD = 0.008) at the second trial (late spring). After 21 or 22 days of growth, alevins weighted on average 0.118 g (SD = 0.015) at the end of first trial and 0.133 g (SD = 0.052) at the end of the second trial. Individual weight gain was higher in the control than in the impacted cages for both trials (0.028 vs -0.018 and 1.221 vs 0.237 – $P(\alpha > 0) = 0.100$ and $P(\gamma > 0) = 0.000$).

Finally, there was no significant difference in fish productivity, but it tended to be higher in the control cages than in the impacted ones at both trials (2.349 vs 1.384 and 4.510 vs 2.928 – $P(\alpha > 0) = 0.110$ and $P(\gamma > 0) = 0.294$). Whatever the considered variable, alevins performances were lower at the first trial than at the second one ($P(\beta > 0) = 0.984, 1.000$ and 0.964 for survival, individual weight gain and fish productivity).

*Table 5 Mean of the posterior probability distribution functions of the effect of flood (α), season (β) and the interaction flood*season (γ) on the survival probability, on individual weight gain of alevins and on fish productivity. Parameters were considered significant when $P(X > 0)$ is less than 0.10 or above 0.90 (in bold).*

Alevins performance	Parameters	Mean	$P(X > 0)$
Survival probability	α	-0.963	0.004
	β	1.086	0.984
	γ	2.373	0.995
Individual weight gain	α	-0.046	0.100
	β	1.192	1.000
	γ	-0.938	0.000
Fish productivity	α	-0.965	0.110
	β	2.161	0.964
	γ	-0.618	0.294

A



B

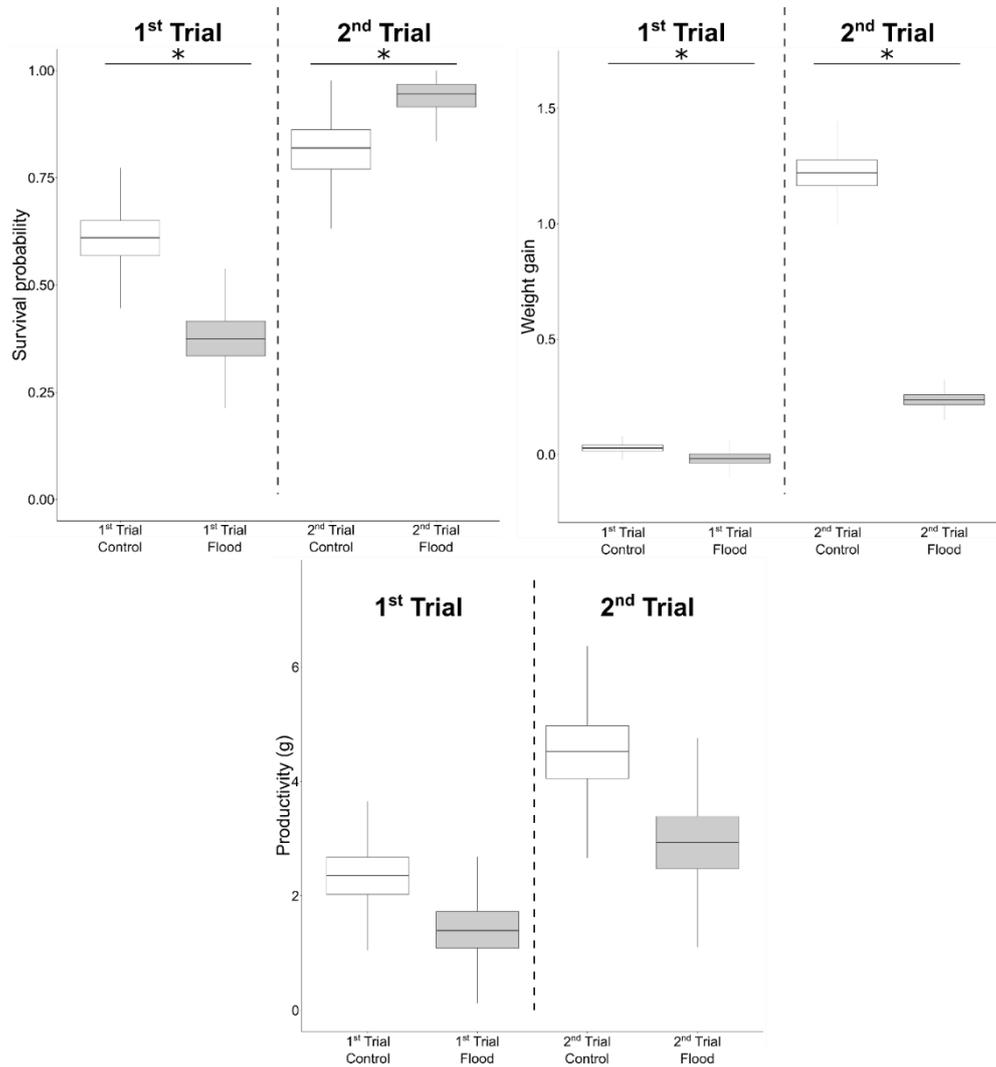


Figure 3 Raw data (A) of the survival probability and mean weight gain by cage and model estimates (B) of the survival probability, the individual weight gain and fish productivity in control (in white) and impacted (in grey) cages in the first and the second trial. Boxplots indicate the 1, 25, 50, 75 and 99 percentiles of posterior distributions. Significant differences are shown by *.

To sum up:

- Survival was low in early spring (50%) and high in late spring (80%),
- Growth was lower in early spring (close to zero) than in late spring (70% increase in weight),
- The artificial flood diminished the survival and growth in early spring,
- The artificial flood enhanced the survival, while the growth was low in late spring.

Behaviour of alevins

During the first trial (early spring), mortalities were recorded at the end of the 21 days of the observation survey, but we ignore when they arose. There was only 50% of survival with only two alevins surviving out on the four initially set up in each cage. Survivors grew, between 0.083 and 0.327 in the control and between 0.143 and 0.748 in the impacted cages (Table S2). Alevins appeared to do more attempts to catch prey in the control cages and there were also more aggressive interactions. However, because the number of alevins in the cage changed over time, it was not possible to analyse behavioural data for the first trial.

During the late spring surveys, we found a strong common pattern in the distribution of the four alevins within each cage. The dominant fish was positioned in the upstream part of the cage, immediately behind the net through which the invertebrates penetrated by drift into the cage. Then, the subdominant fish was positioned side by side or just downstream the dominant. The last two subordinates were generally positioned in the downstream part of the cage and accessed only to prey uncaught by the two upstream fish. The dominant and the subdominant fish gained the most weight and caught the most prey (Table 6), except in cage 2 in which differences in growth and feeding activity were less marked between alevins ranked B, C and D. There was no significant difference in average individual weight gain of alevins according to the conditions ($P(WG_{\text{Flood}} > WG_{\text{Control}}) = 0.818$).

Table 6 Initial and final weight, individual weight gain, mean number of feeding attempts by survey, One-Side-Attacks made (OSA+), sustained (OSA-), social rank and positioning of the alevins during the second trial (late spring) according to their cage of origin and the colour of their tag.

Condition	Cage	VIE tag	Initial Weight (mg)	Final Weight (mg)	Individual weight gain	Mean Number of feeding attempts by survey	OSA+	OSA-	Aggression Index	Rank	Social status	Position	
Control	1	Green	88	119	0.352	15.63	19	129	0.15	D	Subordinate	Middle	Bottom
		Orange	83	110	0.325	8.17	27	57	0.47	C	Subordinate	Downstream	Bottom
		Blue	70	151	1.157	14.86	139	4	34.75	A	Dominant	Upstream	Bottom
		White	93	165	0.774	12.03	74	69	1.07	B	Subdominant	Up/Mid	Surface
	2	Green	73	93	0.274	1.71	11	25	0.44	C	Subordinate	Up/Mid/Down	Bottom
		Orange	88	117	0.330	4.23	5	20	0.25	D	Subordinate	Middle	Bottom
		Blue	93	120	0.290	1.91	10	8	1.25	B	Subdominant	Downstream	Surface
		White	81	166	1.049	12.86	35	8	4.38	A	Dominant	Upstream	Bottom
Flood	3	Green	93	106	0.140	0.85	2	32	0.06	D	Subordinate	Downstream	Bottom
		Orange	89	224	1.517	17.58	30	4	7.50	A	Dominant	Upstream	Bottom
		Blue	85	167	0.965	10.94	20	9	2.22	B	Subdominant	Downstream	Surface
		White	73	127	0.740	8.06	23	30	0.77	C	Subordinate	Middle	Bottom
	4	Green	87	221	1.540	11.50	32	5	6.40	A	Dominant	Upstream	Bottom
		Orange	74	115	0.554	3.42	5	44	0.11	D	Subordinate	Downstream	Bottom
		Blue	75	86	0.147	1.97	3	9	0.33	C	Subordinate	Mid/Down	Bottom
		White	72	125	0.736	11.50	29	11	2.64	B	Subdominant	Mid/Down	Bottom

Over the 15 days of observation, the probability of activity, the number of attempts to catch prey and the number of aggressive interactions during a survey (five minutes) increased but the intensity of the increase always differed between the control and the impacted cages (Table 7).

Table 7 Mean of the posterior probability distribution functions of the increase in the activity probability or in the occurrence of discrete events (i.e. attempts to catch prey or aggressive interactions) over time (δ_1 being the increase in control and δ_2 in impacted cages) and the effect of cage positioning (λ). Parameters were considered significant when $P(X > 0)$ is less than 0.10 or above 0.90 (in bold).

Behaviours	Parameters	Mean	$P(X > 0)$	$P(\delta_1 > \delta_2)$
Activity probability	δ_1	0.214	1.000	} 0.002
	δ_2	0.331	1.000	
	λ	-1.483	0.000	
Number of attempts to catch prey / survey	δ_1	0.087	1.000	} 1.000
	δ_2	0.076	1.000	
	λ	-0.603	0.000	
Number of aggressive interactions / survey	δ_1	0.197	1.000	} 1.000
	δ_2	0.135	1.000	
	λ	-0.986	0.000	

Activity

Activity increased with time in both control and flood cages. Under impacted conditions, probabilities of being active were slightly higher (Figure 4).

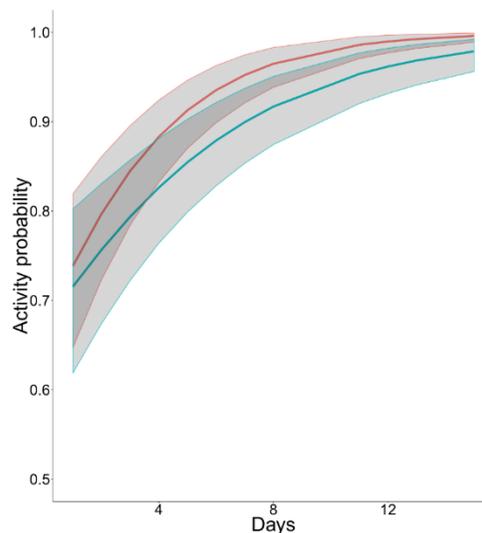


Figure 4 Model estimations of the probability to be active of an alevins according to time (in days) in the control (in blue) or in the impacted conditions (in red). Shaded areas are the 95% probability intervals of posterior distributions.

Feeding attempts

Number of feeding attempts increased with time in both conditions, but the increase was more pronounced under control conditions from day 3-4. Fish made on average 6 feeding attempts by survey on the first day, 11 vs 10 on the seventh day and 21 vs 18 on the fifteenth day (Figure 5).

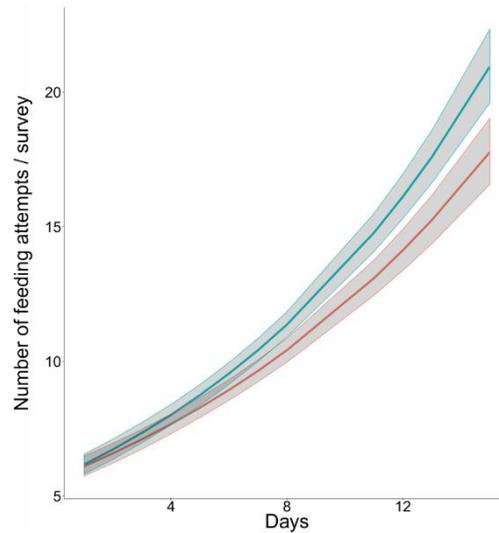


Figure 5 Model estimations of the number of feeding attempts during a survey (five minutes) according to time (in days) in the control (in blue) or in the impacted conditions (in red). Shaded areas are the 95% probability intervals of posterior distributions.

Capture distance

Alevins mostly sought prey around them (0-2 cm, 50.8% of total number of captures) but they also travelled intermediate distances (2-5 cm, 32.1%) or even longer (more than 5 cm, 17.1%). However, there was no significant difference in the temporal trend of the proportions of distances travelled by alevins between control and impacted conditions (Figure 6). It can only be noticed that short distance captures represented 80-90 % of the catches during the first 2 days in the control cages, whilst it established around 60% at the same time in the impacted cages.

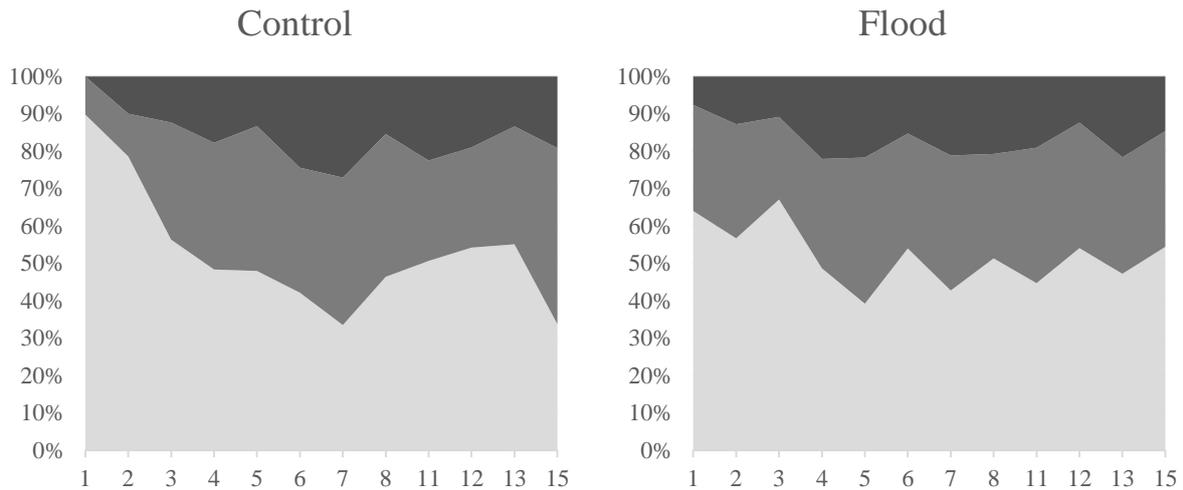


Figure 6 Proportion of short (< 2 cm – in light grey), medium (2-5 cm – in grey) and long (> 5 cm – dark grey) feeding attempts of alevins according to time (in days) in control and in impacted conditions.

Number of aggressive interactions

Aggressiveness increased with time in both conditions, but the increase was more pronounced under control conditions from the sixth day. Alevins made no attack on the first day, 1.5 vs 1 attack on the seventh day and 6.5 vs 2.5 attacks on the fifteenth day (Figure 7).

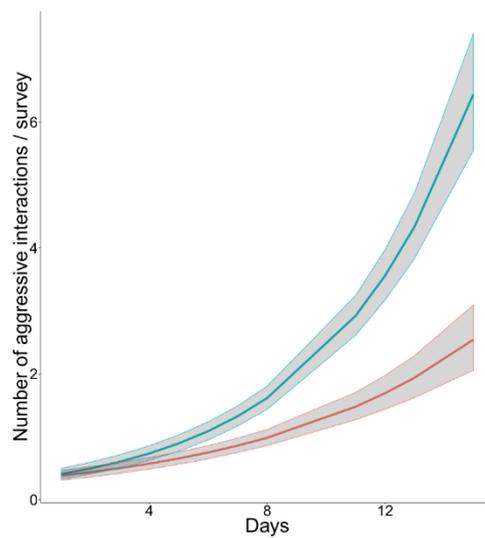


Figure 7 Model estimations of the number of aggressive interactions during five minutes according to time (in days) in the control (in blue) or in the impacted conditions (in red). Shaded areas are the 95% probability intervals of posterior distributions.

Relationship between behaviours

Individual weight gain was correlated with the activity of alevins as well as the average number of attempts to capture prey by survey. Activity of alevins seemed to be correlated with the

average numbers of attempts to capture prey by survey. In all three cases, the fit was higher than 0.50 and slope coefficients (a) were significantly positive (Table 8). The individuals which gained the most weight were the most active and they hunted the most (Figure 8). On the contrary, no link was established with aggressiveness.

Table 8 Correlations between individual weight gain (WG) and the number of survey noted active by alevins, the average number of attempts to catch prey by alevin and the average number of aggressive interactions by alevin. Activity of alevins was also related to the number of attempts to catch prey and the average aggressive interactions. Mean of posterior probability distribution functions of a (the slope coefficient) and b (the intercept) and fit of the models. Parameters were considered significant when $P(X > 0)$ is less than 0.10 or above 0.90 (in bold).

Variable X	Variable Y	a	P(a > 0)	b	P(b > 0)	Equation	Fit
Activity	WG	0.046	1.000	-0.645	0.033	$G = 0.04596 \text{ Activity} + -0.645$	0.534
Attempt to catch prey	WG	0.063	1.000	0.150	0.839	$G = 0.06271 \text{ Catch.Prey} + 0.1497$	0.536
Aggressiveness	WG	0.062	0.673	0.614	0.998	$G = 0.06237 \text{ Agg} + 0.6144$	-0.082
Attempt to catch prey	Activity	1.216	1.000	18.290	1.000	$\text{Activity} = 1.216 \text{ Catch.Prey} + 18.29$	0.692
Aggressiveness	Activity	4.654	0.991	23.080	1.000	$\text{Activity} = 4.654 \text{ Agg} + 23.08$	0.132

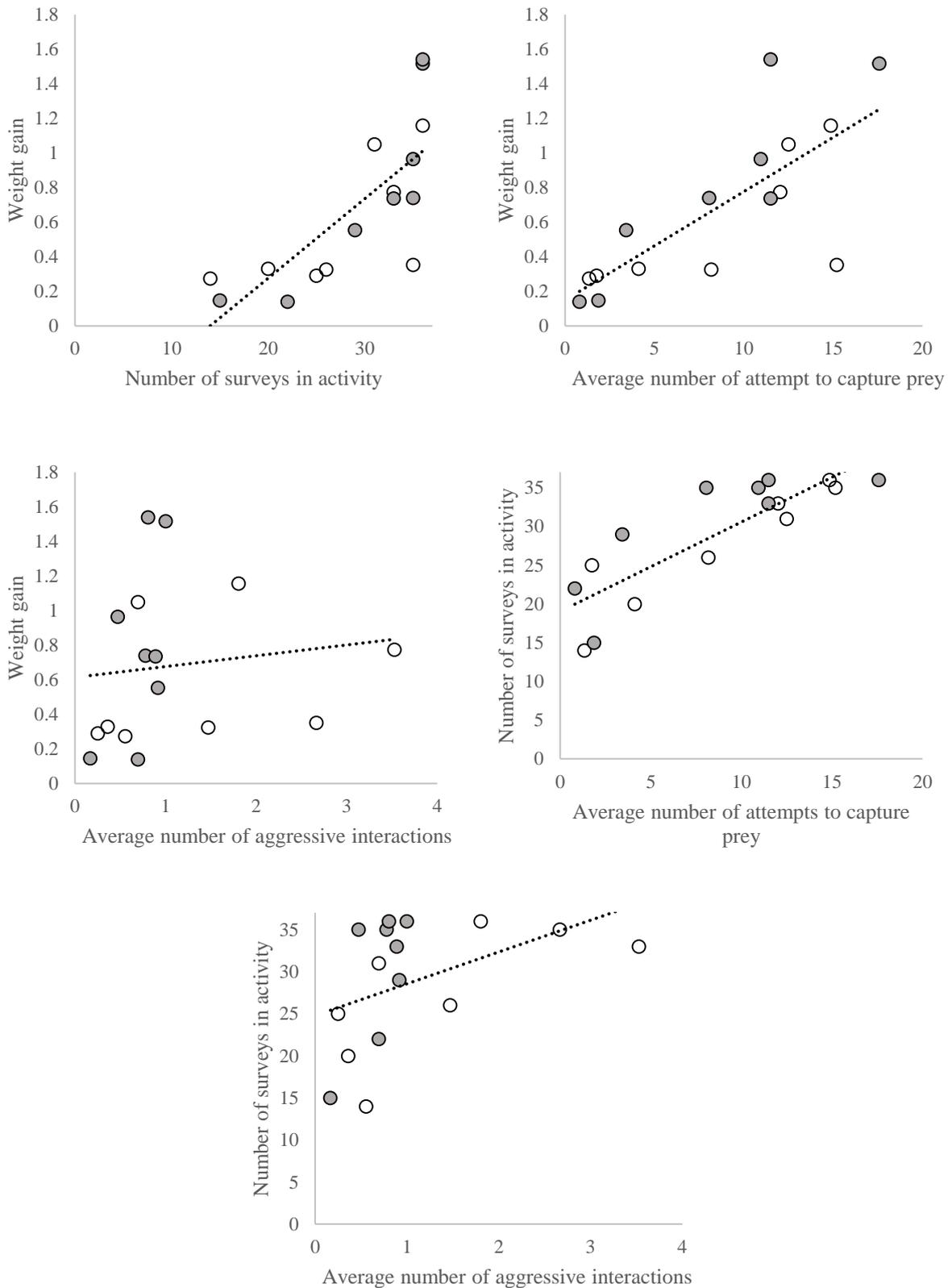


Figure 8 Individual weight gain (WG) according to the number of survey during which alevins were in activity, the average number of attempts to catch prey by alevins and the average number of aggressive interactions by alevins. Activity of alevins was also related to the number of attempts to catch prey and the average aggressive interactions. Empty circles represent alevins in the control cages, grey circles the alevins in the impacted cages and the dotted line the correlation.

Table 9 Synthesis of the main results on the effects of the artificial flood on invertebrate abundances (total and Diptera), alevin performances (survival and growth) and alevin behaviour (activity, feeding activity and competition) in early and late spring.

		Early Spring		Late Spring	
		Control	Flood	Control	Flood
Invertebrates	Abundances ...				
	• Total	LOW	→ LOW	HIGH	↘ MODERATE
	• Diptera	LOW	↘ VERY LOW	HIGH	↘ MODERATE
	Recolonization	LOW	↘ NULL	HIGH	↘ MODERATE
Alevin performances	Survival	LOW 60 %	↘ VERY LOW 40 %	HIGH 80 %	↗ VERY HIGH 95 %
	Growth	NULL 0.03	↘ NEGATIVE -0.02	HIGH 1.2	↘ LOW 0.2
Alevin behaviour	Activity probability				↗
	Feeding attempts				↘
	Strength of competition				↘

Discussion

The experiment conducted in the semi-natural channel of the Lapitxuri highlighted that the artificial flood significantly impact the invertebrate community and the alevin performances and behaviour. However, the impact of the flood differed according to season.

Artificial flood and invertebrate abundances

In early spring, the artificial flood reduced the abundances of Diptera and Ephemeroptera to a third, while the total abundance of invertebrates remained more or less equal. In late spring, the artificial flood decreased by half the abundances of Diptera, Ephemeroptera and the total abundance. It is possible that the low abundances in early spring limited the impact of the artificial flood as observed in other streams when floods arose when invertebrate abundances were low (Arunachalam et al. 1991; Brewin et al. 2000). After twenty days, differences persisted. At that time abundances were the result of both recolonization processes and fish predation (Grosholz & Gallo 2006). Focusing on Diptera and Ephemeroptera, abundances remain lower in the impacted than in the control cages. However, the catch up was much quicker in late spring than in early spring. It could be due to an increase in system productivity that promotes the recolonization process as observed by Miller & Golladay (1996).

Artificial flood and performances of alevins

In early spring, the artificial flood diminished survival in large cages by 23% when compared to control. According to density-dependence mechanism this should reduce competition and promote growth. The growth of alevins was significantly but only slightly lower in the impacted cages. Therefore, the impact of trophic resources has probably been high enough to counter-balance the positive effect of density-dependence. Another possible explanation is that

mortalities occurred only shortly before the end of the experiment, before growth compensation. Then, the occurrence of a flood when the abundance of invertebrates is already low can seriously weaken the strength of the cohort. In late spring, the artificial flood increased unexpectedly the survival by 13%, while it diminished the individual weight gain. Density was higher in the impacted cages and then, the density-dependence mechanisms could partly explain the lower growth. Food availability probably might have had an effect also, since Diptera and Ephemeroptera were more abundant in control than in impacted conditions. It remains difficult to evaluate the respective impact of density and trophic resource limitation on alevin growth (see Supplementary Information Table S3, S4 and Figure S2 for details). The higher survival in impacted cages is surprising and it was probably linked to uncontrolled factors such as habitat availability. Indeed, it remains possible that the artificial flood, by washing fine sediment in between the gravels, favoured the visual isolation between 0⁺ fish since they are very small during their first weeks of growth (23 mm long and 0.1 g on average for a first feeding alevins) and visual isolation is a well know factor that limits the strength of the competition in salmonids (Huntingford et al. 1993; Imre et al. 2002).

Artificial flood and behaviour of alevins

Alevins were seen swimming actively more often in the cages impacted by the artificial flood than in the control ones. This is consistent with Biro et al. (2003) who highlighted that in low food conditions, young trout (*Oncorhynchus mykiss*) needed to increase their activity to maintain a constant supply of food and then, grow. Present results confirm that when resource is scarce, young salmonids exhibit more risky behaviour, increasing their foraging effort and probably increasing the predation risk in the same time.

Our behavioural observations were in accordance with current knowledge: social status matched with the number of feeding attempts and with individual weight gain. The dominant fish was the largest at the beginning of the experiment in only one of the four cages. It suggests that the body condition of fish (weight and/or size) is rather the consequence of dominance than the cause as suggested by Metcalfe (1986). In the behaviour cages, the average individual weight gain did not differ between control and impacted cages, but inter-individual differences were higher in the impacted cages. At low food abundance, dominants monopolized most of the resource leaving a limited access to food for subordinates (Maclean & Metcalfe 2001; Höjesjö et al. 2002). The relationship between fish aggressiveness and food abundance is not yet clear. It has previously been argued that the strength of territorial defence depended on resource availability, with highest levels of territorial defence observed at intermediate levels

of food (Toobaie and Grant 2013). Accordingly, when food is scarce or abundant, the energetic cost of aggressiveness is greater than the benefit. However, results diverge: Symons (1968) and Slaney & Northcote (1974) found that frequency of aggressive encounter was greater at low prey level and during starvation. On the contrary, van Leeuwen et al. (2016) showed that the number of aggressions decreased at low food levels. Subordinates adopted an “energy minimizing” strategy (Johnsson et al. 1996), whereby they did not try to have a good feeding position and they avoided the energetic costs of both swimming against the water flow and potential battles over territories (van Leeuwen et al. 2016). Because investment in territorial defence can be costly for growth, Metcalfe et al. (1986) emphasized that the optimum strategy adopted by subordinate was minimizing energetic expenditures rather than maximizing food intake. This way, subordinates may continue to grow despite receiving a low level of food intake. This hypothesis is clearly strengthened by our results.

Artificial flood and season

In early spring, survival of alevins remained low (50% on average) and they did not gain weight. The same trends was recorded in the observation cages, at least concerning survival. It seems that the low invertebrate abundances impacted significantly the performances of alevins and cause their death. In contrast, in late spring, survival was quite high (90% on average) and their growth was positive, even if much lower in impacted than in control cages. Early emerging fish have less competition pressure to access to the best feeding spots and they can grow efficiently as far as the productivity of the system is not a limiting factor. When late alevins emerge, early survivors are favoured by their residence anteriority on territories (Huntingford & Garcia de Leaniz 1997; Cutts et al. 1999; Harwood et al. 2003). However, the results from the present experiment suggest that if early emerging fish suffer from food shortage, they would undergo high mortality rates as well as negative growth.

Conclusion

Throughout the spring season, temperature as well as sunshine duration and brightness increase and boost the primary production. Correlatively, the invertebrate production increased, the recolonization process was more effective and it partially compensated the flood effects. Therefore, the impact of a scouring flood on invertebrates might depend to a great extent on its timing (Robinson et al. 2004). That timing has to be taken into account when assessing the impact of flood on 0⁺ trout since consequences on survival and growth were not of the same magnitude (harsh in early spring, light in late spring).

Supporting Information of the manuscript in preparation

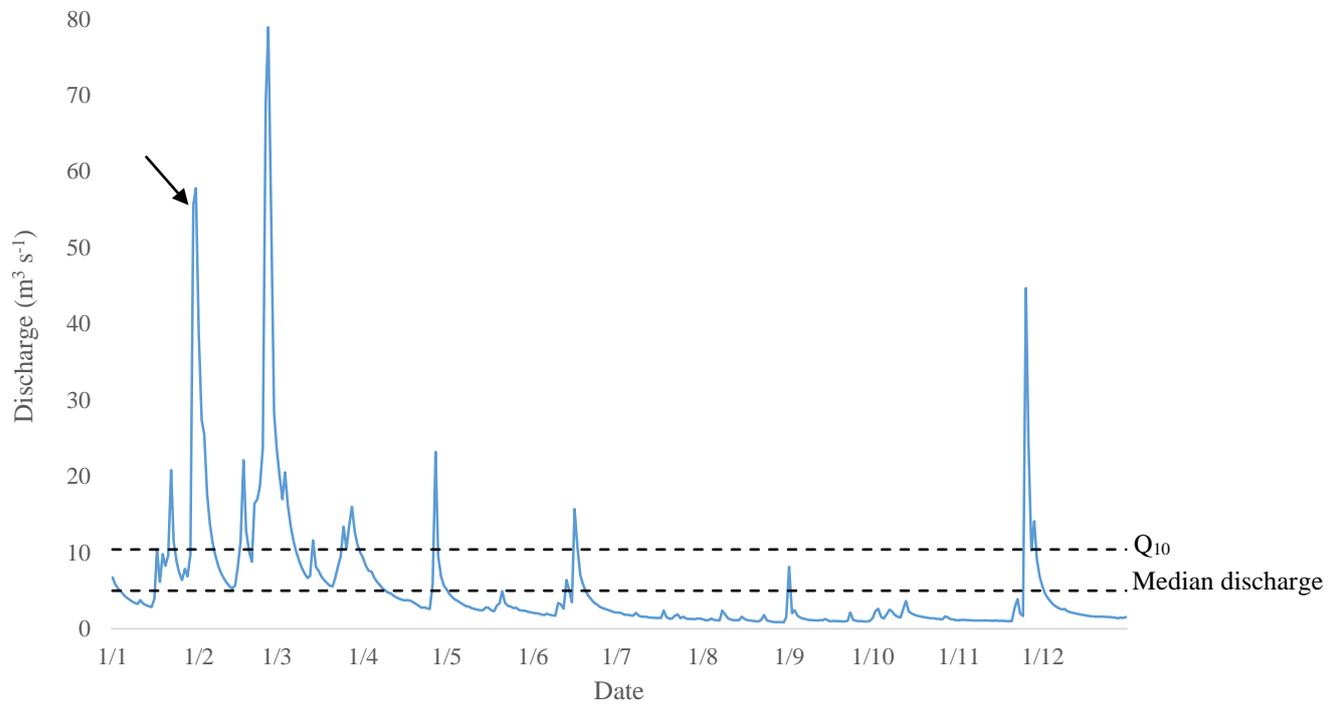


Figure S1: Discharge (in $m^3 s^{-1}$) on the Nivelle river during 2015. Water velocities were recorded on the Lapitxuri brook, a tributary to the Nivelle, the 30/01/2015. This date is represented by a dark arrow and dotted lines represent the Q_{10} and the median discharge of the Nivelle river.

Table S1: Water depth (in cm) and water velocities (in m s⁻¹) recorded on the Lapitxuri brook (the 30/01/2015).

Lapitxuri brook – 30/01/2015 – Nivelle discharge = 55.6 m³ s⁻¹												
	Section	1	2	3	4	5	6	7	8	9	10	11
	Water depth (cm)	13	13	30	32	38	47	50	62	70	50	30
Water velocity (m s ⁻¹) at	80% water depth	0.45	0.48	0.66	0.79	0.83	0.89	0.82	0.87	0.66	0.62	0.38
	40% water depth			0.55	0.63	0.82	0.87	1	1.06	0.83	0.72	0.37
	20% water depth			0.5	0.54	0.75	0.7	0.93	1.09	0.82	0.6	0.43

Table S2 Number of surveys, initial and final weight, individual weight gain, mean number of feeding attempts and of aggressive interactions of the alevins by survey during the first trial (early spring) according to their cage of origin and the colour of their tag.

Condition	Cage	VIE tag	Number of surveys	Initial weight (mg)	Final weight (mg)	Individual weight gain	Mean feeding attempts by survey				Mean number of aggressive interactions by survey		
							Short	Medium	Long	Total	Attacks made	Attacks sustained	Total
Control	1 Up	Green	40	110	146	0.327	3.40	4.28	3.43	11.10	2.18	0.03	2.20
		Orange	40	117	127	0.085	2.90	2.25	1.35	6.50	0.43	2.35	2.78
		Blue	13	86	-	-	0.54	0.00	0.08	0.62	0.00	0.69	0.69
		White	2	109	-	-	0.00	0.00	0.00	0.00	0.00	0.00	0.00
	2 Down	Green	40	121	131	0.083	3.25	3.03	2.68	8.95	0.45	0.25	0.70
		Orange	40	118	153	0.297	3.25	2.53	1.48	7.25	0.98	1.03	2.00
		Blue	36	120	-	-	2.33	1.39	0.89	4.61	0.22	0.33	0.56
		White	35	118	-	-	2.91	2.80	1.69	7.40	0.57	0.17	0.74
Flood	3 Up	Green	31	114	-	-	0.81	0.23	0.00	1.03	0.03	0.13	0.16
		Orange	39	98	112	0.143	3.21	3.28	1.67	8.15	1.00	0.03	1.03
		Blue	23	97	-	-	1.96	0.57	0.35	2.87	0.04	0.39	0.43
		White	35	107	187	0.748	2.31	1.94	1.51	5.77	0.14	0.74	0.89
	4 Down	Green	40	114	136	0.193	3.43	2.80	2.00	8.23	0.43	0.68	1.10
		Orange	40	115	141	0.226	4.48	3.15	2.00	9.63	1.30	0.38	1.68
		Blue	32	108	-	-	2.81	1.41	0.88	5.09	0.06	0.41	0.47
		White	35	110	-	-	3.06	3.03	2.26	8.34	1.40	0.51	1.91

Supporting information: Relationship between the invertebrate abundances and alevin performance

We assessed the strength of link between the mean abundances of invertebrates and (i) the survival, (ii) the average individual weight gain, and (iii) the fish productivity. We tested 60 relationships (10 invertebrate groups x 3 fish performance variables x 2 times, just after the disturbance and at the end of the trial). Correlations were tested with OpenBUGS®. If Y_j was the fish variable of the j^{th} cage, we assumed:

$$Y_j = a \text{ Abond}_j + b$$

Where Y_j is for each j cage, the number of survivors (S_j), the average individual weight gain ($WG_{j[i]}$) or the fish productivity (Prod.Fish_j), a is the slope coefficient, Abond_j is the average of abundance of invertebrates by cage and b is the intercept. Fit of correlation was calculated by dividing the variability unexplained by the variability observed in the data set, as follows:

$$\text{Fit} = 1 - \frac{\text{Variability unexplained by the model}}{\text{Variability of the data set}}$$

When the fit is between 0.5 and 1, the correlation is good. When the fit is between 0 and 0.5, the correlation is intermediate and when the fit is negative, there is no correlation.

Immediately after the flood, abundances of invertebrates fitted with fish performances in 23% of the cases (7/30, fit > 0.500 – Table S3). At the end of the trials, final invertebrate abundances fitted with fish performances in 63% of the cases (19/30, fit > 0.500 – Table S4). Usually positive, the links happen to be negative in five cases but only one link was significantly negative: between fish productivity and abundances of Diptera. Focusing on final Total abundance, and on Diptera and Ephemeroptera (the two groups among which are the main food items of newborn alevins), correlations with the weight gain of alevins were positive and significant (Fit = 0.888, 0.914 and 0.712 respectively – Figure S2). However, they were highly influenced by one point (Control cage in the second trial) characterized by abundances especially high. When this point was not taken into account, correlation was still significant for total abundance (Fit: 0.851; $a = 0.0015$; $P(a > 0) = 0.995$), but no more for Diptera and Ephemeroptera (Fit = -0.712 and -2.325, respectively).

Table S3 Correlations between fish variable (Y) and invertebrate abundances after the flood (X), mean of posterior probability distribution functions of a (the slope coefficient) and b (the intercept) and fit of the models just after flood simulation disturbance. Parameters in bold are significant (when $P(X > 0)$ is less than 0.10 or above 0.90).

Variable Y	Variable X	Groups	a	P(a > 0)	b	P(b > 0)	Equation	Fit
Number of surviving alevins by cage S	Number of invertebrates After the flood	Diptera	0.591	0.967	7.438	0.969	S = 0.591 Diptera + 7.438	0.574
		Trichoptera	-1.515	0.086	14.880	0.997	S = -1.515 Trichoptera + 14.88	0.580
		Ephemeroptera	0.201	0.820	8.719	0.957	S = 0.2005 Ephemeroptera + 8.719	0.313
		Mollusca	3.306	0.870	8.436	0.964	S = 3.306 Mollusca + 8.436	0.407
		Coleoptera	1.084	0.998	0.362	0.546	S = 1.084 Coleoptera + 0.3619	0.832
		Plecoptera	0.026	0.493	11.480	0.990	S = 0.02616 Plecoptera + 11.48	0.342
		Oligochaeta	0.026	0.515	11.100	0.971	S = 0.02557 Oligochaeta + 11.1	0.256
		Crustacea	0.128	0.772	9.943	0.982	S = 0.1276 Crustacea + 9.943	0.289
		Others	0.053	0.660	10.420	0.980	S = 0.05349 Others + 10.42	0.251
		Total	0.122	0.960	4.947	0.854	S = 0.1218 Total + 4.947	0.525
Mean Individual Weight Gain by cages WG	Number of invertebrates After the flood	Diptera	0.025	0.974	-0.168	0.223	WG = 0.02543 Diptera + -0.1681	0.459
		Trichoptera	0.844	0.922	-0.079	0.386	WG = 0.844 Trichoptera + -0.0787	0.076
		Ephemeroptera	0.000	0.500	0.251	0.713	WG = -0.0001604 Ephemeroptera + 0.2508	-0.649
		Mollusca	-0.023	0.469	0.280	0.737	WG = -0.02336 Mollusca + 0.2798	-0.641
		Coleoptera	-0.005	0.459	0.308	0.684	WG = -0.004779 Coleoptera + 0.3083	-0.637
		Plecoptera	1.022	0.766	0.080	0.598	WG = 1.022 Plecoptera + 0.08009	-0.386
		Oligochaeta	0.014	0.970	-0.143	0.261	WG = 0.01387 Oligochaeta + -0.1428	0.416
		Crustacea	-0.003	0.417	0.303	0.802	WG = -0.003036 Crustacea + 0.3032	-0.638
		Others	0.038	0.782	-0.037	0.467	WG = 0.03778 Others + -0.03725	-0.402
		Total	0.007	0.942	-0.404	0.166	WG = 0.00711 Total + -0.404	0.189
Fish Productivity by cages Prod	Number of invertebrates After the flood	Diptera	-0.536	0.172	11.800	0.952	Prod = -0.5361 Diptera + 11.8	0.040
		Trichoptera	4.990	1.000	0.312	0.628	Prod = 4.99 Trichoptera + 0.3116	0.978
		Ephemeroptera	0.428	0.898	3.072	0.701	Prod = 0.4279 Ephemeroptera + 3.072	-0.034
		Mollusca	-1.819	0.332	9.490	0.921	Prod = -1.819 Mollusca + 9.49	-0.111
		Coleoptera	-0.522	0.226	13.490	0.925	Prod = -0.5224 Coleoptera + 13.49	0.068
		Plecoptera	14.100	0.969	3.602	0.805	Prod = 14.1 Plecoptera + 3.602	0.503
		Oligochaeta	-0.152	0.312	9.922	0.913	Prod = -0.1516 Oligochaeta + 9.922	-0.159
		Crustacea	-0.196	0.226	9.917	0.940	Prod = -0.1961 Crustacea + 9.917	-0.116
		Others	0.288	0.941	3.862	0.778	Prod = 0.2876 Others + 3.862	0.117
		Total	-0.264	0.010	25.580	0.997	Prod = -0.2642 Total + 25.58	0.819

Table S4 Correlations between fish variable (Y) and invertebrate abundances at the end of the trials (X), mean of posterior probability distribution functions of a (the slope coefficient) and b (the intercept) and fit of the models at the end of the trials. Parameters in bold are significant (when $P(X > 0)$ is less than 0.10 or above 0.90).

Variable Y	Variable X	Groups	a	P(a > 0)	b	P(b > 0)	Equation	Fit
Number of surviving alevins by cage S	Number of invertebrates At the end of the trials	Diptera	0.094	0.887	10.490	0.986	S = 0.09432 Diptera + 10.49	0.649
		Trichoptera	-0.356	0.323	14.440	0.998	S = -0.3557 Trichoptera + 14.44	0.621
		Ephemeroptera	-0.002	0.455	13.750	0.997	S = -0.002252 Ephemeroptera + 13.75	0.582
		Mollusca	0.667	0.691	12.030	0.988	S = 0.6673 Mollusca + 12.03	0.581
		Coleoptera	0.611	0.915	7.091	0.894	S = 0.6105 Coleoptera + 7.091	0.654
		Plecoptera	3.934	0.850	10.480	0.988	S = 3.934 Plecoptera + 10.48	0.661
		Oligochaeta	0.221	0.956	9.253	0.984	S = 0.2205 Oligochaeta + 9.253	0.727
		Crustacea	-0.090	0.254	14.930	0.998	S = -0.08984 Crustacea + 14.93	0.648
		Others	-0.005	0.401	13.960	0.997	S = -0.004778 Others + 13.96	0.598
		Total	0.052	0.875	8.776	0.945	S = 0.05168 Total + 8.776	0.627
Mean Individual Weight Gain by cages WG	Number of invertebrates At the end of the trials	Diptera	0.011	0.999	-0.602	0.006	WG = 0.01121 Diptera + -0.6015	0.914
		Trichoptera	0.188	1.000	-0.083	0.107	WG = 0.1882 Trichoptera + -0.08306	0.939
		Ephemeroptera	0.090	0.991	-0.296	0.076	WG = 0.08964 Ephemeroptera + -0.2962	0.712
		Mollusca	-0.483	0.090	0.978	0.943	WG = -0.4834 Mollusca + 0.9781	0.027
		Coleoptera	-0.006	0.469	0.323	0.633	WG = -0.006134 Coleoptera + 0.3234	-0.631
		Plecoptera	-0.442	0.146	0.545	0.923	WG = -0.442 Plecoptera + 0.5448	-0.197
		Oligochaeta	0.005	0.994	-0.169	0.123	WG = 0.005161 Oligochaeta + -0.1686	0.773
		Crustacea	0.017	0.904	-0.221	0.284	WG = 0.01688 Crustacea + -0.2214	-0.005
		Others	0.110	0.971	-0.742	0.068	WG = 0.1102 Others + -0.7421	0.425
		Total	0.003	0.998	-0.427	0.012	WG = 0.003139 Total + -0.4272	0.888
Fish Productivity by cages Prod	Number of invertebrates At the end of the trials	Diptera	-0.333	0.014	21.350	0.996	Prod = -0.3325 Diptera + 21.35	0.724
		Trichoptera	5.049	1.000	-0.080	0.466	Prod = 5.049 Trichoptera + -0.07976	0.977
		Ephemeroptera	0.188	0.996	2.488	0.767	Prod = 0.1877 Ephemeroptera + 2.488	0.657
		Mollusca	3.988	0.965	-0.116	0.489	Prod = 3.988 Mollusca + -0.1155	0.267
		Coleoptera	0.142	0.555	6.813	0.783	Prod = 0.1418 Coleoptera + 6.813	-0.217
		Plecoptera	4.670	0.772	4.737	0.771	Prod = 4.67 Plecoptera + 4.737	-0.062
		Oligochaeta	-0.180	0.260	11.400	0.924	Prod = -0.1801 Oligochaeta + 11.4	-0.049
		Crustacea	-0.353	0.140	12.100	0.962	Prod = -0.3531 Crustacea + 12.1	0.094
		Others	0.127	0.999	1.887	0.744	Prod = 0.1274 Others + 1.887	0.761
		Total	-0.099	0.132	17.900	0.955	Prod = -0.09919 Total + 17.9	0.313

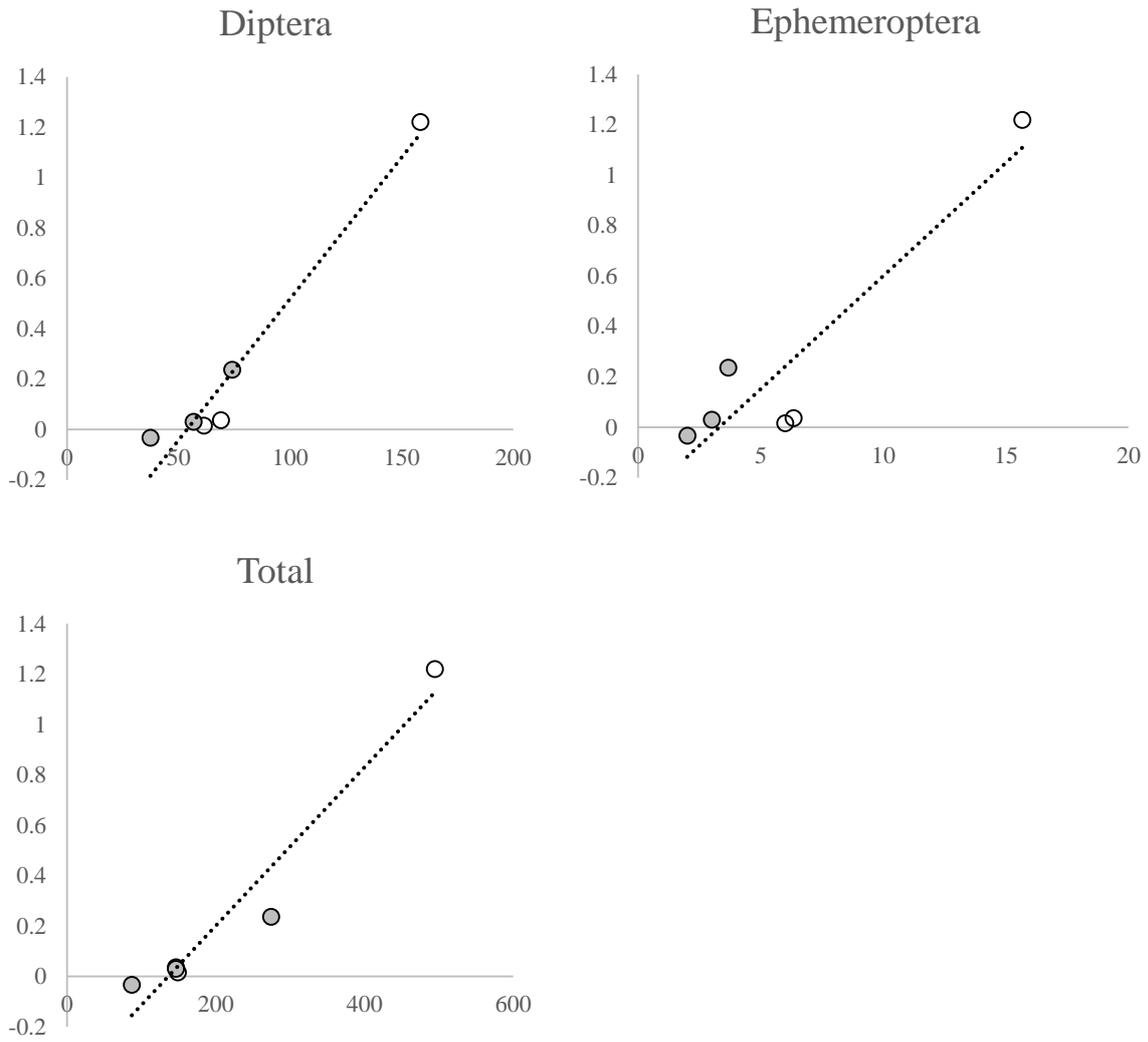


Figure S2 Average individual weight gain of alevins by cage according to the average abundance of Diptera, Ephemeroptera and the total invertebrates by cage at the end of the trials. Empty circles represent the control cages, grey circles the impacted cages and the dotted line the correlation.

Chapter IV Consequences of Global Climate Change on the community of invertebrates, on the survival and growth of first-feeding alevins at low density: flood simulation in a semi-natural environment

Introduction

The previous experiment emphasised that hydrological disturbances decreased the food availability for fish but the consequences at the fish level depended on the productivity of the system. These results stemmed out from an experiment at high fish density (32 fish m⁻²) and one may wonder on the persistence of these effects on the fish population at low alevins density. According to literature, mortality rate at low density is mainly density-independent while density-dependent growth still occurs (Jenkins et al. 1999; Grant & Imre 2005; Imre et al. 2005).

A protocol was set up in the experimental channel of the Lapitxuri in late spring to look at the performances of alevins at low density. A flood was simulated in four half-reaches, while four others were not impacted and used as control. Immediately after the flood, invertebrates were sampled and just-emerged brown trout alevins were introduced in all reaches at low fish density (2.6 fish m⁻²). After 33 days, the surviving alevins were recovered and invertebrates were sampled. We aimed to test the following hypotheses:

1. Flood should diminish the abundance of invertebrates,
2. Flood should diminish growth of alevins, but not survival,
3. A relationship between the abundance of invertebrates and growth is awaited.

Material and Methods

The experimental channel of the Lapitxuri and experimental design

The experiment was conducted from April to the end of May 2015 in an experimental channel fed by the Lapitxuri brook, a tributary of the Nivelle river in south-western France (43°16' N, 1°28' W). Four reaches (10 m long and 2.8 m wide) were used for the experiment, two located at 30 meters and two at 80 meters downstream the flow entry (Figure 4.1). Each reach was divided longitudinally by a tarpaulin wall in two half-reaches of 14 m² (10 m long and 1.4 m wide). Upstream and downstream movements of fish were prevented by nets, and traps at the end of downstream nets allowed to catch drifting fish. Aerial nets protected fish from avian predation. Gravel bed was constituted by cobbles and pebbles, substrate particles ranging approximately from 1 to 5 cm. The average mid-water velocity was maintained during the whole experiment at 12.4 cm s⁻¹ (SD = 3.9 – calculation based on the absolute value of velocity measurements) and the mean water temperature during the experiment was 13.34°C (SD = 1.04). Abiotic conditions in the channel were close to optimal environment for young trout in terms of substratum quality, water quality and temperature values (Roussel & Bardonnnet 1997,

2002; Heggenes et al. 1999). Food supply was provided by invertebrate drift from the Lapitxuri brook and by the production of the channel itself. To recover fish from a previous experiment, an electrofishing was conducted in the totality of the channel on 14/04/2015. On 15/04/2015, a flood was simulated with a motor-pump in the 4 left-bank reaches (Figure 4.1). The motor-pump was used at the extreme right point during 5 seconds, then the pump was displaced by 20 cm to the left and used again during 5 seconds. This procedure was repeated over the entire width of the impacted reach (7 times) and such transects were made every 50 cm to cover the entire length of each impacted reach. The use of the motor-pump increased the water velocity up to 92.2 cm s^{-1} (SD = 18.9) on average, ranging from 45 to 146 cm s^{-1} in the 50 cm in front of the outflow. After the flood simulation, the gravel bed was flattened again.



Figure 4.1 Experimental design set up in the semi-natural channel of the Lapitxuri. Eight reaches were used. Four reaches were impacted by the flood (left bank – in grey) and four reaches served as controls (right bank – in white).

Fish sampling

On 16/01/2015, eggs were obtained through the artificial fertilization of gametes of wild brown trout caught in the Nivelles watershed (43°21' N, 1°33' W). Eggs and alevins were reared at 8.33°C (SD = 1.40) until complete yolk sac depletion (752 degree-days; survival: 92%). On 16/04/2015, 40 randomly selected alevins were individually weighed and measured to

characterize the biometry of the batch. Then, remaining individuals were randomly split into eight groups of 37 alevins and released in the eight experimental reaches of the Lapitxuri semi-natural stream. The fish density in each reach was of 2.6 fish m⁻² (~8% of the density used for the previous experiment). Downstream traps were checked every morning, and after counting, fish were immediately released back into the reach they came from. After 33 days of growth, surviving alevins were recovered by successive removal of electrofishing (19–22 May). They were weighted, measured and subjected to a benzocaine lethal anesthesia. Eight alevins of each reach were randomly selected and all invertebrates contained in their stomachs were identified to family level when possible and counted. Then a total of sixty-four gut contents were analysed.

Benthic invertebrate sampling

Immediately after the flood simulation (15/04/2015) and at the end of the trial (19/05/2015), 3 invertebrate samples by reach were randomly collected using a Surber net (30 cm x 30 cm, 500 µm mesh). Samples were preserved in 70% ethanol. Invertebrates were identified and assigned to different groups: Diptera, Trichoptera, Ephemeroptera, Mollusca, Coleoptera, Plecoptera, Oligochaeta, Crustacea and Exogenous.

Statistical analyses

Abundance of invertebrates as well as the logit of the alevins survival probability, alevins individual weight gain and fish productivity (sum of final weights) were analysed using a bayesian modelling approach computed with OpenBUGS®. All the models were built with the same logic: we assumed that the variables of interest followed a normal distribution. Then, the mean of the variable (μ .Variable) was modelled according to two parameters: a fixed-effect relative to the flood (α) and a random effect relative to each j reach (β_j), which integrated all the potential sources of variation other than the flood. The number of prey ingested by alevin followed a Poisson law and the mean (μ .Prey) was modelled according to these same two effects (α and β_j).

Abundance of invertebrates

If $Abond_i$ is the number of invertebrates counted by sample, we assume:

$$\mu.\log(Abond_i + 1)_i = \mu + \alpha Flood_i + \beta_{j[i]}$$

Where μ is the average of the logarithm of the invertebrate abundance, α is a fixed-effect parameter for the impact of the flood and β is a random effect corresponding to each j reach. The same modelling treatment was applied to the total abundance of invertebrates as well as to

the abundance of each group identified (*i.e.* Diptera, Trichoptera, Ephemeroptera, Mollusca, Coleoptera, Plecoptera, Oligochaeta and Crustacea) and for exogenous individuals.

Survival probability of alevins

For each j reach, number of survivors S_j is estimated from the number of fish caught at each electrofishing passage ($C1_j$, $C2_j$, $C3_j$ and $C4_j$), the number of remaining fish after each passage ($R1_j$, $R2_j$ and $R3_j$) and the fish efficiency ($p.F_j$):

$$C1_j \sim \text{Binomial}(p.F_j, S_j)$$

$$R1_j = S_j - C1_j ; C2_j \sim \text{Binomial}(p.F_j, R1_j)$$

$$R2_j = R1_j - C2_j ; C3_j \sim \text{Binomial}(p.F_j, R2_j)$$

$$R3_j = R2_j - C3_j ; C4_j \sim \text{Binomial}(p.F_j, R3_j)$$

Then, if $p.S_j$ is the probability of survival in the j reach and N_j the number of fish introduced in each reach at the beginning of the experiment, we assume:

$$S_j \sim \text{Binomial}(p.S_j, N_j)$$

$$\mu.\text{Logit}(p.S_j) = \mu + \alpha \text{Flood}_j + \beta_j$$

Where μ is the average of the logit survival probability, α is a fixed-effect parameter for the impact of the flood and β is a random effect corresponding to each j reach.

Individual weight gain of alevins

The initial weight (IW) of the 40 alevins sub-sampled at the beginning of the experiment allowed to estimate the mean ($\mu.IW$) and the standard deviation ($\sigma.IW$) of the alevins initial weight distribution. For each k survivor recovered at the end, its initial weight ($W.Beg_k$) was drawn in the alevin initial weight distribution (according to $\mu.IW$ and $\sigma.IW$), while its final weight was known ($W.End_k$). By j reach, the standard deviation of the final weight of alevins ($\sigma.W.End_j$) was also modelled to test if the flood affected the variability of alevins final weight.

Individual weight gain (WG_k) was obtained by dividing the difference between the final weight and the initial weight by the initial weight. Then, we assume:

$$\mu.WG_k = \mu + \alpha \text{Flood}_k + \beta_{j[k]}$$

Where μ is the average weight gain of alevins, α is a fixed-effect parameter for the impact of the flood and β is a random effect corresponding to each j reach.

Fish productivity

For each j reach, the final weights were summed to obtain the fish production per reach (Prod.Fish $_j$).

$$\mu. Prod. Fish_j = \mu + \alpha Flood_j + \beta_j$$

Where μ is the average fish production, α is a fixed-effect parameter for the impact of the flood and β is a random effect corresponding to each j reach.

Relationship between the community of invertebrate and alevins

We linked (i) the number of surviving alevins by reach, (ii) the average individual weight gain of alevins by reach and (iii) fish productivity by reach according to the average invertebrate abundance by reach immediately after the flood or at the end of the experiment (33 days after). Then, 60 relationships were tested (10 invertebrate groups x 3 variables for alevin performances x 2 times). Correlations were tested with OpenBUGS®. If Y_j was the variable of the j^{th} reach, we assume:

$$Y_j = a Abond_j + b$$

Where Y_j is the number of survivors by reach (S_j), the fish weight gain by reach ($WG_{j[i]}$) or the fish productivity by reach (Prod.Fish $_j$), a is the slope coefficient, $Abond_j$ is the average invertebrate abundance by reach and b is the intercept.

Fit of the correlation is calculated by dividing the variability unexplained by the correlation model with the variability observed in the data set, as follows:

$$Fit = 1 - \frac{Variability\ unexplained\ by\ the\ model}{Variability\ of\ the\ data\ set}$$

When the fit is negative, there is no correlation. When the fit is between 0 and 0.5, the correlation is intermediate. When the fit is between 0.5 and 1, the correlation is good.

Gut contents

For the description of the diet, data were pooled according to conditions (control vs. impacted). The total number of prey (N), the relative abundance (A) and the occurrence of prey in fish (F, where the number of alevins containing this kind of prey is divided by the total number of alevins sampled) were calculated for each family.

Total number of prey ingested by alevins and the number of individuals of Chironomidae and Baetidae families (which are the main food items just-emerged alevins) were analysed. Then, if $Prey_l$ is the number of prey ingested by the l^{th} alevins, we assume:

$$Prey_l \sim Poisson(\mu \cdot Prey_l)$$

$$\log(\mu \cdot Prey_l) = \mu + \alpha Flood_j + \beta_j$$

Where μ is the average number of prey ingested, α is a fixed-effect parameter for the impact of the flood and β is a random effect corresponding to each j reach.

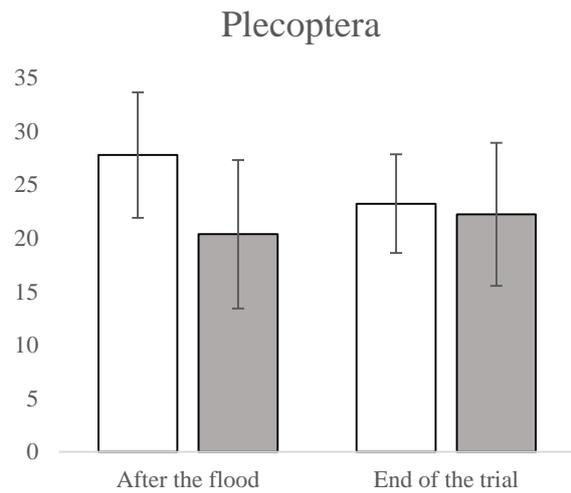
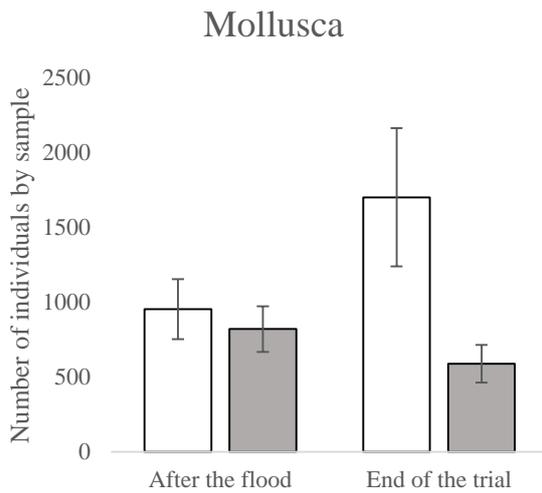
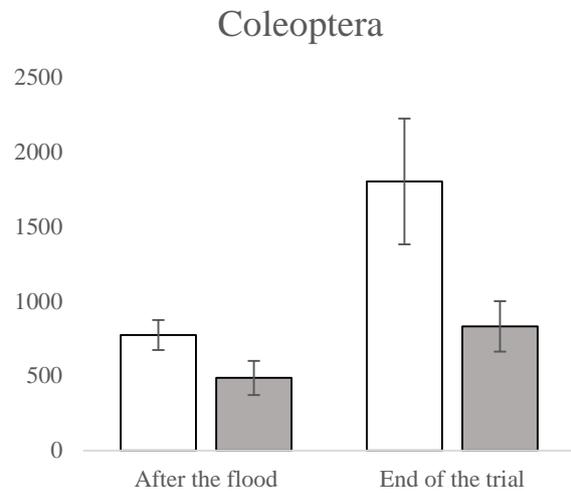
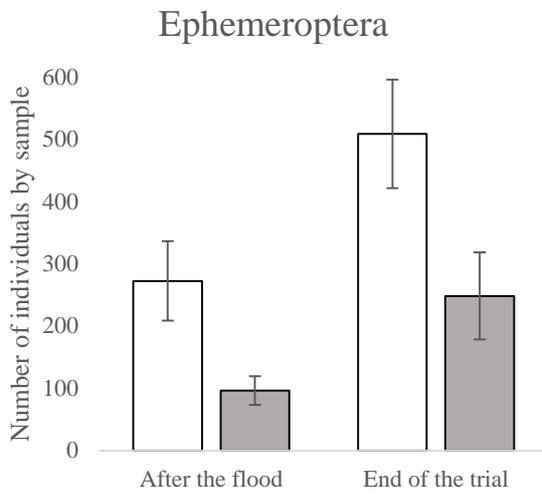
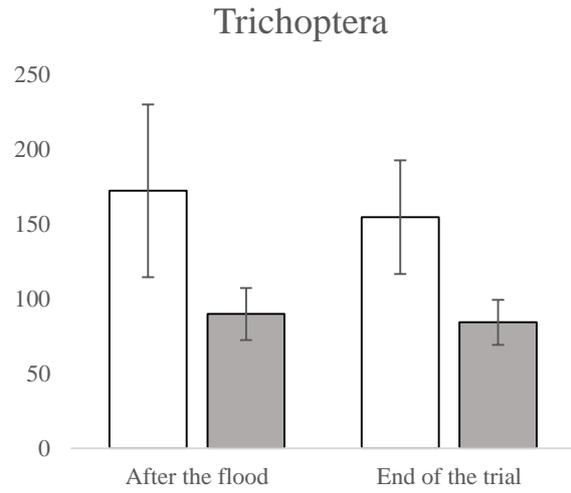
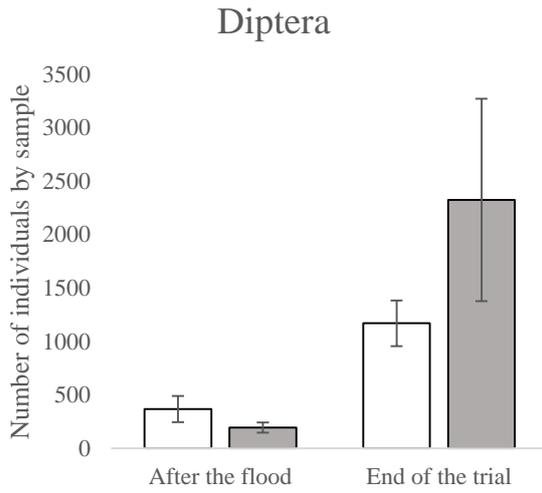
Bayesian computations

Parameters were given independent “weakly informative” priors. Following recommendations of Gelman & Hill (2007), μ , α , $\mu.IW$, $\sigma.IW$, a and b were sampled in a normal distribution, $N(0, 100)$. β_j were drawn in a normal distribution, $N(0, \sigma_\beta^2)$, with σ_β sampled in a truncated half-Cauchy distribution, $t(0, 1, 1)$. $p.F_j$ were sampled in a beta distribution, $beta(1, 1)$. Hyperparameters of $\sigma.W.End_j$ ($B.\sigma.W.End$ and $E.\sigma.W.End$) were sampled in a gamma distribution, $gamma(1, 1)$. We used each time three independent chains, the first 10 000 iterations were discarded as an initial burn-in period. Then, 10 000 further iterations (resulting from one every ten runs) were performed. The convergence of the chains to their ergodic distribution was tested *via* the Gelman-Rubin (GR) diagnostics integrated in OpenBUGS®. The significance of the parameters was tested with the step function implemented in OpenBUGS®. At each iteration for a variable X , $step(X)$ equaled 1 if $X \geq 0$ and equaled 0 if $X < 0$. At the end of the run, if $P(X > 0)$ was lower than 0.1 or higher than 0.9, the parameter X was considered to be different from 0.

Results

Effect of the flood on the community of invertebrates

Immediately after the artificial flood, most invertebrate groups and the total abundance of invertebrates were negatively impacted (except Mollusca – Figure 4.2). However, this tendency was never significant (except for Coleoptera ($P(\alpha_1 > 0) = 0.054$, Table 4.1). At the end of the trial, all groups (except Diptera) were still less abundant in the impacted reaches, but again differences were not significant, (except for Ephemeroptera and Mollusca ($P(\alpha_2 > 0) = 0.037$ and 0.052 respectively).



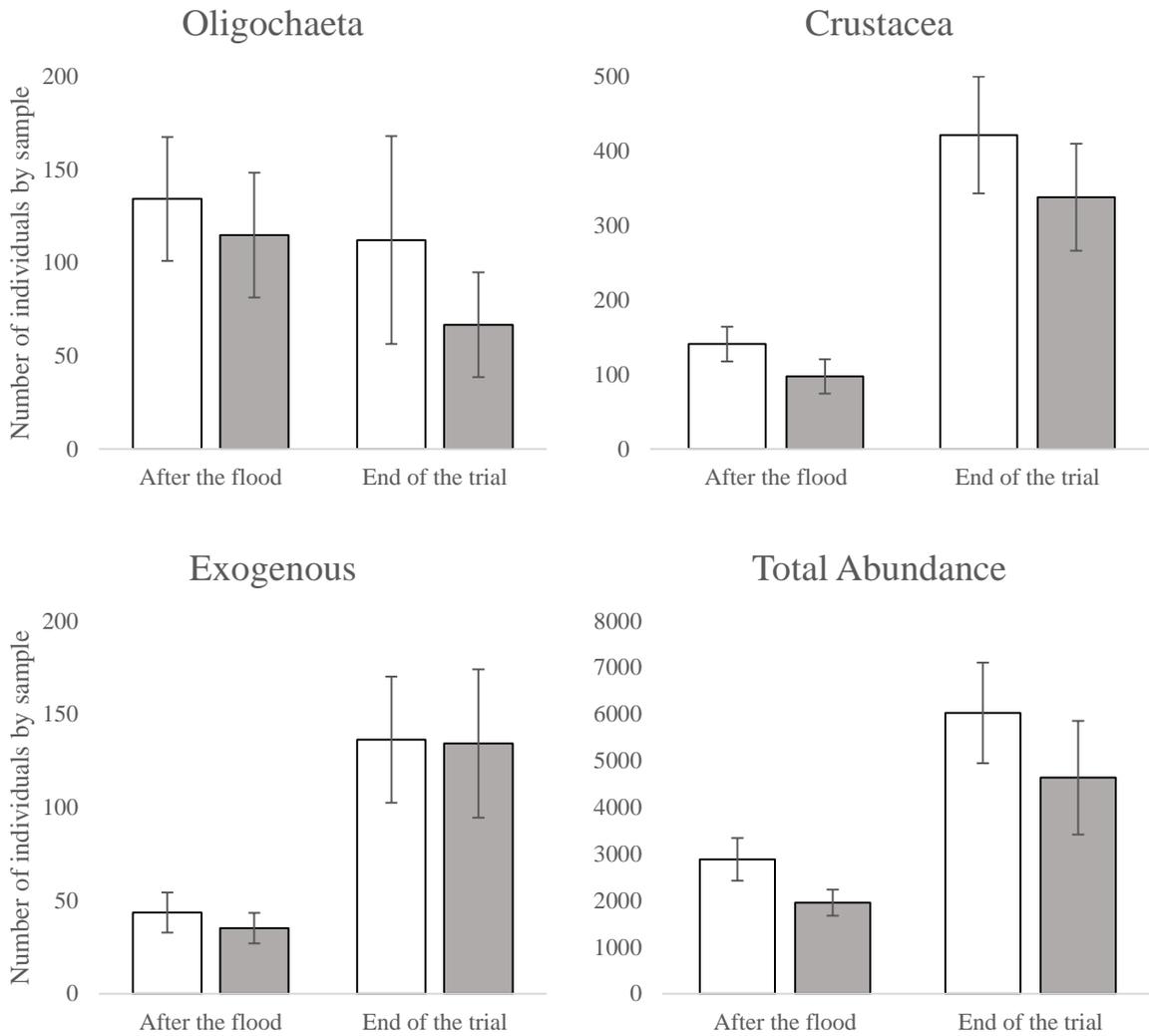


Figure 4.2 Number of individuals per sample (average \pm standard error) of the groups of invertebrates identified according to treatment (Control in white and Impacted by flood in grey) immediately and at the end of the trial.

Table 4.1 Mean of the posterior probability distribution functions of α , the effect of flood on the logarithm of the abundance of invertebrates. The effect was estimated for each group and for the total abundance, immediately after the flood and at the end of the trial. Parameters were considered significant when $P(X > 0)$ is less than 0.10 or above 0.90 (in bold).

Groups	After the flood		End of the trial	
	α_1	$P(\alpha_1 > 0)$	α_2	$P(\alpha_2 > 0)$
Diptera	-0.375	0.264	0.078	0.553
Trichoptera	-0.440	0.163	-0.724	0.117
Ephemeroptera	-0.844	0.156	-1.307	0.037
Mollusca	0.015	0.512	-1.282	0.052
Coleoptera	-0.680	0.054	-0.737	0.122
Plecoptera	-0.500	0.246	-0.565	0.208
Oligochaeta	0.014	0.503	-0.763	0.232
Crustacea	-0.185	0.370	-0.616	0.196
Exogenous	0.060	0.545	-0.402	0.281
Total	-0.301	0.186	-0.519	0.168

Effect of the flood on alevins

Survival probability reached 84% in the control and only 63% in the impacted reaches (Figure 4.3). At the beginning of the experiment, the batch of 40 alevins weighted on average 0.104 g and the SD was low (0.004). At the end of the trial (33 days of growth), fish were lighter in the control (0.459 g on average), than in the impacted reaches (0.505 g). Accordingly, weight gain was lower in the control (3.411) than in the impacted reaches (3.854). In addition, alevins weights were more homogenous in the impacted reaches ($SD_{F1} = 0.060$, $SD_{F2} = 0.065$, $SD_{F3} = 0.087$ and $SD_{F4} = 0.090$), than in the control ones ($SD_{C1} = 0.098$, $SD_{C2} = 0.104$, $SD_{C3} = 0.092$ and $SD_{C4} = 0.113$). Finally, fish productivity reached on average 13.23 g in control reaches and only 11.31 g in the impacted ones.

To sum up, alevin survival probability and fish productivity were significantly lower in the impacted reaches than in the control ones ($P(\alpha > 0) = 0.012$ and 0.084, respectively – Table 4.2), while individual weight gain was higher ($P(\alpha > 0) = 0.907$).

Table 4.2 Mean of the posterior probability distribution functions of α , the effect of flood on the survival probability, on individual weight gain of alevins and on fish productivity. Parameters were considered significant when $P(X > 0)$ is less than 0.10 or above 0.90 (in bold).

Effect of the flood (α) on ...	Mean	$P(\alpha > 0)$
Survival probability	-1.169	0.012
Weight gain	0.443	0.907
Productivity	-1.923	0.084

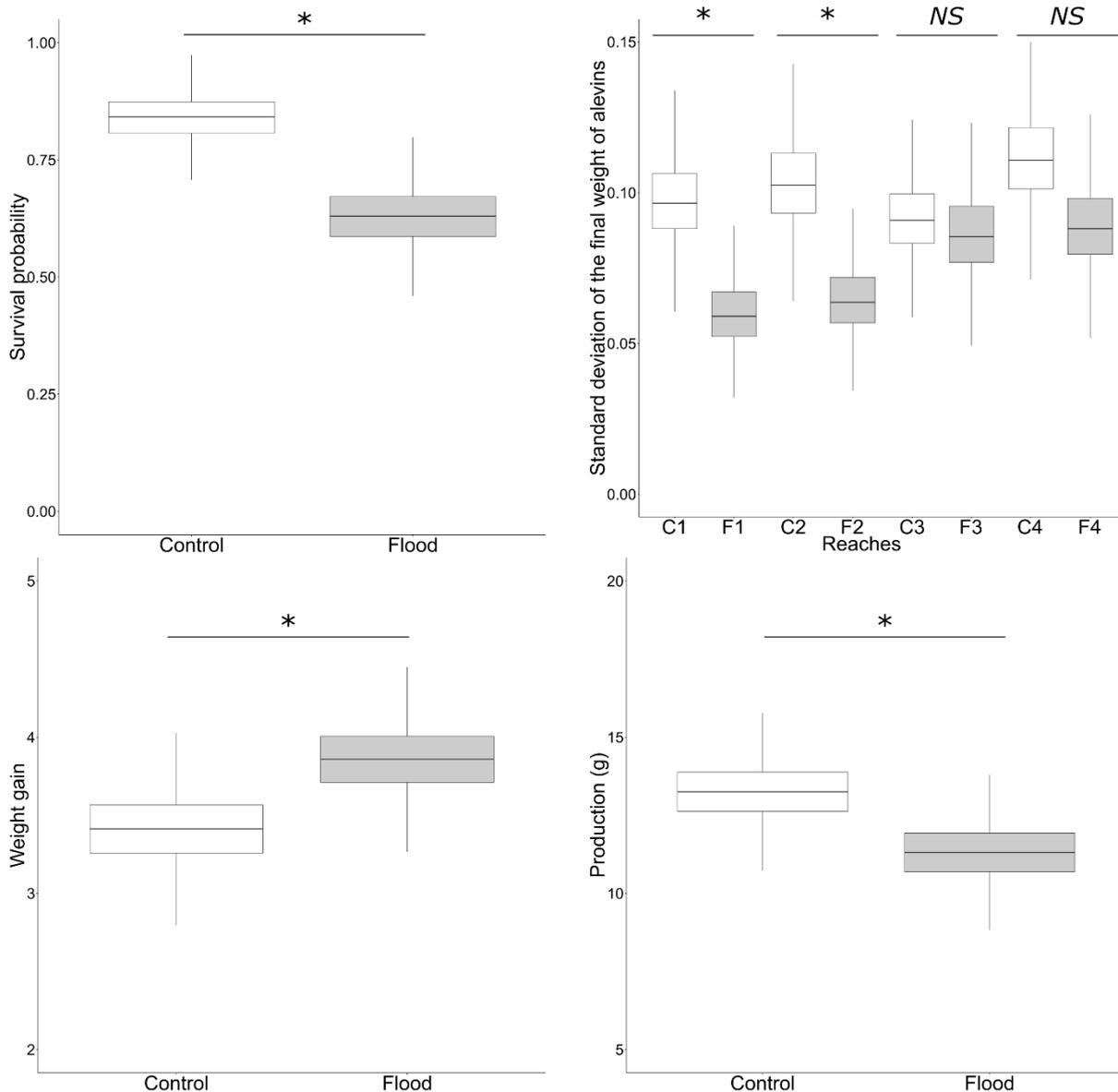


Figure 4.3 Model estimates of the survival probability, of standard deviation of the final weight, of weight gain of alevins and of fish productivity of control (in white) and impacted by flood reaches (in grey). Boxplots indicate the 1, 25, 50, 75 and 99 percentiles of posterior distributions. Significant differences are shown by *.

Relationship between the community of invertebrate and alevins

Alevins performances (number of survivors, average individual weight gain, fish productivity) were related to the abundances of each invertebrate group (10 groups: Diptera, Trichoptera, Ephemeroptera, Mollusca, Coleoptera, Plecoptera, Oligochaeta, Crustacea, Exogenous and Total) both immediately after the flood (Table 4.3) and at the end of the trial (Table 4.4), then 60 relationships were tested. On the 60 relationships tested, 45 fits of correlations were negative and 15 fits were positive but lower than 0.50. This meant that the link between the variables tested was low or inexistent. Focusing on Diptera and Ephemeroptera orders, despite the poor

quality of the correlations, on 12 relationships, the slope coefficient was significantly positive five times ($P(a > 0) > 0.9$) and significantly negative once ($P(a > 0) < 0.1$), leading to the following hypotheses: the initial abundance of Diptera and Ephemeroptera increased the survival and the productivity of fish (Table 4.3). However, the growth of alevins decreased the abundance of Ephemeroptera at the end of the trial (Table 4.4).

Table 4.3 Correlations between invertebrate abundances immediately after the flood (X) and fish variable (Y), mean of posterior probability distribution functions of a (the slope coefficient) and b (the intercept) and fit of the models. Parameters were considered significant when $P(X > 0)$ is less than 0.10 or above 0.90 (in bold).

Fish variable (Y)	Invertebrate variable (X)	Groups	a	P(a > 0)	b	P(b > 0)	Equations	Fit
Number of surviving alevins by reach S	Abundance of invertebrates by reach Immediately after the flood	Diptera	0.009	0.912	23.890	1.000	S = 0.008572 Diptera + 23.89	-0.198
		Trichoptera	0.028	0.951	22.680	1.000	S = 0.02825 Trichoptera + 22.68	-0.127
		Ephemeroptera	0.016	0.967	23.510	1.000	S = 0.01583 Ephemeroptera + 23.51	0.034
		Mollusca	0.008	0.943	19.440	0.996	S = 0.007867 Mollusca + 19.44	-0.400
		Coleoptera	0.011	0.973	19.530	0.998	S = 0.01103 Coleoptera + 19.53	-0.164
		Plecoptera	0.198	0.952	21.610	0.999	S = 0.198 Plecoptera + 21.61	-0.191
		Oligochaeta	0.040	0.917	21.340	0.999	S = 0.03996 Oligochaeta + 21.34	-0.372
		Crustacea	0.067	0.990	18.700	0.999	S = 0.06662 Crustacea + 18.7	0.068
		Exogenous	0.102	0.915	22.290	1.000	S = 0.1021 Exogenous + 22.29	-0.324
		Total	0.003	0.969	20.230	0.999	S = 0.002573 Total + 20.23	-0.167
Average individual weight gain of alevins by reach WG	Abundance of invertebrates by reach Immediately after the flood	Diptera	0.000	0.750	3.518	1.000	WG = 0.0004204 Diptera + 3.518	-0.236
		Trichoptera	0.001	0.618	3.567	1.000	WG = 0.0005287 Trichoptera + 3.567	-0.333
		Ephemeroptera	0.000	0.616	3.587	1.000	WG = 0.0002745 Ephemeroptera + 3.587	-0.326
		Mollusca	0.000	0.838	3.226	1.000	WG = 0.0004618 Mollusca + 3.226	-0.108
		Coleoptera	0.000	0.663	3.477	1.000	WG = 0.0002509 Coleoptera + 3.477	-0.304
		Plecoptera	0.001	0.537	3.610	1.000	WG = 0.001053 Plecoptera + 3.61	-0.344
		Oligochaeta	0.000	0.554	3.589	1.000	WG = 0.0003731 Oligochaeta + 3.589	-0.337
		Crustacea	0.000	0.511	3.628	1.000	WG = 0.00007267 Crustacea + 3.628	-0.346
		Exogenous	0.007	0.835	3.374	1.000	WG = 0.006682 Exogenous + 3.374	-0.121
		Total	0.000	0.738	3.417	1.000	WG = 0.00009113 Total + 3.417	-0.258
Fish productivity by reach Prod	Abundance of invertebrates by reach Immediately after the flood	Diptera	0.003	0.950	11.440	1.000	Prod = 0.003001 Diptera + 11.44	0.159
		Trichoptera	0.009	0.956	11.160	1.000	Prod = 0.008547 Trichoptera + 11.16	0.177
		Ephemeroptera	0.005	0.979	11.330	1.000	Prod = 0.00519 Ephemeroptera + 11.33	0.344
		Mollusca	0.002	0.942	10.320	1.000	Prod = 0.002216 Mollusca + 10.32	0.088
		Coleoptera	0.003	0.964	10.360	1.000	Prod = 0.003065 Coleoptera + 10.36	0.204
		Plecoptera	0.044	0.886	11.220	1.000	Prod = 0.04409 Plecoptera + 11.22	-0.035
		Oligochaeta	0.007	0.801	11.380	1.000	Prod = 0.007145 Oligochaeta + 11.38	-0.173
		Crustacea	0.017	0.968	10.280	1.000	Prod = 0.01685 Crustacea + 10.28	0.224
		Exogenous	0.032	0.941	11.010	1.000	Prod = 0.03226 Exogenous + 11.01	0.106
		Total	0.001	0.973	10.410	1.000	Prod = 0.0007759 Total + 10.41	0.271

Table 4.4 Correlations between invertebrate abundances at the end of the trial (X) and fish variable (Y), mean of posterior probability distribution functions of a (the slope coefficient) and b (the intercept) and fit of the models. Parameters were considered significant when $P(X > 0)$ is less than 0.10 or above 0.90 (in bold).

Fish variable (Y)	Invertebrate variable (X)	Groups	a	P(a > 0)	b	P(b > 0)	Equations	Fit
Number of surviving alevins by reach S	Abundance of invertebrates by reach At the end of the trial (33 days)	Diptera	-0.001	0.213	27.850	1.000	S = -0.0008931 Diptera + 27.85	-0.046
		Trichoptera	0.037	0.954	22.060	1.000	S = 0.03713 Trichoptera + 22.06	-0.164
		Ephemeroptera	0.017	0.994	20.400	1.000	S = 0.01676 Ephemeroptera + 20.4	0.250
		Mollusca	0.004	0.993	21.840	1.000	S = 0.004215 Mollusca + 21.84	0.283
		Coleoptera	0.002	0.939	23.050	1.000	S = 0.002463 Coleoptera + 23.05	-0.172
		Plecoptera	0.262	0.877	20.470	0.995	S = 0.2616 Plecoptera + 20.47	-0.566
		Oligochaeta	0.019	0.808	24.520	1.000	S = 0.01855 Oligochaeta + 24.52	-0.339
		Crustacea	0.016	0.934	20.260	0.997	S = 0.01626 Crustacea + 20.26	-0.393
		Exogenous	0.017	0.756	23.820	0.999	S = 0.0173 Exogenous + 23.82	-0.475
		Total	0.001	0.897	21.930	0.999	S = 0.0008152 Total + 21.93	-0.415
Average individual weight gain of alevins by reach WG	Abundance of invertebrates by reach At the end of the trial (33 days)	Diptera	0.000	0.851	3.412	1.000	WG = 0.0001282 Diptera + 3.412	-0.095
		Trichoptera	-0.002	0.128	3.916	1.000	WG = -0.00239 Trichoptera + 3.916	-0.046
		Ephemeroptera	-0.001	0.081	4.015	1.000	WG = -0.001018 Ephemeroptera + 4.015	0.104
		Mollusca	0.000	0.029	4.007	1.000	WG = -0.0003253 Mollusca + 4.007	0.344
		Coleoptera	0.000	0.057	3.937	1.000	WG = -0.0002251 Coleoptera + 3.937	0.185
		Plecoptera	0.008	0.659	3.454	1.000	WG = 0.008094 Plecoptera + 3.454	-0.307
		Oligochaeta	-0.002	0.171	3.794	1.000	WG = -0.001739 Oligochaeta + 3.794	-0.129
		Crustacea	-0.001	0.186	3.966	1.000	WG = -0.0008761 Crustacea + 3.966	-0.147
		Exogenous	0.000	0.483	3.646	1.000	WG = -0.00006981 Exogenous + 3.646	-0.351
		Total	0.000	0.222	3.871	1.000	WG = -0.00004401 Total + 3.871	-0.198
Fish productivity by reach Prod	Abundance of invertebrates by reach At the end of the trial (33 days)	Diptera	0.000	0.184	12.880	1.000	Prod = -0.0003482 Diptera + 12.88	-0.077
		Trichoptera	0.004	0.693	11.830	1.000	Prod = 0.003705 Trichoptera + 11.83	-0.244
		Ephemeroptera	0.003	0.880	11.220	1.000	Prod = 0.002825 Ephemeroptera + 11.22	-0.054
		Mollusca	0.001	0.826	11.590	1.000	Prod = 0.0005948 Mollusca + 11.59	-0.138
		Coleoptera	0.000	0.579	12.110	1.000	Prod = 0.0001153 Coleoptera + 12.11	-0.264
		Plecoptera	0.021	0.619	11.810	1.000	Prod = 0.02081 Plecoptera + 11.81	-0.275
		Oligochaeta	-0.001	0.422	12.360	1.000	Prod = -0.001085 Oligochaeta + 12.36	-0.258
		Crustacea	0.001	0.618	11.870	1.000	Prod = 0.001065 Crustacea + 11.87	-0.268
		Exogenous	-0.001	0.457	12.350	1.000	Prod = -0.0005867 Exogenous + 12.35	-0.264
		Total	0.000	0.518	12.190	1.000	Prod = 0.00001373 Total + 12.19	-0.277

Gut contents

All the sixty-four alevins had prey in the stomach. The number of preys ingested was 1.5 lower in the control reaches when compared to the impacted ones (Table 4.5). The Chironomidae family was found in 100% of alevins from the control reaches and in 96.9% of alevins from the impacted ones. Chironomidae represented 68.3% and 76.1% of the prey consumed in the control and impacted reaches, respectively. Baetidae was the second most represented group (17.5% of prey in the control and 12.7% in the impacted reaches). The other groups represented less than 3% of the total amount of prey ingested.

Model estimations highlighted that fish consumed significantly less prey in the control (12.3 prey on average) than in the impacted reaches (17.4 prey on average, $P(\alpha > 0) = 0.998$ – Figure 4.4). Accordingly, fish ate significantly less Chironomidae in the control (8.4 in average) than in the impacted reaches (13.2 in average, $P(\alpha > 0) = 993$ – Figure 4.4). However, there was no difference in the number of Baetidae ingested (2.1 vs 2.2 respectively, in the control and in the impacted reaches – $P(\alpha > 0) = 0.559$).

Table 4.5 Diet composition. Number of prey (N), relative abundance (A) and frequency of occurrence in fish (F) in the control and in the impacted reaches.

	Control			Flood		
	N	A (%)	F (%)	N	A (%)	F (%)
Diptera						
<i>Chironomidae</i>	269	68.3	100	424	76.1	96.9
<i>Simuliidae</i>	8	2.0	21.9	5	0.9	15.6
<i>Empididae</i>	-	-	-	1	0.2	3.1
Brachycera	-	-	-	1	0.2	3.1
?	2	0.5	6.3	1	0.2	3.1
Trichoptera						
<i>Polycentropodidae</i>	7	1.8	18.8	9	1.6	21.9
<i>Philopotanidae</i>	-	-	-	1	0.2	3.1
<i>Hydropsychidae</i>	1	0.3	3.1	3	0.5	9.4
<i>Psychomyiidae</i>	-	-	-	1	0.2	3.1
?	5	1.3	15.6	5	0.9	12.5
Ephemeroptera						
<i>Baetidae</i>	69	17.5	87.5	71	12.7	71.9
<i>Heptageniidae</i>	3	0.8	9.4	4	0.7	6.3
<i>Ephemerellidae</i>	11	2.8	21.9	3	0.5	9.4
<i>Caenidae</i>	1	0.3	3.1	-	-	-
?	1	0.3	3.1	1	0.2	3.1
Coleoptera						
<i>Elmidae</i>	1	0.3	3.1	2	0.4	6.3
Amphipoda						
<i>Gammaridae</i>	2	0.5	6.3	3	0.5	6.3
Isopoda						
<i>Asellidae</i>	3	0.8	9.4	-	-	-
Oligochaeta						
	1	0.3	3.1	-	-	-
Others						
Hydracaria	4	1.0	3.1	6	1.1	6.3
Hemiptera	-	-	-	1	0.2	3.1
Nematoda	-	-	-	5	0.9	3.1
?	1	0.3	3.1	1	0.2	3.1
Exogenous	5	1.3	15.6	9	1.6	25.0
Total	394			557		

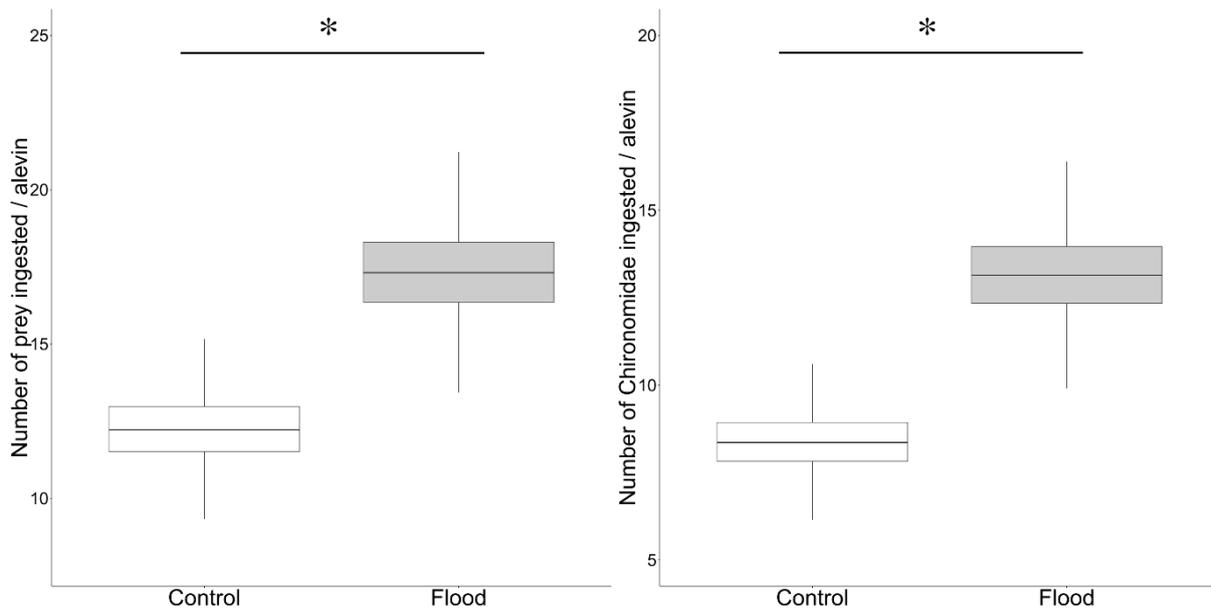


Figure 4.4 Model estimations of the number of prey ingested and the number of Chironomidae ingested by alevins in the control (in white) and in the impacted (in grey) reaches. Boxplots indicate the 1, 25, 50, 75 and 99 percentiles of posterior distributions. Significant differences are shown by *.

Table 4.6 Synthesis of the main results on the effects of the artificial flood on invertebrate abundances (total and Diptera), alevin performances (survival and growth) and alevin diet (number of prey eaten and number of Chironomids eaten) in late spring.

		Late Spring	
		Control	Flood
Invertebrates	Abundances ...		
	• Total	MODERATE	→ MODERATE
	• Diptera	HIGH	↗ VERY HIGH
	Recolonization	HIGH	↗ VERY HIGH
Alevin performances	Survival	HIGH 85 %	↘ MODERATE 60 %
	Growth	HIGH 3.4	↗ VERY HIGH 3.9
Alevin diet	Number of prey	12	↗ 17
	Number of Chironomids	8	↗ 12

Discussion

We failed at pointing out a significant effect of the artificial flood on the community of invertebrates. There was a high variability among the 3 samples characterizing a given reach, and this is probably why results were not significant. Thus, despite the homogeneity of the reaches in terms of water velocity, depth and gravel size, this suggests a huge heterogeneity in

spatial distribution of invertebrates as observed in the field (Downes et al. 1995; Heino et al. 2004). Nevertheless, the repeated trend for every taxa was a decrease in abundance in the impacted reaches and this trend persisted thirty-three days after the flood (except concerning Diptera). Because Diptera are essential in fish diet (see gut contents results), they deserve a specific attention. In control reaches, Diptera abundances double during the May month, probably in accompaniment to the increase in primary productivity and temperature. This happened also in the impacted reach, the average effect was much higher (more than 10 times increase), even if quite heterogeneous among reaches. All groups of invertebrates showed the same trend but Diptera was the only group exhibiting such a high increase in the impacted reaches.

As there was no significant impact of the artificial flood immediately afterwards, it was not expected to detect differences at the end of the trial, thirty-three days later. However, there were significantly less Ephemeroptera and Mollusca in the impacted reaches. Although there was no significant difference immediately after the disturbance, enough adults could have been displaced and/or the clutches hooked to the gravel (Gaino & Rebora 2001; Tachet et al. 2010) could have been decimated, weakening the next generation of these two orders.

Surviving alevins were more homogenous in weight in the impacted reaches than in the control ones. It is likely that the lack of food affected first the weaker ones, which would die because of their inability to establish a feeding territory and a lower amount of reserves than bigger alevins. This hypothesis is supported by Good et al. (2001) who concluded that early mortality associated with hydroclimatic events was high for small Atlantic salmon as they were not strong enough to survive. Then, massive mortalities caused by hydrological events reduce variability in individual growth rate (Vincenzi et al. 2012). The higher weight gain of alevins in impacted reaches than in the control ones could be related to density-dependent mechanisms with few survivors sharing the same amount of resources. Another explanation may rely on the difference in habitat availability. Indeed, floods mobilise fine particles filling the interstices of the gravel bed (Kondolf & Wilcock 1996; Mürle et al. 2003; Lepori & Hjerdt 2006), which maintain the heterogeneity of the substrate and provide shelters. Habitat heterogeneity increases the visual isolation between alevins and reduces the strength of the aggressive interactions (Kalleberg 1958; Bolliet et al. 2005). The combined effect of a diminution in the numbers of competitors and a heterogeneous habitat may have enhanced fish growth (Letcher & Terrick 1998). Gut content analyses highlighted a higher number of prey ingested by alevins in the impacted than in the control reaches and a large amount of these prey were Chironomidae. Cross et al. (2011)

found that a flood in the Colorado River (USA) decreased the number of benthic invertebrates while the production of rainbow trout (*Oncorhynchus mykiss*) increased. The trout production was mainly supported by invertebrates from the Simuliidae and the Chironomidae families, which both represented 50% of the prey ingested. They suggested that the flood created favourable conditions for age-0 trout: the scouring of the benthos associated with the flood enhanced the quality of the habitat and increased the production of Simuliidae and Chironomidae, which need a clean substrate for attachment. These taxa are also the pioneers at the recolonization time after hydrological disturbances (Otermin et al. 1998). Despite the absence of significant difference in the invertebrate community, it can be argued that the Chironomidae preferentially colonized the impacted reaches because the habitat was of better quality but the intrinsic variability of the experimental channel masked this phenomenon. Therefore, the increase in Diptera in the impacted reach may have also favour growth. However, the highest growth of alevins did not compensate for their lower survival and fish productivity was reduced by the impacted reach.

We predicted that reaches with a good food availability (high abundance of invertebrates) would enable the settlement of a stronger fish population (high numbers of survivors, high average weight gain and high productivity) but it was difficult to put forward this kind of relationship. Although abiotic factors were controlled and the habitat simplified compared to natural environment, the invertebrate community was unevenly spread and the heterogeneity in the invertebrate distribution limited the information provided by benthic samples. Additionally, Weber et al. (2017) demonstrated that measuring the biomass of invertebrates drifting was more accurate to estimate food supply for fish. Even if there is a link between benthic invertebrates and drifting invertebrates, direct relationship between benthic community composition and the productivity of salmonids is not well established (Faush et al. 1988). Moreover, sampling at a given time does not reflect the dynamics of the invertebrate population and the food availability on the 33 days that lasted the experiment. More accurate analysis taking into account only small larvae of Chironomidae, Simuliidae and Baetidae might improve our understanding of the mechanisms at work.

Conclusion

The impact of the artificial flood was low on the invertebrate assemblage, while it was obvious on brown trout young stages. At low fish density (2.6 fish m⁻²), the artificial flood decreased fish survival and improved fish growth (probably in relation with a mix between habitat, density and food availability). Despite our inability to identify the proximal factors underpinning fish

performances, emergence appears to be a critical stage, highly sensitive to hydrological disturbances since even at low density, and with relatively low impact on benthic invertebrates, fish productivity was negatively impacted.

General discussion

In the following section, I will highlight how results from different chapters complement each other to answer the original question: what could be the consequences of Global Climate Change (GCC, and notably an increase in temperature and flood frequency/intensity) on the trophic availability in invertebrate larvae for young trout stages. Then, the discussion will go through a more speculative part on the potential evolutionary consequences of GCC on salmonid phenology, before concluding on some perspectives.

Studying the ecological consequences of a flood by sampling in the wild is a tricky challenge. Indeed, because of the unpredictability of floods it is highly hazardous to gather enough points to assess the effects of flood on young emerging salmonids within a 3-year study. In addition, the instream habitat heterogeneity can mask or modify the effect of a disturbance. For all these reasons, we worked in a semi-natural environment. These experiments allow to evaluate the consequences of a flood in a context where invertebrate production and renewal is very close to the wild. The simulation of the flood with a motor pump created a flush that disturbed the gravel like a scouring flood. However, the impact of the simulated flush was restricted both in space and time. In addition, abiotic factors characterizing natural floods such as the gradual increase in water level and in turbidity were not reproduced. These cues announce the flood to the biota, which can shelter (Doeg & Milledge 1991; Suren & Jowett 2001). However, in the case of a scouring flood, the substratum is mobilised and most of the invertebrates are dislodged, including the sheltered ones. Water velocities obtained by the use of the motor pump were probably of the same order than velocities observed during natural scouring floods. Invertebrate species responded quite differently to the flood disturbance, but at least for Diptera and Ephemeroptera, densities were reduced.

In the chapters 3 and 4, we conducted experiments at two fish contrasted densities. In nursery streams, clutches are aggregated and it can result in high fish densities on the spawning grounds (Elliott 1989). We worked at 32 and 2.6 fish m⁻² and this last value corresponds to the carrying capacity in 0⁺ trout of the Lapitxuri semi-natural stream after emigration post-emergence (A. Bardonnnet, pers. com.). Analyses of alevin performances in late spring were quite disrupting between high (cages experiment) and low (reach experiment) density as the impacts of the flood diverged. At low density, survival was lower and growth higher in the impacted condition when compared to control, while the opposite was observed at high density. In addition, survival rates were quite similar according to density in the control conditions (0.84 at low density vs 0.81 at high density) but survival was much lower at low density in flooded conditions (0.63 at low density vs 0.94 at high density). Looking at weight gain, growth performances were 5 times

higher at low density in control, and even more in the impacted conditions. One possible explanation of such pattern may lie in the rhythm of the invertebrate recolonization process. Invertebrates sampling differed between low and high density protocol, preventing the comparison of abundances. However, we can compare differences in the pattern of abundances between the start and the end of the experiment. At high fish density, the flood diminished the abundances of Diptera immediately after the disturbance and one month later, Diptera did not recolonize and their abundance keep on declining, while the opposite was observed at low fish density. This was probably due to a high fish predation rate decreasing the Diptera abundances in the cages. In the reaches, fish density was too low to affect Diptera abundances and allowed the course of the recolonization process. We can then hypothesize that at the start of the experiment, the abundances of good sized preys, mainly chironomids, in the cage (high density) and in the reach (low density) were similarly low. However, prey were easy to catch in the cages because water velocity was lower, hunting surface was restricted and the prey shelters was limited to a small gravel layer. Therefore, alevins in the reaches were more likely to face a period of starvation leading to mortality than in the cages. Later, Diptera dramatically decreased in the cages, limiting fish growth. In the reach, the recolonization process was very efficient leading to the replenishment of Diptera and to excellent growth performances (confirmed by the number of chironomids in the gut content).

Of course, this scenario is speculative and many uncontrolled factors might have shape our results. Among them, temperature was on average one degree higher during the low density experiment and it may have interact. Water temperature averaged 10.2, 12.3°C at high density (chapter 3) and 13.3°C at low density (chapter 4). The last value is close to the upper thermal tolerance of alevins (Ojanguren & Braña 2003; Lahnsteiner 2012), and it may have increased the mortality rate of starving fish (as observed in chapter 2). Temperature may also interfere with growth. However in chapter 2 we observed no difference in growth between 8 and 11°C for fish fed *ad libitum* (despite the fact that they almost double the food intake at warm temperature). In addition, after a delayed access to food, alevins were more capable to resume a normal diet and growth was higher at low temperature (12% differences in growth catch-up between the two temperatures). This was quite surprising and could be due to the short duration of the experiment, but it could also be the consequence of a growth optimum closer to 8 than to 11°C in trout early ontogenesis (Elliott & Elliott 2010).

Fish growth and survival are linked in a complex and hardly predictable manner and the intensity of the density-dependent mechanisms partially enlighten the results of the Lapitxuri

experiments. At high population abundance, space is limited and density-dependent mortality and emigration occur. At low population abundance, mortality rate is mainly density independent while density-dependence only operates on growth during the earliest time period (Jenkins et al. 1999; Grant & Imre 2005; Imre et al. 2005; Lobón-Cerviá 2007). It was only during a second time period that density-dependence operates on both growth and mortality. This way, the population can persist after catastrophic disturbances and recover quickly, which reduces the extinction risk of stream dwelling salmonid populations in variable environments. Vincenzi et al. (2008) pointed out that the population abundance of marble trout (*Salmo marmoratus*) was diminished by severe floods, while growth of surviving juveniles was stimulated. This phenomenon was linked to both high food supply and low competition due to a low population density following the event. Moreover, females were larger to sexual maturity and produced more eggs. The increase in fertility allowed the population to recover fast and to be highly resilient.

Throughout spring, temperature, sunshine and brightness duration increase and initiate both primary and secondary production. Consequently, stream productivity is higher in late spring than in early spring. Emergence is spread during spring and according to literature (e.g. Einum & Fleming 2000), the timing of salmonid emergence is under high selective pressure. The timing of emergence is the result of a trade-off between food and territory availabilities (Bromage et al. 2001; Letcher et al. 2004 – Figure 1). Alevins emerging in early spring undergo harsh environmental conditions with a reduced amount of food. However, the low density of alevins allows them to establish feeding territories on the best hunting spots, giving them a growth advantage. In contrast, alevins emerging in late spring face good environmental conditions with an abundant amount of food but the number of territories available in optimal habitat is limited and intensifies the strength of the competition between congeners. The prior residency of the early emergent alevins gives them a significant advantage (Johnsson et al. 1999; Harwood et al. 2003). These mechanisms maintain a variability in the timing of emergence but this could be disrupted by Global Climate Change.

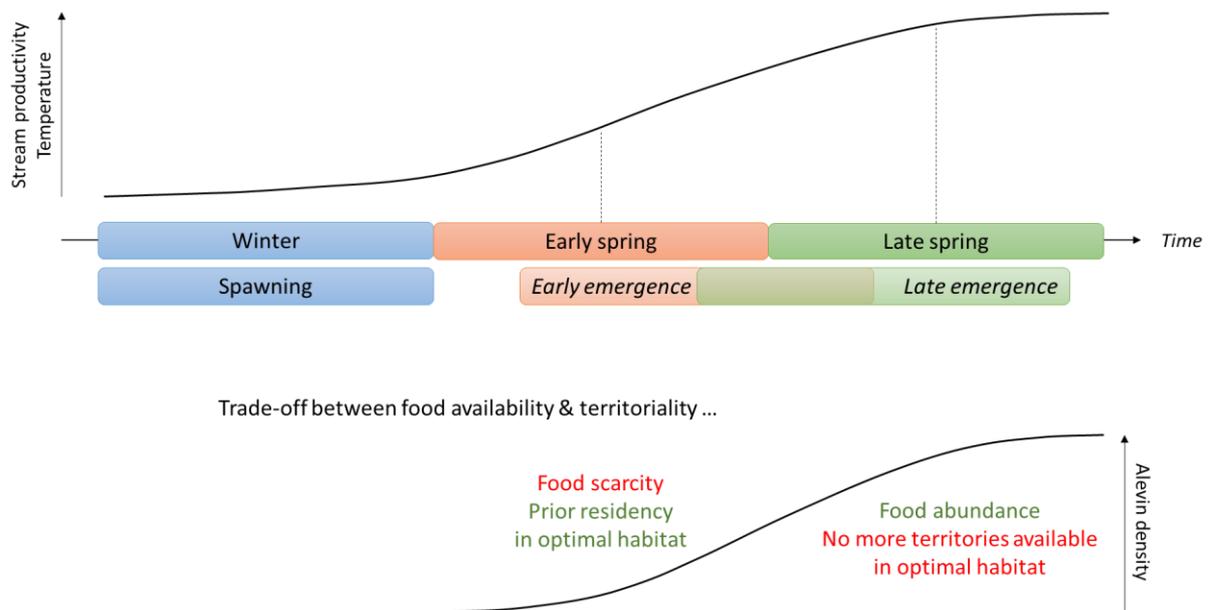


Figure 1 Stream productivity increases during the spring. Emergence is spread from early spring to late spring and alevin density is a trade-off between food availability and territoriality.

Currently, prey availability matches with the timing of emergence. GCC should increase the intensity and the frequency of stochastic hydrologic events such as catastrophic floods. This could directly diminish the production, the abundances of stream invertebrates by displacing and killing them and consequently, reduced the prey availability for carnivorous fish. Additionally, GCC should increase the temperature and affect the phenology of brown trout, including the timing of emergence. As brown trout is an ectotherm, warmer temperature will reduce the development duration and advance the timing of emergence. Consequently, GCC should delayed the peak of prey availability while the emergence date should be advanced (Figure 2).

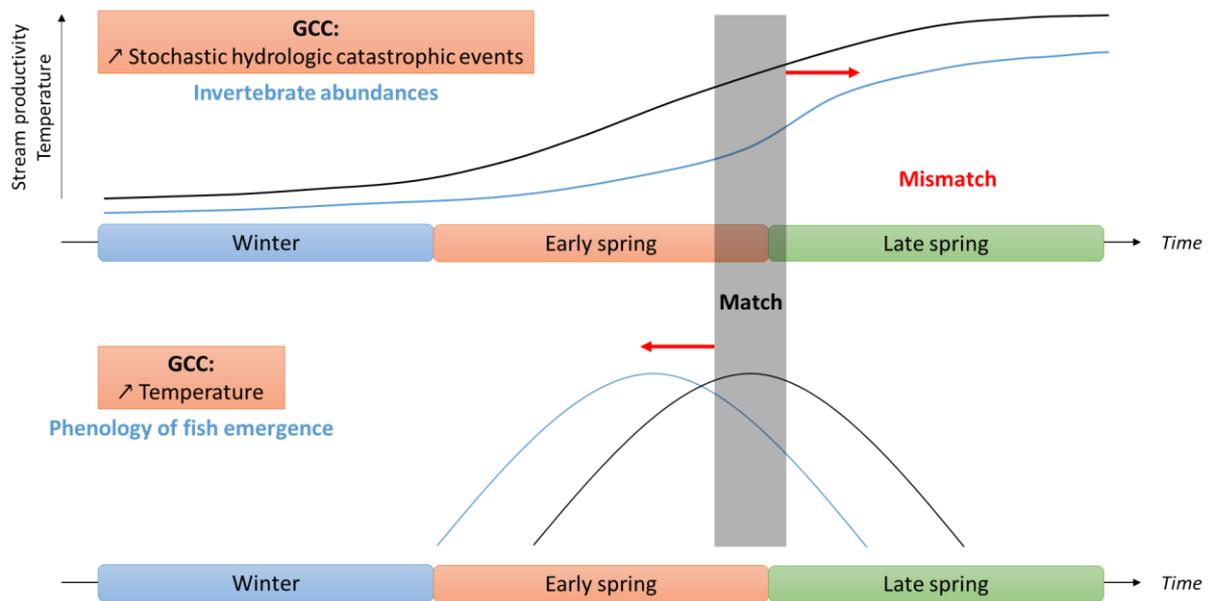


Figure 2 Currently, fish emergence timing matches with invertebrate abundance (in black) but Global Climate Change should delay the increase in invertebrate abundance and advance the emergence date (in blue) and create a mismatch between prey availability and predator requirements (in red).

Nevertheless, temperature affects the whole ecosystem metabolism. According to Woodward et al. (2010), warmer temperature should put forward the initiation of both primary and secondary productivity. Therefore, a move forward in the emergence date might be supported by an earlier primary production (Demars et al. 2011) and a higher leaf decomposition rate (Pereira et al. 2017), leading to high invertebrate production. Invertebrate drift should also be enhanced: “mechanically” with the increase in invertebrate abundance, and also in relation to drifting behaviour. Results from chapter 1 suggested that *Baetis* drift may depend on temperature. This experiment was not designated to look at the effect of temperature, but differences in temperature (12.2 vs 14.7°C) accompanied the spring and autumn trials. In the autumn trial, the warmest, *Baetis* drift rate was increased by nearly a third (26%). The increase in temperature could then mitigate the negative effect of flood on invertebrates.

Temperature should also affect the proximal mechanisms affecting the timing of emergence (Figure 3). We know that an increase in temperature will at first reduce the time needed for the embryo-larval development as an immediate physiological response to temperature. This will lead to an earlier emergence date. The quicker development of the embryo-larval stages can be compensated by a delay in the spawning season, which depends strongly on the photoperiod and falling autumn temperature (Beacham & Murray 1990; Van Der Kraak and Pankhurst 1997; Pankhurst & Porter 2003; Pankhurst & King 2010; Pankhurst & Munday 2011). Literature

suggests that genitors adjust the spawning date to match the emergence with favourable conditions of water flow, temperature and food supply (Crozier et al. 2008). Hence, Unwin et al. (2000) demonstrated that later spawning occurs in chinook salmon populations where embryos develop in warmer water. Similarly, Webb & McLay (1996) highlighted that spawning time of Atlantic salmon varied along an altitudinal gradient in Scotland rivers. At high altitudes, temperatures were lower and salmons spawned earlier. The same trend was observed by Warren et al. (2012), warmer temperatures delayed spawning of brook trout (*Salvelinus fontinalis*) to coincide the date of emergence with the peak of prey production. These two phenomenon (*i.e.* a quicker development associated with a delayed spawning season) could compensate each other but it is still difficult to say whether the emergence date will be changed. The third mechanism affecting the timing of emergence is the variation in egg size. Johnston & Legget (2002) demonstrated that warmer temperature increased the egg size but this affects the survival of eggs. Large eggs are stenotherm (Régnier et al. 2013). Moreover, maximal egg size is related to the female size (Hendry & Day 2003) and maternal fitness is a trade-off between egg size and egg number (Einum et al. 2002). Then, warmer temperature should increase the mortality rate of large eggs and the egg size increase should decrease the egg number. The pre-emergence mortality should be higher, resulting in fewer emerging fish. However, large eggs develop faster and emerge early (Einum et al. 2002; Rollinson & Hutchings 2010). Early emerged alevins have higher metabolic rates (Régnier et al. 2012a) and the energy conversion efficiency into growth is higher for large eggs than for small eggs (Régnier et al 2012b), which produces larger alevins. These four factors should support post-emergence survival and confer a competitive advantage to alevins hatching from large eggs but it is difficult to evaluate if the strategy to produce large eggs should be favoured by GCC.

Effects of Global Climate Change on Emergence: Proximal mechanisms

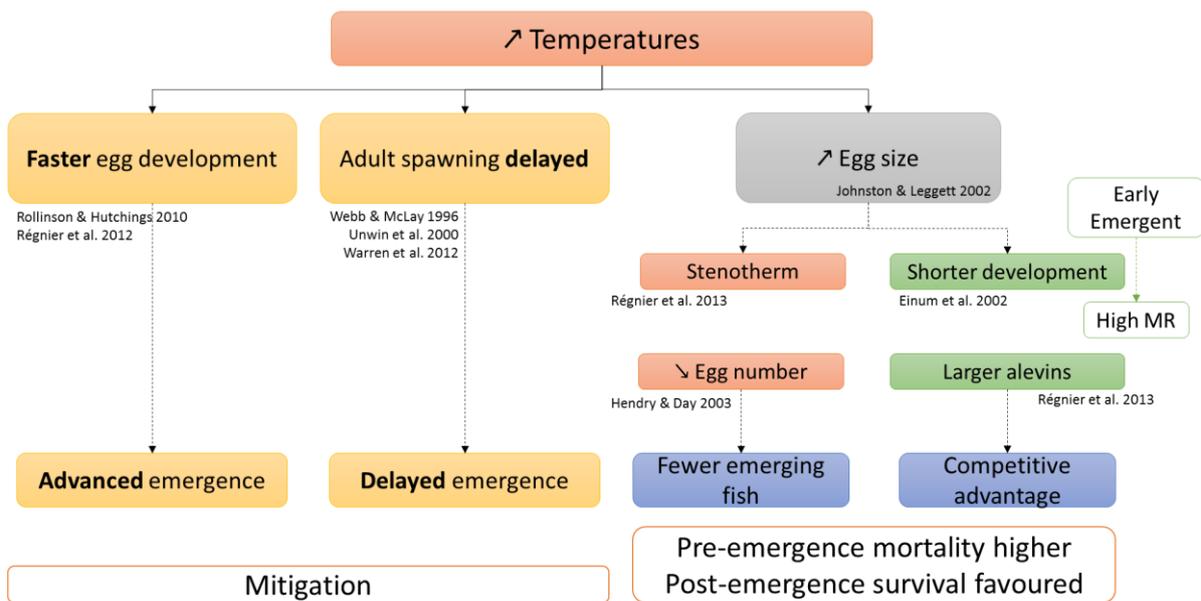


Figure 3 Effects of Global Climate Change on the proximal mechanisms affecting the emergence date (i.e. the egg development duration, the timing of the spawning season and the size of the eggs).

GCC should also be a selective force by selecting individuals according to their temperaments or personalities, which induce individual differences in exploration and activity (Le Galliard et al. 2013). These behaviours are associated with foraging, territorial defence, mate finding, and dispersal as well as habitat choice. For example, individuals with high activity increase both growth and the risk of mortality from predation, thus resulting in a similar fitness than low activity individuals (Stamps 2007). Réale et al. (2010) associated behaviour (i.e. activity, superficial exploration, boldness and aggressiveness) with physiology (i.e. immunity, metabolism and oxidative stress). This so-called “pace-of-life” syndrome emphasizes integration of behavioural variation within a slow-fast demographic and metabolic continuum. If we try to evaluate the impacts of GCC in this theoretical context of “pace-of-life” syndrome, alevins should be selected according to their probability to avoid starvation or their ability to withstand it during period of food scarcity (Figure 4). With a high prey production in late spring, alevins emerging later diminish their risk of starvation. Likewise, alevins with low metabolic rate and a high amount of reserves have a high ability to survive during starvation. Late emerging alevins have low metabolic rates, then it is possible that alevins exhibit these two traits simultaneously as they are probably correlated but these alevins usually have a low amount of reserves. According to the “pace-of-life” syndrome (Ricklefs & Wikelski 2002; Réale et al. 2010), shy alevins which are associated with low metabolic rate could be favoured.

This selection could have consequences on adult spawning strategies, on maternal investment and on the whole population functioning and dynamics. Despite this very speculative scenario, it is still difficult to predict which direction will take the selection with Global Climate Change and, for now, very few elements are available.

Effects of Global Climate Change on Emergence: **Ultimate mechanisms**

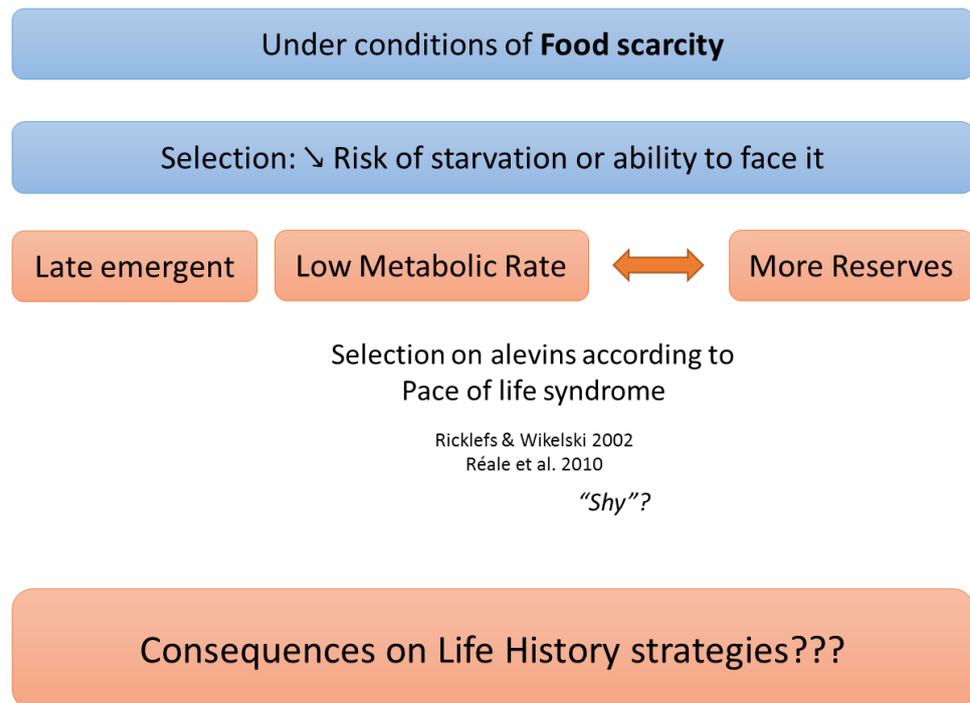


Figure 4 According to the environmental conditions (food scarcity), alevins could be selected depending on their ability to avoid or to face periods of starvation. This could have consequences on life history strategies and on the whole population functioning.

We did not succeed in increasing sufficiently the water velocities in our experimental facilities to approach flood velocities. However, the link between moderate water velocities and the invertebrate propensity to drift has been clarified for the three species we focused on. Even if results depended in part on the experimental conditions, we found three distinct patterns and we provided a precise quantification of the drift probability of the taxa studied. Laboratory experiment also allowed us to assess the ability of alevins to face starvation, the consequences of such stress on their metabolism and their ability to recover according to temperature. These two experiments could be reproduced with temperatures ranging from 4 to 14°C, which corresponds to the limits of the thermal tolerance of brown trout alevins (Elliott 1994) and temperature that alevins can face throughout their distributional range (Jonsson & Jonsson 2009). Consequently, according to thermal conditions, the drift propensity of invertebrates

would allow to evaluate the trophic availability for alevins (and more widely for drift-feeding fish) and the metabolic, survival and growth consequences of alevins facing periods of starvation would be nearly known. These data (those already acquired and those from future potential experiments) should be used as basis for calibrating and developing a global individual-based model (Grimm 1999) to simulate scenarios of GCC and to better understand the consequences on brown trout alevin survival and growth (Clark et al. 2001). Incrementing results from studies on the phenology and the productivity of systems could provide complementary elements for modelling climate change related scenarios and the consequences of rising temperature and occurrence of flood on a brown trout population.

It is forecasted that Global Climate Change should increase the intensity of hydrological events as well as their frequency. Connell (1978) proposed the Intermediate Disturbance Hypothesis (IDH), which states that in absence of disturbance, species richness is expected to be low with more competitive species dominating and outcompeting the others. Similarly, when disturbances are too frequent, few species are able to resist to the changes in their environment or to colonize during the brief periods between disturbances. Under a disturbance regime that is intermediate in frequency and intensity, both rapid colonizers and more competitive species co-occur and species richness is maximised (Townsend et al. 1997). The increase in flood frequency associated with GCC should diminish species diversity of invertebrates and this could potentially affect the quality of prey for brown trout. Chironomids are pioneer species, have short life cycle and quick turn-over rates. As they constitute the main part of brown trout diet, frequent floods could increase the production of Chironomids and make them more available for brown trout.

Lastly, in Southern and Mediterranean Europe, GCC should induce more droughts in spring. Droughts reduce the volume of water available for fish, impeding or preventing their migration and adversely affecting water quality, especially water temperature and dissolved oxygen (Elliott 2000). The aggregation of organisms (invertebrates and fish) could modify the intra-/inter-specific interactions, as well as the amount of prey and their availability for predators. Then, it should be interesting to study the consequences of drought on the invertebrate assemblage and on just-emerged alevins.

To conclude, this work highlighted the possible consequences of GCC on the invertebrate fauna and on young trout stages of brown trout. In a local management context, it is difficult to mitigate the effects of GCC on riverine ecosystems. The building of a flood control dam would limit the intensity of floods but would not reduce their occurrence. It would be necessary to

study the consequences of these facilities on the hydrological river regime and on the biota in order to evaluate their effectiveness. However, it appears necessary to limit other sources of disturbance to the biota (*i.e.* pollution or habitat fragmentation) in order to limit the stress induced by the interaction between GCC and others potential factors.

References

- Aas, Ø., Haider, W., & Hunt, L. (2000). Angler responses to potential harvest regulations in a Norwegian sport fishery: a conjoint-based choice modeling approach. *North American Journal of Fisheries Management*, **20**(4), 940-950.
- Allan, J. D., & Castillo, M. M. (2007). *Stream ecology: structure and function of running waters*. Springer Science & Business Media.
- Allen, K. R. (1951). *The Horokiwi Stream: a study of a trout population* (No. 10). New Zealand Marine Department.
- Alvarez, D., & Nicieza, A. G. (2003). Predator avoidance behaviour in wild and hatchery-reared brown trout: the role of experience and domestication. *Journal of Fish Biology*, **63**(6), 1565-1577.
- Argerich, A., Puig García, M., & Pupilli, E. (2004). Effect of floods of different magnitude on the macroinvertebrate communities of Matarranya stream (Ebro river basin, NE Spain). *Limnetica*, **23**(3-4), 283-294.
- Armstrong, J. D., & Nislow, K. H. (2006). Critical habitat during the transition from maternal provisioning in freshwater fish, with emphasis on Atlantic salmon (*Salmo salar*) and brown trout (*Salmo trutta*). *Journal of Zoology*, **269**(4), 403-413.
- Armstrong, J. D., Kemp, P. S., Kennedy, G. J. A., Ladle, M., & Milner, N. J. (2003). Habitat requirements of Atlantic salmon and brown trout in rivers and streams. *Fisheries research*, **62**(2), 143-170.
- Bacon, P. J., Gurney, W. S. C., Jones, W., McLaren, I. S., & Youngson, A. F. (2005). Seasonal growth patterns of wild juvenile fish: partitioning variation among explanatory variables, based on individual growth trajectories of Atlantic salmon (*Salmo salar*) parr. *Journal of Animal Ecology*, **74**(1), 1-11.
- Baglinière, J. L., & Maisse, G. (1991). *La truite. Biologie et écologie*. Editions Quae.
- Bal, G., Rivot, E., Baglinière, J. L., White, J., & Prévost, E. (2014). A hierarchical Bayesian model to quantify uncertainty of stream water temperature forecasts. *PloS one*, **9**(12), e115659.
- Bar, N. (2014). Physiological and hormonal changes during prolonged starvation in fish. *Canadian Journal of Fisheries and Aquatic Sciences*, **71**(10), 1447-1458.

- Bardonnnet, A., & Heland, M. (1994). The influence of potential predators on the habitat preferenda of emerging brown trout. *Journal of Fish Biology*, **45**(sA), 131-142.
- Beacham, T. D., & Murray, C. B. (1990). Temperature, egg size, and development of embryos and alevins of five species of Pacific salmon: a comparative analysis. *Transactions of the American Fisheries Society*, **119**(6), 927-945.
- Benke, A. C., Hall, C. A., Hawkins, C. P., Lowe-McConnell, R. H., Stanford, J. A., Suberkropp, K., & Ward, J. V. (1988). Bioenergetic considerations in the analysis of stream ecosystems. *Journal of the North American Benthological Society*, **7**(4), 480-502.
- Bewick, S., Cantrell, R. S., Cosner, C., & Fagan, W. F. (2016). How resource phenology affects consumer population dynamics. *The American Naturalist*, **187**(2), 151-166.
- Blöschl, G., Hall, J., Parajka, J., Perdigão, R. A., Merz, B., Arheimer, B., ... & Čanjevac, I. (2017). Changing climate shifts timing of European floods. *Science*, **357**(6351), 588-590.
- Bolliet, V., Bardonnnet, A., Jarry, M., Vignes, J. C., & Gaudin, P. (2005). Does embeddedness affect growth performance in juvenile salmonids? An experimental study in brown trout, *Salmo trutta* L. *Ecology of Freshwater Fish*, **14**(3), 289-295.
- Bozek, M. A., DeBrey, L. D., & Lockwood, J. A. (1994). Diet overlap among size classes of Colorado River cutthroat trout (*Oncorhynchus clarki pleuriticus*) in a high-elevation mountain stream. *Hydrobiologia*, **273**(1), 9-17.
- Bratrich, C., Truffer, B., Jorde, K., Markard, J., Meier, W., Peter, A., ... & Wehrli, B. (2004). Green hydropower: a new assessment procedure for river management. *River Research and Applications*, **20**(7), 865-882.
- Bretschko, G., & Moog, O. (1990). Downstream effects of intermittent power generation. *Water Science and Technology*, **22**(5), 127-135.
- Brittain, J. E., & Eikeland, T. J. (1988). Invertebrate drift—a review. *Hydrobiologia*, **166**(1), 77-93.
- Bromage, N., Porter, M., & Randall, C. (2001). The environmental regulation of maturation in farmed finfish with special reference to the role of photoperiod and melatonin. *Aquaculture*, **197**(1), 63-98.

- Brooker, M. P., & Hemsworth, R. J. (1978). The effect of the release of an artificial discharge of water on invertebrate drift in the R. Wye, Wales. *Hydrobiologia*, **59**(3), 155-163.
- Bruno, M. C., Siviglia, A., Carolli, M., & Maiolini, B. (2013). Multiple drift responses of benthic invertebrates to interacting hydropeaking and thermopeaking waves. *Ecohydrology*, **6**(4), 511-522.
- Butler, J. R., Radford, A., Riddington, G., & Laughton, R. (2009). Evaluating an ecosystem service provided by Atlantic salmon, sea trout and other fish species in the River Spey, Scotland: the economic impact of recreational rod fisheries. *Fisheries Research*, **96**(2), 259-266.
- Cada, G. F., Loar, J. M., & Sale, M. J. (1987). Evidence of food limitation of rainbow and brown trout in southern Appalachian soft-water streams. *Transactions of the American Fisheries Society*, **116**(5), 692-702.
- Cattaneo, F., Hugueny, B., & Lamouroux, N. (2003). Synchrony in brown trout, *Salmo trutta*, population dynamics: a 'Moran effect' on early-life stages. *Oikos*, **100**(1), 43-54.
- Cattaneo, F., Lamouroux, N., Breil, P., & Capra, H. (2002). The influence of hydrological and biotic processes on brown trout (*Salmo trutta*) population dynamics. *Canadian journal of fisheries and aquatic sciences*, **59**(1), 12-22.
- Céréghino, R., & Lavandier, P. (1998). Influence of hypolimnetic hydropeaking on the distribution and population dynamics of Ephemeroptera in a mountain stream. *Freshwater Biology*, **40**(2), 385-399.
- Céréghino, R., Cugny, P., & Lavandier, P. (2002). Influence of intermittent hydropeaking on the longitudinal zonation patterns of benthic invertebrates in a mountain stream. *International review of hydrobiology*, **87**(1), 47-60.
- Clark, M. E., Rose, K. A., Levine, D. A., & Hargrove, W. W. (2001). Predicting climate change effects on appalachian trout: combining gis and individual-based modeling. *Ecological Applications*, **11**(1), 161-178.
- Cobb, D. G., Galloway, T. D., & Flannagan, J. F. (1992). Effects of discharge and substrate stability on density and species composition of stream insects. *Canadian Journal of Fisheries and Aquatic Sciences*, **49**(9), 1788-1795.

- Connell, J. H. (1978). Diversity in tropical rain forests and coral reefs. *Science*, **199**(4335), 1302-1310.
- Crespin De Billy, V., & Usseglio-Polatera, P. (2002). Traits of brown trout prey in relation to habitat characteristics and benthic invertebrate communities. *Journal of Fish Biology*, **60**(3), 687-714.
- Cross, W. F., Baxter, C. V., Donner, K. C., Rosi-Marshall, E. J., Kennedy, T. A., Hall, R. O., ... & Rogers, R. S. (2011). Ecosystem ecology meets adaptive management: food web response to a controlled flood on the Colorado River, Glen Canyon. *Ecological Applications*, **21**(6), 2016-2033.
- Crozier, L. G., Hendry, A. P., Lawson, P. W., Quinn, T. P., Mantua, N. J., Battin, J., ... & Huey, R. B. (2008). Potential responses to climate change in organisms with complex life histories: evolution and plasticity in Pacific salmon. *Evolutionary Applications*, **1**(2), 252-270.
- Cushing, D. H. (1969). The regularity of the spawning season of some fishes. *ICES Journal of Marine Science*, **33**(1), 81-92.
- Cushing, D. H. (1990). Plankton production and year-class strength in fish populations: an update of the match/mismatch hypothesis. *Advances in marine biology*, **26**, 249-293.
- Dabrowski, K. (1984). The feeding of fish larvae: present «state of the art» and perspectives. *Reproduction Nutrition Développement*, **24**(6), 807-833.
- Dahl, J., & Greenberg, L. (1996). Impact on stream benthic prey by benthic vs drift feeding predators: a meta-analysis. *Oikos*, 177-181.
- Dankers, R., & Feyen, L. (2008). Climate change impact on flood hazard in Europe: An assessment based on high-resolution climate simulations. *Journal of Geophysical Research: Atmospheres*, **113**(D19).
- Demars, B. O., Russell Manson, J., Olafsson, J. S., Gislason, G. M., Gudmundsdóttir, R., Woodward, G. U. Y., ... & Friberg, N. (2011). Temperature and the metabolic balance of streams. *Freshwater Biology*, **56**(6), 1106-1121.
- Doeg, T. J., & Milledge, G. A. (1991). Effect of experimentally increasing concentration of suspended sediment on macroinvertebrate drift. *Marine and Freshwater Research*, **42**(5), 519-526.

- Domagała, J., Krepski, T., Czerniawski, R., & Pilecka-Rapacz, M. (2015). Prey availability and selective feeding of sea trout (*Salmo trutta* L., 1758) fry stocked in small forest streams. *Journal of Applied Ichthyology*, **31**(2), 375-380.
- Dou, S. Z., Masuda, R., Tanaka, M., & Tsukamoto, K. (2005). Effects of temperature and delayed initial feeding on the survival and growth of Japanese flounder larvae. *Journal of Fish Biology*, **66**(2), 362-377.
- Downes, B. J., Lake, P. S., & Schreiber, E. S. G. (1995). Habitat structure and invertebrate assemblages on stream stones: a multivariate view from the riffle. *Austral Ecology*, **20**(4), 502-514.
- Durant, J. M., Anker-Nilssen, T., & Stenseth, N. C. (2003). Trophic interactions under climate fluctuations: the Atlantic puffin as an example. *Proceedings of the Royal Society of London B: Biological Sciences*, **270**(1523), 1461-1466.
- Durant, J. M., Hjermmann, D. Ø., Anker-Nilssen, T., Beaugrand, G., Mysterud, A., Pettoelli, N., & Stenseth, N. C. (2005). Timing and abundance as key mechanisms affecting trophic interactions in variable environments. *Ecology Letters*, **8**(9), 952-958.
- Durant, J. M., Hjermmann, D. Ø., Ottersen, G., & Stenseth, N. C. (2007). Climate and the match or mismatch between predator requirements and resource availability. *Climate research*, **33**(3), 271-283.
- Ehrlich, K. F. (1974). Chemical changes during growth and starvation of larval *Pleuronectes platessa*. *Marine Biology*, **24**(1), 39-48.
- Einum, S., & Fleming, I. A. (2000). Selection against late emergence and small offspring in Atlantic salmon (*Salmo salar*). *Evolution*, **54**(2), 628-639.
- Einum, S., Hendry, A. P., & Fleming, I. A. (2002). Egg-size evolution in aquatic environments: does oxygen availability constrain size? *Proceedings of the Royal Society of London B: Biological Sciences*, **269**(1507), 2325-2330.
- Elliott, J. M. (1967). The food of trout (*Salmo trutta*) in a Dartmoor stream. *Journal of Applied Ecology*, 59-71.
- Elliott, J. M. (1971). The distances travelled by drifting invertebrates in a Lake District stream. *Oecologia*, **6**(4), 350-379.

- Elliott, J. M. (1986). Spatial distribution and behavioural movements of migratory trout *Salmo trutta* in a Lake District stream. *The Journal of Animal Ecology*, 907-922.
- Elliott, J. M. (1989). The critical-period concept for juvenile survival and its relevance for population regulation in young sea trout, *Salmo trutta*. *Journal of Fish Biology*, **35**(sA), 91-98.
- Elliott, J. M. (1994). *Quantitative ecology and the brown trout*. Oxford University Press.
- Elliott, J. M. (2000). Pools as refugia for brown trout during two summer droughts: trout responses to thermal and oxygen stress. *Journal of fish biology*, **56**(4), 938-948.
- Elliott, J. M., & Elliott, J. A. (2010). Temperature requirements of Atlantic salmon *Salmo salar*, brown trout *Salmo trutta* and Arctic charr *Salvelinus alpinus*: predicting the effects of climate change. *Journal of fish biology*, **77**(8), 1793-1817.
- Elliott, J. M., Hurley, M. A., & Elliott, J. A. (1997). Variable effects of droughts on the density of a sea-trout *Salmo trutta* population over 30 years. *Journal of Applied Ecology*, 1229-1238.
- Elwood, J. W., & Waters, T. F. (1969). Effects of floods on food consumption and production rates of a stream brook trout population. *Transactions of the American Fisheries Society*, **98**(2), 253-262.
- Fahy, E. (1980). Prey selection by young trout fry (*Salmo trutta*). *Journal of Zoology*, **190**(1), 27-37.
- Fausch, K. D. (1984). Profitable stream positions for salmonids: relating specific growth rate to net energy gain. *Canadian journal of zoology*, **62**(3), 441-451.
- Fausch, K. D., Hawkes, C. L., & Parsons, M. G. (1988). Models that predict standing crop of stream fish from habitat variables: 1950-85.
- Fuller, R. L., Griego, C., Muehlbauer, J. D., Dennison, J., & Doyle, M. W. (2010). Response of stream macroinvertebrates in flow refugia and high-scour areas to a series of floods: a reciprocal replacement study. *Journal of the North American Benthological Society*, **29**(2), 750-760.
- Gaino, E., & Reborá, M. (2001). Synthesis and function of the fibrous layers covering the eggs of *Siphonurus lacustris* (Ephemeroptera, Siphonuridae). *Acta Zoologica*, **82**(1), 41-48.
- Galliard, J. F., Paquet, M., Cisel, M., & Montes-Poloni, L. (2013). Personality and the pace-of-life syndrome: variation and selection on exploration, metabolism and locomotor performances. *Functional Ecology*, **27**(1), 136-144.

- Gelman, A., & Hill, J. (2007). *Data analysis using regression and multilevel hierarchical models* (Vol. 1). New York, NY, USA: Cambridge University Press.
- Giller, P. S., & Malmqvist, B. (1998). *The biology of streams and rivers*. Oxford University Press.
- Giroux, F., Ovidio, M., Phillippart, J. C., & Baras, E. (2000). Relationship between the drift of macroinvertebrates and the activity of brown trout in a small stream. *Journal of Fish Biology*, **56**(5), 1248-1257.
- Good, S. P., Dodson, J. J., Meekan, M. G., & Ryan, D. A. (2001). Annual variation in size-selective mortality of Atlantic salmon (*Salmo salar*) fry. *Canadian Journal of Fisheries and Aquatic Sciences*, **58**(6), 1187-1195.
- Gore, J. A., Layzer, J. B., & Mead, J. I. M. (2001). Macroinvertebrate instream flow studies after 20 years: a role in stream management and restoration. *River Research and Applications*, **17**(4-5), 527-542.
- Grant, J. W. A., & Imre, I. (2005). Patterns of density-dependent growth in juvenile stream-dwelling salmonids. *Journal of Fish Biology*, **67**(sB), 100-110.
- Grant, J. W., & Kramer, D. L. (1990). Territory size as a predictor of the upper limit to population density of juvenile salmonids in streams. *Canadian Journal of Fisheries and Aquatic Sciences*, **47**(9), 1724-1737.
- Grant, J. W., Girard, I. L., Breau, C., & Weir, L. K. (2002). Influence of food abundance on competitive aggression in juvenile convict cichlids. *Animal Behaviour*, **63**(2), 323-330.
- Grant, J. W., Steingrímsson, S. Ó., Keeley, E. R., & Cunjak, R. A. (1998). Implications of territory size for the measurement and prediction of salmonid abundance in streams. *Canadian Journal of Fisheries and Aquatic Sciences*, **55**(S1), 181-190.
- Grimm, V. (1999). Ten years of individual-based modelling in ecology: what have we learned and what could we learn in the future? *Ecological modelling*, **115**(2), 129-148.
- Harwood, A. J., Metcalfe, N. B., Griffiths, S. W., & Armstrong, J. D. (2002). Intra- and inter-specific competition for winter concealment habitat in juvenile salmonids. *Canadian Journal of Fisheries and Aquatic Sciences*, **59**(9), 1515-1523.

- Harwood, A. J., Griffiths, S. W., Metcalfe, N. B., & Armstrong, J. D. (2003). The relative influence of prior residency and dominance on the early feeding behaviour of juvenile Atlantic salmon. *Animal Behaviour*, **65**(6), 1141-1149.
- Hayes, J. W., Hughes, N. F., & Kelly, L. H. (2007). Process-based modelling of invertebrate drift transport, net energy intake and reach carrying capacity for drift-feeding salmonids. *Ecological Modelling*, **207**(2), 171-188.
- Hayes, J. W., Stark, J. D., & Shearer, K. A. (2000). Development and test of a whole-lifetime foraging and bioenergetics growth model for drift-feeding brown trout. *Transactions of the American Fisheries Society*, **129**(2), 315-332.
- Hecketsweiler, B., & Hecketsweiler, P. (2004). Voyage en biochimie: circuits en biochimie humaine, nutritionnelle et métabolique.
- Heggenes, J. (1988). Effects of short-term flow fluctuations on displacement of, and habitat use by, brown trout in a small stream. *Transactions of the American Fisheries Society*, **117**(4), 336-344.
- Heggenes, J., & Traaen, T. (1988). Downstream migration and critical water velocities in stream channels for fry of four salmonid species. *Journal of Fish Biology*, **32**(5), 717-727.
- Heggenes, J., Bagliniere, J. L., & Cunjak, R. A. (1999). Spatial niche variability for young Atlantic salmon (*Salmo salar*) and brown trout (*S. trutta*) in heterogeneous streams. *Ecology of Freshwater Fish*, **8**(1), 1-21.
- Heino, J., Louhi, P., & Muotka, T. (2004). Identifying the scales of variability in stream macroinvertebrate abundance, functional composition and assemblage structure. *Freshwater Biology*, **49**(9), 1230-1239.
- Héland, M. (1999). Social organization and territoriality in brown trout juveniles during ontogeny. *Biology and Ecology of the Brown and Sea trout*, **151**, 115.
- Hendry, A. P., & Day, T. (2003). Revisiting the positive correlation between female size and egg size. *Evolutionary Ecology Research*, **5**(3), 421-429.
- Hjort, J. (1914). Fluctuations in the great fisheries of northern Europe viewed in the light of biological research. ICES.

- Horta, F., Santos, H., Tavares, L., Antunes, M., Pinheiro, P., & Callisto, M. (2009). Assessment of benthic macroinvertebrate habitat suitability in a tropical watershed. In *Proceedings of the 7th International Symposium on Ecohydraulics* (pp. 170-179).
- Horton, R. E. (1945). Erosional development of streams and their drainage basins; hydrophysical approach to quantitative morphology. *Geological society of America bulletin*, **56**(3), 275-370.
- Hubert, W. A., & Rhodes, H. A. (1992). Sizes of prey consumed by age-0 brown trout in Douglas Creek, Wyoming. *Journal of Freshwater Ecology*, **7**(3), 277-282.
- Huet, M. (1949). Aperçu des relations entre la pente et les populations piscicoles des eaux courantes. *Aquatic Sciences-Research Across Boundaries*, **11**(3), 332-351.
- Huet, M. (1954). Biologie, profils en long et en travers des eaux courantes. *Bulletin français de Pisciculture*, **175**, 41-53.
- Hunter, J. R. (1981). Feeding Ecology and Predation of Marine Fish Larvae. *R. Lasker ed*, **1**, 34-77.
- Huryn, A. D. (1996). An appraisal of the Allen paradox in a New Zealand trout stream. *Limnology and Oceanography*, **41**(2), 243-252.
- Imre, I., Grant, J. W. A., & Cunjak, R. A. (2005). Density-dependent growth of young-of-the-year Atlantic salmon *Salmo salar* in Catamaran Brook, New Brunswick. *Journal of Animal Ecology*, **74**(3), 508-516.
- IPCC. (2014). Part A: Global and Sectoral Aspects. (Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change). Climate Change 2014: Impacts, Adaptation, and Vulnerability., 1132. Retrieved from https://www.ipcc.ch/pdf/assessment-report/ar5/wg2/WGIIAR5-FrontMatterA_FINAL.pdf
- Jenkins, T. M. (1969). Social structure, position choice and micro-distribution of two trout species (*Salmo trutta* and *Salmo gairdneri*) resident in mountain streams. *Animal behaviour monographs*, **2**, 55-123.
- Jenkins, T. M., Diehl, S., Kratz, K. W., & Cooper, S. D. (1999). Effects of population density on individual growth of brown trout in streams. *Ecology*, **80**(3), 941-956.

- Jensen, A. J., & Johnsen, B. O. (1999). The functional relationship between peak spring floods and survival and growth of juvenile Atlantic salmon (*Salmo salar*) and brown trout (*Salmo trutta*). *Functional Ecology*, **13**(6), 778-785.
- Johnson, J. H., McKenna, J. E., & Douglass, K. A. (2013). Movement and feeding ecology of recently emerged steelhead in Lake Ontario tributaries. *Journal of Applied Ichthyology*, **29**(1), 221-225.
- Johnsson, J. I., Nöbbelein, F., & Bohlin, T. (1999). Territorial competition among wild brown trout fry: effects of ownership and body size. *Journal of Fish Biology*, **54**(2), 469-472.
- Jonas, J. L., & Wahl, D. H. (1998). Relative importance of direct and indirect effects of starvation for young walleyes. *Transactions of the American Fisheries Society*, **127**(2), 192-205.
- Jonsson, B., & Jonsson, N. (2009). A review of the likely effects of climate change on anadromous Atlantic salmon *Salmo salar* and brown trout *Salmo trutta*, with particular reference to water temperature and flow. *Journal of fish biology*, **75**(10), 2381-2447.
- Jonsson, B., & Jonsson, N. (2014). Early environment influences later performance in fishes. *Journal of Fish Biology*, **85**(2), 151-188.
- Jowett, I. G., & Richardson, J. (1990). Microhabitat preferences of benthic invertebrates in a New Zealand river and the development of in-stream flow-habitat models for *Deleatidium* spp. *New Zealand journal of marine and freshwater research*, **24**(1), 19-30.
- Kalleberg, H. (1958). Observations in a stream tank of territoriality and competition in juvenile salmon and trout. *Inst. Freshw. Res. Drottingholm Rep.*, **39**, 55-98.
- Kamalam, B. S., Medale, F., & Panserat, S. (2017). Utilisation of dietary carbohydrates in farmed fishes: New insights on influencing factors, biological limitations and future strategies. *Aquaculture*, **467**, 3-27.
- Keeley, E. R., & Grant, J. W. (2001). Prey size of salmonid fishes in streams, lakes, and oceans. *Canadian Journal of Fisheries and Aquatic Sciences*, **58**(6), 1122-1132.
- Kennedy, B. P., Nislow, K. H., & Folt, C. L. (2008). Habitat-mediated foraging limitations drive survival bottlenecks for juvenile salmon. *Ecology*, **89**(9), 2529-2541.

- Klemetsen, A., Amundsen, P. A., Dempson, J. B., Jonsson, B., Jonsson, N., O'Connell, M. F., & Mortensen, E. (2003). Atlantic salmon *Salmo salar* L., brown trout *Salmo trutta* L. and Arctic charr *Salvelinus alpinus* (L.): a review of aspects of their life histories. *Ecology of freshwater fish*, **12**(1), 1-59.
- Kondolf, G. M., & Wilcock, P. R. (1996). The flushing flow problem: defining and evaluating objectives. *Water Resources Research*, **32**(8), 2589-2599.
- Korman, J., Kaplinski, M., & Melis, T. S. (2011). Effects of fluctuating flows and a controlled flood on incubation success and early survival rates and growth of age-0 rainbow trout in a large regulated river. *Transactions of the American Fisheries Society*, **140**(2), 487-505.
- Ladago, B. J., Marsden, J. E., & Evans, A. N. (2016). Early feeding by lake trout fry. *Transactions of the American Fisheries Society*, **145**(1), 1-6.
- Lagarrigue, T., Céréghino, R., Lim, P., Reyes-Marchant, P., Chappaz, R., Lavandier, P., & Belaud, A. (2002). Diel and seasonal variations in brown trout (*Salmo trutta*) feeding patterns and relationship with invertebrate drift under natural and hydropeaking conditions in a mountain stream. *Aquatic living resources*, **15**(2), 129-137.
- Lahnsteiner, F. (2012). Thermotolerance of brown trout, *Salmo trutta*, gametes and embryos to increased water temperatures. *Journal of Applied Ichthyology*, **28**(5), 745-751.
- Laliberte, J. J., Post, J. R., Rosenfeld, J. S., & Mee, J. A. (2016). Modelling Temperature, Body Size, Prey Density, and Stream Gradient Impacts on Longitudinal Patterns of Potential Production of Drift-Feeding Trout. *River Research and Applications*, **32**(10), 2045-2055.
- Latterell, J. J., Fausch, K. D., Gowan, C., & Riley, S. C. (1998). Relationship of trout recruitment to snowmelt runoff flows and adult trout abundance in six Colorado mountain streams. *Rivers*, **6**(4), 240-250.
- Lehner, B., Döll, P., Alcamo, J., Henrichs, T., & Kaspar, F. (2006). Estimating the impact of global change on flood and drought risks in Europe: a continental, integrated analysis. *Climatic Change*, **75**(3), 273-299.
- Leigh, C., Bush, A., Harrison, E. T., Ho, S. S., Luke, L., Rolls, R. J., & Ledger, M. E. (2015). Ecological effects of extreme climatic events on riverine ecosystems: insights from Australia. *Freshwater Biology*, **60**(12), 2620-2638.

- Lepori, F., & Hjerdt, N. (2006). Disturbance and aquatic biodiversity: reconciling contrasting views. *AIBS Bulletin*, **56**(10), 809-818.
- Letcher, B. H., & Terrick, T. D. (1998). Maturation of male age-0 Atlantic salmon following a massive, localized flood. *Journal of Fish Biology*, **53**(6), 1243-1252.
- Letcher, B. H., Dubreuil, T., O'donnell, M. J., Obedzinski, M., Griswold, K., & Nislow, K. H. (2004). Long-term consequences of variation in timing and manner of fry introduction on juvenile Atlantic salmon (*Salmo salar*) growth, survival, and life-history expression. *Canadian Journal of Fisheries and Aquatic Sciences*, **61**(12), 2288-2301.
- Leung, E. S., Rosenfeld, J. S., & Bernhardt, J. R. (2009). Habitat effects on invertebrate drift in a small trout stream: implications for prey availability to drift-feeding fish. *Hydrobiologia*, **623**(1), 113-125.
- Li, F., Cai, Q., Fu, X., & Liu, J. (2009). Construction of habitat suitability models (HSMs) for benthic macroinvertebrate and their applications to instream environmental flows: a case study in Xiangxi River of Three Gorges Reservoir region, China. *Progress in Natural Science*, **19**(3), 359-367.
- Li, F., Chung, N., Bae, M. J., Kwon, Y. S., & Park, Y. S. (2012). Relationships between stream macroinvertebrates and environmental variables at multiple spatial scales. *Freshwater Biology*, **57**(10), 2107-2124.
- Lobón-Cerviá, J. (2004). Discharge-dependent covariation patterns in the population dynamics of brown trout (*Salmo trutta*) within a Cantabrian river drainage. *Canadian Journal of Fisheries and Aquatic Sciences*, **61**(10), 1929-1939.
- Lobón-Cerviá, J. (2005). Spatial and temporal variation in the influence of density dependence on growth of stream-living brown trout (*Salmo trutta*). *Canadian Journal of Fisheries and Aquatic Sciences*, **62**(6), 1231-1242.
- Lobón-Cerviá, J. (2007). Density-dependent growth in stream-living Brown Trout *Salmo trutta* L. *Functional Ecology*, **21**(1), 117-124.
- Lobón-Cerviá, J., & Mortensen, E. (2005). Population size in stream-living juveniles of lake-migratory brown trout *Salmo trutta* L.: the importance of stream discharge and temperature. *Ecology of Freshwater fish*, **14**(4), 394-401.

- Mackay, R. J. (1992). Colonization by lotic macroinvertebrates: a review of processes and patterns. *Canadian Journal of Fisheries and Aquatic Sciences*, **49**(3), 617-628.
- Mäki-Petäys, A., Muotka, T., & Huusko, A. (1999). Densities of juvenile brown trout (*Salmo trutta*) in two subarctic rivers: assessing the predictive capability of habitat preference indices. *Canadian Journal of Fisheries and Aquatic Sciences*, **56**(8), 1420-1427.
- Malmqvist, B. (2002). Aquatic invertebrates in riverine landscapes. *Freshwater Biology*, **47**(4), 679-694.
- McCormack, J. C. (1962). The food of young trout (*Salmo trutta*) in two different becks. *The Journal of Animal Ecology*, 305-316.
- McCue, M. D. (2010). Starvation physiology: reviewing the different strategies animals use to survive a common challenge. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, **156**(1), 1-18.
- McMullen, L. E., & Lytle, D. A. (2012). Quantifying invertebrate resistance to floods: a global-scale meta-analysis. *Ecological Applications*, **22**(8), 2164-2175.
- Melo, A. S., Niyogi, D. K., Matthaei, C. D., & Townsend, C. R. (2003). Resistance, resilience, and patchiness of invertebrate assemblages in native tussock and pasture streams in New Zealand after a hydrological disturbance. *Canadian Journal of Fisheries and Aquatic Sciences*, **60**(6), 731-739.
- Mennigen, J. A., Skiba-Cassy, S., & Panserat, S. (2013). Ontogenetic expression of metabolic genes and microRNAs in rainbow trout alevins during the transition from the endogenous to the exogenous feeding period. *Journal of Experimental Biology*, **216**(9), 1597-1608.
- Mesa, L. M. (2010). Effect of spates and land use on macroinvertebrate community in Neotropical Andean streams. *Hydrobiologia*, **641**(1), 85-95.
- Miller, S. W., & Judson, S. (2014). Responses of macroinvertebrate drift, benthic assemblages, and trout foraging to hydropeaking. *Canadian Journal of Fisheries and Aquatic Sciences*, **71**(5), 675-687.
- Milner, N. J., Elliott, J. M., Armstrong, J. D., Gardiner, R., Welton, J. S., & Ladle, M. (2003). The natural control of salmon and trout populations in streams. *Fisheries Research*, **62**(2), 111-125.

- Moog, O. (1993). Quantification of daily peak hydropower effects on aquatic fauna and management to minimize environmental impacts. *River Research and Applications*, *8*(1-2), 5-14.
- Mürle, U., Ortlepp, J., & Zahner, M. (2003). Effects of experimental flooding on riverine morphology, structure and riparian vegetation: The River Spöl, Swiss National Park. *Aquatic Sciences-Research Across Boundaries*, *65*(3), 191-198.
- Nakano, S., Kawaguchi, Y., Taniguchi, Y., Miyasaka, H., Shibata, Y., Urabe, H., & Kuhara, N. (1999). Selective foraging on terrestrial invertebrates by rainbow trout in a forested headwater stream in northern Japan. *Ecological Research*, *14*(4), 351-360.
- Naman, S. M., Rosenfeld, J. S., & Richardson, J. S. (2016). Causes and consequences of invertebrate drift in running waters: from individuals to populations and trophic fluxes. *Canadian Journal of Fisheries and Aquatic Sciences*, *73*(8), 1292-1305.
- Negishi, J. N., & Richardson, J. S. (2006). An experimental test of the effects of food resources and hydraulic refuge on patch colonization by stream macroinvertebrates during spates. *Journal of Animal Ecology*, *75*(1), 118-129.
- Nehring, R. B., & Anderson, R. M. (1993). Determination of population-limiting critical salmonid habitats in Colorado streams using the Physical Habitat Simulation System. *Rivers*, *4*(1), 1-19.
- NRC, 2011. Carbohydrates and Fibre, in: Nutrient Requirements of Fish and Shrimp. The National Academies Press, Washington DC, pp. 135–162
- Nuhfer, A. J., Clark, R. D., & Alexander, G. R. (1994). *Recruitment of brown trout in the south branch of the Au Sable River, Michigan in relation to stream flow and winter severity*. Lansing: Michigan Department of Natural Resources.
- O'Hop, J., & Wallace, J. B. (1983). Invertebrate drift, discharge, and sediment relations in a southern Appalachian headwater stream. *Hydrobiologia*, *98*(1), 71-84.
- Ojanguren, A. F., & Brana, F. (2003). Thermal dependence of embryonic growth and development in brown trout. *Journal of Fish Biology*, *62*(3), 580-590.
- Orpwood, J. E., Griffiths, S. W., & Armstrong, J. D. (2003). Effects of body size on sympatric shelter use in over-wintering juvenile salmonids. *Journal of Fish Biology*, *63*(s1), 166-173.

- Otermin, A., Basaguren, A., & Pozo, J. (2002). Re-colonization by the macroinvertebrate community after a drought period in a first-order stream (Agüera Basin, Northern Spain). *Limnetica*, **21**(1-2), 117-128.
- Pankhurst, N. W., & King, H. R. (2010). Temperature and salmonid reproduction: implications for aquaculture. *Journal of Fish Biology*, **76**(1), 69-85.
- Pankhurst, N. W., & Munday, P. L. (2011). Effects of climate change on fish reproduction and early life history stages. *Marine and Freshwater Research*, **62**(9), 1015-1026.
- Pankhurst, N. W., & Porter, M. J. R. (2003). Cold and dark or warm and light: variations on the theme of environmental control of reproduction. *Fish Physiology and Biochemistry*, **28**(1), 385-389.
- Pankhurst, N. W., & Van Der Kraak, G. (1997). *Effects of stress on reproduction and growth of fish* (pp. 73-93). Cambridge, UK: Cambridge University Press.
- Pereira, A., Trabulo, J., Fernandes, I., Pascoal, C., Cássio, F., & Duarte, S. (2017). Spring stimulates leaf decomposition in moderately eutrophic streams. *Aquatic Sciences*, **79**(1), 197-207.
- Perkins, D. M., Reiss, J., Yvon-Durocher, G., & Woodward, G. (2010). Global change and food webs in running waters. *Hydrobiologia*, **657**(1), 181-198.
- Poff, N. L., Olden, J. D., Vieira, N. K., Finn, D. S., Simmons, M. P., & Kondratieff, B. C. (2006). Functional trait niches of North American lotic insects: traits-based ecological applications in light of phylogenetic relationships. *Journal of the North American Benthological Society*, **25**(4), 730-755.
- Pyke, G. H., Pulliam, H. R., & Charnov, E. L. (1977). Optimal foraging: a selective review of theory and tests. *The quarterly review of biology*, **52**(2), 137-154.
- Quinn, J. M., & Hickey, C. W. (1990). Characterisation and classification of benthic invertebrate communities in 88 New Zealand rivers in relation to environmental factors. *New Zealand journal of marine and freshwater research*, **24**(3), 387-409.
- Quinn, T. P., & Peterson, N. P. (1996). The influence of habitat complexity and fish size on over-winter survival and growth of individually marked juvenile coho salmon (*Oncorhynchus kisutch*) in Big Beef Creek, Washington. *Canadian Journal of Fisheries and Aquatic Sciences*, **53**(7), 1555-1564.

- Rader, R. B. (1997). A functional classification of the drift: traits that influence invertebrate availability to salmonids. *Canadian Journal of Fisheries and Aquatic Sciences*, **54**(6), 1211-1234.
- Rawi, C. S., Al-Shami, S. A., Madrus, M. R., & Ahmad, A. H. (2014). Biological and ecological diversity of aquatic macroinvertebrates in response to hydrological and physicochemical parameters in tropical forest streams of Gunung Tebu, Malaysia: implications for ecohydrological assessment. *Ecohydrology*, **7**(2), 496-507.
- Réale, D., Garant, D., Humphries, M. M., Bergeron, P., Careau, V., & Montiglio, P. O. (2010). Personality and the emergence of the pace-of-life syndrome concept at the population level. *Philosophical Transactions of the Royal Society of London B: Biological Sciences*, **365**(1560), 4051-4063.
- Régnier, T., Bolliet, V., Gaudin, P., & Labonne, J. (2012b). Female effects on offspring energetic status and consequences on early development in yolk feeding Brown trout (*Salmo trutta*). *Journal of Experimental Zoology Part A: Ecological Genetics and Physiology*, **317**(6), 347-358.
- Régnier, T., Bolliet, V., Gaudin, P., & Labonne, J. (2013). Bigger is not always better: egg size influences survival throughout incubation in brown trout (*Salmo trutta*). *Ecology of Freshwater Fish*, **22**(2), 169-177.
- Régnier, T., Labonne, J., Gaudin, P., & Bolliet, V. (2012a). Influence of energetic status on ontogenetic niche shifts: emergence from the redd is linked to metabolic rate in brown trout. *Oecologia*, **168**(2), 371-380.
- Ricklefs, R. E., & Wikelski, M. (2002). The physiology/life-history nexus. *Trends in Ecology & Evolution*, **17**(10), 462-468.
- Ringler, N. H. (1985). Individual and temporal variation in prey switching by brown trout, *Salmo trutta*. *Copeia*, 918-926.
- Robinson, C. T., Uehlinger, U. R. S., & Monaghan, M. T. (2004). Stream ecosystem response to multiple experimental floods from a reservoir. *River Research and Applications*, **20**(4), 359-377.

- Rollinson, N., & Hutchings, J. A. (2010). Why does egg size increase with maternal size? Effects of egg size and egg density on offspring phenotypes in Atlantic salmon (*Salmo salar*). *Evolutionary Ecology Research*, **12**(8), 949-960.
- Romaniszyn, E. D., Hutchens, J. J., & Bruce Wallace, J. (2007). Aquatic and terrestrial invertebrate drift in southern Appalachian Mountain streams: implications for trout food resources. *Freshwater Biology*, **52**(1), 1-11.
- Roussel, J. M., & Bardonnnet, A. (1997). Diel and seasonal patterns of habitat use by fish in a natural salmonid brook: an approach to the functional role of the riffle-pool sequence. *Bulletin Français de la Pêche et de la Pisciculture* **346**, 573-588.
- Roussel, J. M., & Bardonnnet, A. (2002). The habitat of juvenile brown trout (*Salmo trutta* L.) in small streams: Preferences, movements, diel and seasonal variations. *Bulletin Français de la Pêche et de la Pisciculture (France)*.
- Ryer, C. H., & Olla, B. L. (1996). Growth depensation and aggression in laboratory reared coho salmon: the effect of food distribution and ration size. *Journal of Fish Biology*, **48**(4), 686-694.
- Sánchez-Hernández, J., Servia, M. J., Vieira-Lanero, R., & Cobo, F. (2012). Ontogenetic dietary shifts in a predatory freshwater fish species: the brown trout as an example of a dynamic fish species. In *New advances and contributions to fish biology*. InTech.
- Sánchez-Hernández, J., Vieira-Lanero, R., Servia, M. J., & Cobo, F. (2011). First feeding diet of young brown trout fry in a temperate area: disentangling constraints and food selection. *Hydrobiologia*, **663**(1), 109-119.
- Sandri, M. (2010). Autophagy in health and disease. 3. Involvement of autophagy in muscle atrophy. *American Journal of Physiology-Cell Physiology*, **298**(6), C1291-C1297.
- Schülting, L., Feld, C. K., & Graf, W. (2016). Effects of hydro- and thermopeaking on benthic macroinvertebrate drift. *Science of The Total Environment*, **573**, 1472-1480.
- Seegrist, D. W., & Gard, R. (1972). Effects of floods on trout in Sagehen Creek, California. *Transactions of the American Fisheries Society*, **101**(3), 478-482.
- Shearer, K. A., Hayes, J. W., Jowett, I. G., & Olsen, D. A. (2015). Habitat suitability curves for benthic macroinvertebrates from a small New Zealand river. *New Zealand journal of marine and freshwater research*, **49**(2), 178-191.

- Simpkins, D. G., Hubert, W. A., Del Rio, C. M., & Rule, D. C. (2003). Physiological responses of juvenile rainbow trout to fasting and swimming activity: effects on body composition and condition indices. *Transactions of the American Fisheries Society*, **132**(3), 576-589.
- Skoglund, H., & Barlaup, B. T. (2006). Feeding pattern and diet of first feeding brown trout fry under natural conditions. *Journal of Fish Biology*, **68**(2), 507-521.
- Skoglund, H., Einum, S., & Robertsen, G. (2011). Competitive interactions shape offspring performance in relation to seasonal timing of emergence in Atlantic salmon. *Journal of Animal Ecology*, **80**(2), 365-374.
- Sogard, S. M. (1997). Size-selective mortality in the juvenile stage of teleost fishes: a review. *Bulletin of marine science*, **60**(3), 1129-1157.
- St Pierre, J. I., & Kovalenko, K. E. (2014). Effect of habitat complexity attributes on species richness. *Ecosphere*, **5**(2), 1-10.
- Stewart, P. M., Butcher, J. T., & Swinford, T. O. (2000). Land use, habitat, and water quality effects on macroinvertebrate communities in three watersheds of a Lake Michigan associated marsh system. *Aquatic Ecosystem Health & Management*, **3**(1), 179-189.
- Strahler, A. N. (1954). Statistical analysis in geomorphic research. *The Journal of Geology*, **62**(1), 1-25.
- Strahler, A. N. (1957). Quantitative analysis of watershed geomorphology. *Eos, Transactions American Geophysical Union*, **38**(6), 913-920.
- Strange, E. M., Moyle, P. B., & Foin, T. C. (1993). Interactions between stochastic and deterministic processes in stream fish community assembly. *Environmental Biology of Fishes*, **36**(1), 1-15.
- Sundby, A., Eliassen, K. A., Blom, A. K., & Asgard, T. (1991). Plasma insulin, glucagon, glucagon-like peptide and glucose levels in response to feeding, starvation and life long restricted feed ration in salmonids. *Fish physiology and biochemistry*, **9**(3), 253-259.
- Suren, A. M., & Jowett, I. G. (2001). Effects of deposited sediment on invertebrate drift: an experimental study. *New Zealand Journal of Marine and Freshwater Research*, **35**(4), 725-737.
- Tachet, H., Richoux, P., Bournaud, M., & Usseglio-Polatera, P. (2010). *Invertébrés d'eau douce: systématique, biologie, écologie*(Vol. 15). Paris: CNRS editions.

- Theodoropoulos, C., Vourka, A., Stamou, A., Rutschmann, P., & Skoulikidis, N. (2017). Response of freshwater macroinvertebrates to rainfall-induced high flows: A hydroecological approach. *Ecological Indicators*, **73**, 432-442.
- Thompson, J. N., & Beauchamp, D. A. (2016). Growth of juvenile steelhead *Oncorhynchus mykiss* under size-selective pressure limited by seasonal bioenergetic and environmental constraints. *Journal of fish biology*, **89**(3), 1720-1739.
- Townsend, C. R., Doledec, S., Norris, R., Peacock, K., & Arbuckle, C. (2003). The influence of scale and geography on relationships between stream community composition and landscape variables: description and prediction. *Freshwater Biology*, **48**(5), 768-785.
- Townsend, C. R., Scarsbrook, M. R., & Dolédec, S. (1997). The intermediate disturbance hypothesis, refugia, and biodiversity in streams. *Limnology and oceanography*, **42**(5), 938-949.
- Unwin, M. J., Quinn, T. P., Kinnison, M. T., & Boustead, N. C. (2000). Divergence in juvenile growth and life history in two recently colonized and partially isolated chinook salmon populations. *Journal of Fish Biology*, **57**(4), 943-960.
- van Vliet, M. T., Franssen, W. H., Yearsley, J. R., Ludwig, F., Haddeland, I., Lettenmaier, D. P., & Kabat, P. (2013). Global river discharge and water temperature under climate change. *Global Environmental Change*, **23**(2), 450-464.
- Vannote, R. L., Minshall, G. W., Cummins, K. W., Sedell, J. R., & Cushing, C. E. (1980). The river continuum concept. *Canadian journal of fisheries and aquatic sciences*, **37**(1), 130-137.
- Vignes, J., & Heland, M. (1995). Comportement alimentaire au cours du changement d'habitat lié à l'émergence chez le saumon atlantique *Salmo salar* L. et la truite commune *Salmo trutta* L., en conditions semi-naturelles. *Bulletin Français de la Pêche et de la Pisciculture*, (337-338-339), 207-214.
- Vincenzi, S., Crivelli, A. J., Giske, J., Satterthwaite, W. H., & Mangel, M. (2012). Selective consequences of catastrophes for growth rates in a stream-dwelling salmonid. *Oecologia*, **168**(2), 393-404.
- Vincenzi, S., Crivelli, A. J., Jesensek, D., & De Leo, G. A. (2008). The role of density-dependent individual growth in the persistence of freshwater salmonid populations. *Oecologia*, **156**(3), 523-534.

- Vøllestad, L. A., Olsen, E. M., & Forseth, T. (2002). Growth-rate variation in brown trout in small neighbouring streams: evidence for density-dependence? *Journal of Fish Biology*, **61**(6), 1513-1527.
- Wallace, J. B., & Webster, J. R. (1996). The role of macroinvertebrates in stream ecosystem function. *Annual review of entomology*, **41**(1), 115-139.
- Wankowski, J. W. J. (1979). Morphological limitations, prey size selectivity, and growth response of juvenile Atlantic salmon, *Salmo salar*. *Journal of Fish Biology*, **14**(1), 89-100.
- Ward, D. M., Nislow, K. H., & Folt, C. L. (2009). Increased population density and suppressed prey biomass: relative impacts on juvenile Atlantic salmon growth. *Transactions of the American Fisheries Society*, **138**(1), 135-143.
- Ware, D. M. (1972). Predation by rainbow trout (*Salmo gairdneri*): the influence of hunger, prey density, and prey size. *Journal of the Fisheries Board of Canada*, **29**(8), 1193-1201.
- Ware, D. M. (1972). Predation by rainbow trout (*Salmo gairdneri*): the influence of hunger, prey density, and prey size. *Journal of the Fisheries Board of Canada*, **29**(8), 1193-1201.
- Warren, D. R., Robinson, J. M., Josephson, D. C., Sheldon, D. R., & Kraft, C. E. (2012). Elevated summer temperatures delay spawning and reduce redd construction for resident brook trout (*Salvelinus fontinalis*). *Global Change Biology*, **18**(6), 1804-1811.
- Waters, T. F. (1988). Fish production-benthos production relationships in trout streams. *Polskie Archiwum Hydrobiologii/Polish Archives of Hydrobiology*, **35**(3), 545-561.
- Webb, J. H., & McLay, H. A. (1996). Variation in the time of spawning of Atlantic salmon (*Salmo salar*) and its relationship to temperature in the Aberdeenshire Dee, Scotland. *Canadian Journal of Fisheries and Aquatic Sciences*, **53**(12), 2739-2744.
- Weber, N., Bouwes, N., & Jordan, C. (2017). Accounting for spatial and temporal variation in macroinvertebrate community abundances when measuring the food supply of stream salmonids. *Freshwater Science*, **36**(2), 364-376.
- Wenger, S. J., Isaak, D. J., Dunham, J. B., Fausch, K. D., Luce, C. H., Neville, H. M., ... & Chandler, G. L. (2011). Role of climate and invasive species in structuring trout distributions in the interior Columbia River Basin, USA. *Canadian Journal of Fisheries and Aquatic Sciences*, **68**(6), 988-1008.

- Werner, E. E., & Gilliam, J. F. (1984). The ontogenetic niche and species interactions in size-structured populations. *Annual review of ecology and systematics*, **15**(1), 393-425.
- Wildhaber, M. L., & Lamberson, P. J. (2004). Importance of the habitat choice behavior assumed when modeling the effects of food and temperature on fish populations. *Ecological Modelling*, **175**(4), 395-409.
- Winterbourn, M. J. (1981). The use of aquatic invertebrates in studies of stream water quality. *Water and soil publication*, **22**, 5-16.
- Woodward, G. U. Y. (2009). Biodiversity, ecosystem functioning and food webs in fresh waters: assembling the jigsaw puzzle. *Freshwater Biology*, **54**(10), 2171-2187.
- Woodward, G., Perkins, D. M., & Brown, L. E. (2010). Climate change and freshwater ecosystems: impacts across multiple levels of organization. *Philosophical Transactions of the Royal Society of London B: Biological Sciences*, **365**(1549), 2093-2106.
- Wysujack, K., Greenberg, L. A., Bergman, E., & Olsson, I. C. (2009). The role of the environment in partial migration: food availability affects the adoption of a migratory tactic in brown trout *Salmo trutta*. *Ecology of Freshwater Fish*, **18**(1), 52-59.
- Yokota, T., Nakagawa, T., Murakami, N., Chimura, M., Tanaka, H., Yamashita, Y., & Funamoto, T. (2016). Effects of starvation at the first feeding stage on the survival and growth of walleye pollock *Gadus chalcogrammus* larvae. *Fisheries science*, **82**(1), 73-83.
- Young, K. A. (2003). Evolution of fighting behavior under asymmetric competition: an experimental test with juvenile salmonids. *Behavioral Ecology*, **14**(1), 127-134.

Article references of Chapter 1

- Abràmoff, M. D., P. J. Magalhães, & S. J. Ram, 2004. Image processing with imageJ. *Biophotonics International* **11**: 36–41.
- Allan, J. D., 1984. The size composition of invertebrate drift in a rocky mountain stream. *Oikos* **43**: 68–76.
- Berg, M. B., 1995. *Larval food and feeding behaviour The Chironomidae: Biology and Ecology of Non-biting Midges*. Springer Netherlands, Dordrecht: 136–168.
- Bowles, D. E., & R. A. Short, 1988. Size composition of invertebrate drift and fish predation in a Texas stream. *Southwestern Naturalist* **33**: 177–184.

- Brittain, J., & T. Eikeland, 1988. Invertebrate drift - a review. *Hydrobiologia* **166**: 77–93.
- Bruno, M. C., M. J. Cashman, B. Maiolini, S. Biffi, & G. Zolezzi, 2016. Responses of benthic invertebrates to repeated hydropeaking in semi-natural flume simulations. *Ecohydrology* **9**: 68–82.
- Carling, P. A., 1987. *Bed stability in gravel streams, with reference to stream regulation and ecology* In Richards, K. (ed), *River Channels: Environment and Process*. Oxford: 321–347.
- Ciborowski, J. J. H., 1983. Influence of current velocity, density, and detritus on drift of two mayfly species (Ephemeroptera). *Canadian Journal of Zoology* **61**: 119–125.
- Charpentier, B., & Morin, A., 1994. Effect of current velocity on ingestion rates of black fly larvae. *Canadian Journal of Fisheries and Aquatic Sciences* **51**, 1615–1619.
- Fenoglio, S., F. Boano, T. Bo, R. Revelli, & L. Ridolfi, 2013. The impacts of increasing current velocity on the drift of *Simulium monticola* (Diptera: Simuliidae): a laboratory approach. *Italian Journal of Zoology* **80**: 443–448.
- Finelli, C. M., D. D. Hart, & R. A. Merz, 2002. Stream insects as passive suspension feeders: Effects of velocity and food concentration on feeding performance. *Oecologia* **131**: 145–153.
- Fingerut, J. T., D. M. Fonseca, J. R. Thomson, & D. D. Hart, 2015. Seeking shelter from the storm: responses of benthic stream invertebrates to natural and experimental floods. *Freshwater Science* **34**: 897–908.
- Fjellheim, A., 1996. Distribution of benthic invertebrates in relation to stream flow characteristics in a Norwegian river. *Regulated Rivers-Research & Management* **12**: 263–271.
- Fonseca, D. M., & D. D. Hart, 1996. Density-dependent dispersal of black fly neonates is mediated by flow. *Oikos* **75**: 49–58.
- Gelman, A., & J. Hill, 2006. *Data analysis using regression and multilevel/hierarchical models*. Policy Analysis. Cambridge University Press.
- Gibbins, C. N., E. Scott, C. Soulsby, & I. McEwan, 2005. The relationship between sediment mobilisation and the entry of *Baetis* mayflies into the water column in a laboratory flume. *Hydrobiologia* **533**: 115–122.
- Gibbins, C. N., D. Vericat, & R. J. R. Batalla, 2007. When is stream invertebrate drift catastrophic? The role of hydraulics and sediment transport in initiating drift during flood

events. *Freshwater Biology* **52**: 2369–2384.

Giller, P. S., & B. Malmqvist, 1998. *The biology of streams and rivers*. Oxford University Press.

Hay, C. H., T. G. Franti, D. B. Marx, E. J. Peters, & L. W. Hesse, 2008. Macroinvertebrate drift density in relation to abiotic factors in the Missouri River. *Hydrobiologia* **598**: 175–189.

Hayes, J. W., N. F. Hughes, & L. H. Kelly, 2007. Process-based modelling of invertebrate drift transport, net energy intake and reach carrying capacity for drift-feeding salmonids. *Ecological Modelling* **207**: 171–188.

Holomuzki, J. R., & B. J. F. Biggs, 2003. Sediment texture mediates high-flow effects on lotic macroinvertebrates. *Journal of the North American Benthological Society* **22**: 542–553.

Imbert, B. J., & J. A. Perry, 2000. Drift and benthic invertebrate responses to stepwise and abrupt increases in non-scouring flow. *Hydrobiologia* **436**: 191–208.

James, A., Z. Dexson, & R. Death, 2008. The effect of experimental flow reductions on macroinvertebrate drift in natural and streamside channels. *River Research and Applications* **24**: 22–35.

Kennedy, T. A., C. B. Yackulic, W. F. Cross, P. E. Grams, M. D. Yard, & A. J. Copp, 2014. The relation between invertebrate drift and two primary controls, discharge and benthic densities, in a large regulated river. *Freshwater Biology* **59**: 557–572.

Kohler, S. L., 1983. Positioning on substrates, positioning changes, and diel drift periodicities in mayflies. *Canadian Journal of Zoology* **61**: 1362–1368.

Kohler, S. L., 1985. Identification of stream drift mechanisms: an experimental and observational approach. *Ecology* **66**: 1749–1761.

Lake, P. S., 2000. Disturbance, patchiness, and diversity in streams. *Journal of the North American Benthological Society* **19**: 573–592.

Lancaster, J., 1999. Small-scale movements of lotic macroinvertebrates with variations in flow. *Freshwater Biology* **41**: 605–619.

Lancaster, J., 2000. Geometric scaling of microhabitat patches and their efficacy as refugia during disturbance. *Journal of Animal Ecology* **69**: 442–457.

Lehmkuhl, D. M., & N. H. Anderson, 1972. Microdistribution and density as factors affecting the downstream drift of mayflies. *Ecology* **53**: 661–667.

- Long, A., W. Ashe, K. Ravana, & K. S. Simon, 2011. The effects of water velocity and sediment size on *Acroneuria abnormis* (Plecoptera: Perlidae) entrainment. *Aquatic Insects* **33**: 105–112.
- Matthaei, C. D., & C. R. Townsend, 2000. Long-term effects of local disturbance history on mobile stream invertebrates. *Oecologia* **125**: 119–126.
- Matthaei, C., U. R. S. Uehlinger, & A. Frutiger, 1997. Response of benthic invertebrates to natural versus experimental disturbance in a Swiss prealpine river. *Freshwater Biology* **37**: 61–77.
- Naman, S. M., J. S. Rosenfeld, J. S. Richardson, & J. L. Way, 2016. Species traits and channel architecture mediate flow disturbance impacts on invertebrate drift. *Freshwater Biology* **62**: 340–355.
- Nislow, K. H., F. J. Magilligan, C. L. Folt, & B. P. Kennedy, 2002. Within-basin variation in the short-term effects of a major flood on stream fishes and invertebrates. *Journal of Freshwater Ecology* **17**: 305–318.
- Phillipson, J., 1957. The effect of current speed on the distribution of the larvae of the blackflies, *Simulium variegatum* (Mg.) and *Simulium monticola* Fried (Diptera). *Bulletin of Entomological Research* **48**: 811–819.
- Poff, N. L., & J. V. Ward, 1991. Drift response of benthic invertebrates to experimental streamflow variation in a hydrologically stable stream. *Canadian Journal of Fisheries and Aquatic Sciences* **48**: 1926–1936.
- Radford, D., & R. Hartland-Rowe, 1971. A preliminary investigation of bottom fauna and invertebrate drift in an unregulated and a regulated stream in Alberta. *Journal of Applied Ecology* **8**: 883–903.
- Rempel, L. L., J. S. Richardson, & M. C. Healey, 2000. Macroinvertebrate community structure along gradients of hydraulic and sedimentary conditions in a large gravel-bed river. *Freshwater Biology* **45**: 57–73.
- Richardi, V. S., D. Rebecchi, J. M. R. Aranha, & M. A. Navarro-Silva, 2013. Determination of larval instars in *Chironomus sancticaroli* (Diptera: Chironomidae) using novel head capsule structures. *Zoologia* **30**: 211–216.
- Robinson, C. T., U. Uehlinger, & M. T. Monaghan, 2004. Stream ecosystem response to multiple experimental floods from a reservoir. *River Research and Applications* **20**: 359–377.

- Sagar, P. M., & G. J. Glova, 1992. Diel changes in the abundance and size composition of invertebrate drift in five rivers in South Island, New Zealand. *New Zealand Journal of Marine and Freshwater Research* **26**: 103–114.
- Sánchez-Hernández, J., R. Vieira-Lanero, M. J. M. J. Servia, F. Cobo, J. Sánchez-Hernández, R. Vieira-Lanero, M. J. M. J. Servia, & F. Cobo, 2011. First feeding diet of young brown trout fry in a temperate area: disentangling constraints and food selection. *Hydrobiologia* **663**: 109–119.
- Skinner, W. D., 1985. Night-day drift patterns and the size of larvae of two aquatic insects. *Hydrobiologia* **124**: 283–285.
- Spiegelhalter, D. J., N. G. Best, B. P. Carlin, & A. Linde, 2002. Bayesian measures of model complexity and fit. *Journal of the Royal Statistical Society: Series B (Statistical Methodology)* **64**: 583–639.
- Stewart, K. W., & S. W. Szczytko, 1983. Drift of Ephemeroptera and Plecoptera in two Colorado rivers. *Freshwater Invertebrate Biology* **2**: 117–131.
- Stoneburner, D. L., & L. A. Smock, 1979. Seasonal fluctuations of macroinvertebrate drift in a south carolina piedmont stream. *Hydrobiologia* **63**: 49–56.
- Tachet, H., Richoux, P., Bournaud, M., & P. Usseglio-Polatera, 2010. *Invertébrés d'eau douce: systématique, biologie, écologie* (Vol. 15). Paris: CNRS editions.
- Taniguchi, H., & M. Tokeshi, 2004. Effects of habitat complexity on benthic assemblages in a variable environment. *Freshwater Biology* **49**: 1164–1178.
- Theodoropoulos, C., A. Vourka, A. Stamou, P. Rutschmann, & N. Skoulikidis, 2017. Response of freshwater macroinvertebrates to rainfall-induced high flows: A hydroecological approach. *Ecological Indicators* **73**: 432–442.
- Vericat, D., R. Batalla, & C. N. Gibbins, 2008. Sediment entrainment and depletion from patches of fine material in a gravel-bed river. *Water Resources Research* **44**: 1–15.
- Walton, O. E., S. R. Reice, & R. W. Andrews, 1977. The effects of density, sediment particle size and velocity on drift of *Acroneuria abnormis* (Plecoptera). *Oikos* **28**: 291–298.
- Waters, T. F., 1965. Interpretation of invertebrate drift in streams. *Ecology* **46**: 327–334.
- Woodward, G., N. Bonada, H. B. Feeley, & P. S. Giller, 2015. Resilience of a stream

community to extreme climatic events and long-term recovery from a catastrophic flood. *Freshwater Biology* **60**: 2497–2510.

Wotton, R. S., 1985. The reaction of larvae of *Simulium noelleri* (Diptera) to different current velocities. *Hydrobiologia* **123**: 215–218.

Article references of Chapter 2

Abràmoff, M. D., Magalhães, P. J. & Ram, S. J. (2004) Image Processing with imageJ. *Biophotonics International* **11**, 36–41.

Allan, J. D. & Castillo, M. M. (2007) *Stream Ecology: Structure and Function of Running Waters*. Springer Science & Business Media.

Bal, G., Rivot, E., Baglinière, J.-L., White, J. & Prévost, E. (2014) A Hierarchical Bayesian Model to Quantify Uncertainty of Stream Water Temperature Forecasts. *PLoS ONE* **9**, 1–24.

Bilton, H. T. & Robins, G. L. (1973) The Effects of Starvation and Subsequent Feeding on Survival and Growth of Fulton Channel Sockeye Salmon Fry (*Oncorhynchus nerka*). *J. Fish. Res. Board Can.* **30**, 1–5.

Blaxter, J. H. S. (1969) Development: Eggs and Larvae. *Fish Physiology* **3**, 177–252.

Blaxter, J. H. S. & Hempel, G. (1963) The Influence of Egg Size on Herring Larvae (*Clupea harengus* L.). *ICES Journal of Marine Science* **28**, 211–240.

Dou, S., Masuda, R., Tanaka, M. & Tsukamoto, K. (2002) Feeding Resumption, Morphological Changes and Mortality during Starvation in Japanese Flounder Larvae. *Journal of Fish Biology* **60**, 1363–1380.

Edsall, T. A., Manny, B. A. & Kennedy, G. W. (2003) Starvation Resistance in Lake Trout Fry. *Journal of Great Lakes Research* **29**, 375–382.

Elliott, J. M. (1986) Spatial Distribution and Behavioural Movements of Migratory Trout *Salmo trutta* in a Lake District Stream. *Journal of Animal Ecology* **55**, 907–922.

Elliott, J. M. (1994) *Quantitative Ecology and the Brown Trout*, Oxford Uni. Oxford.

Elliott, J. M. & Hurley, M. A. (2001) Modelling Growth of Brown Trout, *Salmo Trutta*, in Terms of Weight and Energy Units. *Freshwater Biology* **46**, 679–692.

Garcia de Leaniz, C., Fraser, N., Mikheev, V. & Huntingford, F. (1994) Individual Recognition of Juvenile Salmonids Using Melanophore Patterns. *Journal of Fish Biology* **45**, 417–422.

- Gelman, A. & Hill, J. (2006) *Data Analysis Using Regression and Multilevel/hierarchical Models*. Cambridge University Press.
- Giller, P. S. & Malmqvist, B. (1998) *The Biology of Streams and Rivers*. Oxford University Press.
- IPCC (2013) *Climate change 2013: the physical science basis. In: Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change* (Eds T.F. Stocker, D. Qin, G.-K. Plattner, M. Tignor, S.K. Allen, J. Boschung et al.), p. 1265. IPCC, Cambridge, UK and New York, NY.
- Kennedy, B. P., Nislow, K. H. & Folt, C. L. (2008) Habitat-Mediated Foraging Limitations Drive Survival Bottlenecks for Juvenile Salmon. *Ecology* **89**, 2529–2541.
- Koss, D. R. & Bromage, N. R. (1990) Influence of the Timing of Initial Feeding on the Survival and Growth of Hatchery-Reared Atlantic Salmon (*Salmo salar* L.). *Aquaculture* **89**, 149–163.
- Malzahn, A. M., Clemmesen, C. & Rosenthal, H. (2003) Temperature Effects on Growth and Nucleic Acids in Laboratory-Reared Larval Coregonid Fish. *Marine Ecology Progress Series* **259**, 285–293.
- Marandel, L., Lepais, O., Arbenoits, E., Véron, V., Dias, K., Zion, M. & Panserat, S. (2016) Remodelling of the Hepatic Epigenetic Landscape of Glucose-Intolerant Rainbow Trout (*Oncorhynchus mykiss*) by Nutritional Status and Dietary Carbohydrates. *Scientific reports* **6**.
- Marr, D. H. A. (1966) Influence of Temperature on the Efficiency of Growth of Salmonid Embryos. *Nature* **212**, 957–959.
- McGurk, M. D. (1984) Effects of Delayed Feeding and Temperature on the Age of Irreversible Starvation and on the Rates of Growth and Mortality of Pacific Herring Larvae. *Marine Biology* **84**, 13–26.
- Mizushima, N. (2007) Autophagy : Process and Function. *Genes and Development* 2861–2873.
- Nicieza, A. G. & Metcalfe, N. B. (1997) Growth Compensation in Juvenile Atlantic Salmon : Responses to Depressed Temperature and Food Availability. *Ecology* **78**, 2385–2400.
- Panserat, S., Marandel, L., Geurden, I., Veron, V., Dias, K., Plagnes-Juan, E., Pegourié, G., Arbenoits, E., Santigosa, E., Weber, G., et al. (2017) Muscle Catabolic Capacities and Global Hepatic Epigenome Are Modified in Juvenile Rainbow Trout Fed Different Vitamin Levels at

First Feeding. *Aquaculture* **468**, 515–523.

Pfaffl, M. W. (2001) A New Mathematical Model for Relative Quantification in Real-Time RT-PCR. *Nucleic acids research* **29**, e45-e45.

Régnier, T., Bolliet, V., Gaudin, P. & Labonne, J. (2013) Bigger Is Not Always Better: Egg Size Influences Survival throughout Incubation in Brown Trout (*Salmo trutta*). *Ecology of Freshwater Fish* **22**, 169–177.

Robinowitz, J. & White, E. (2010) Autophagy and Metabolism. *Science* **330**, 1344–1348.

Robinson, C. T., Aebischer, S. & Uehlinger, U. (2004) Immediate and Habitat-Specific Responses of Macroinvertebrates to Sequential, Experimental Floods. *Journal of the North American Benthological Society* **23**, 853–867.

Romaniszyn, E. D., Hutchens, J. J. & Wallace, B. J. (2007) Aquatic and Terrestrial Invertebrate Drift in Southern Appalachian Mountain Streams: Implications for Trout Food Resources. *Freshwater Biology* **52**, 1–11.

Salem, M., Silverstein, J., Rexroad, C. E. & Yao, J. (2007) Effect of Starvation on Global Gene Expression and Proteolysis in Rainbow Trout (*Oncorhynchus mykiss*). *BMC genomics* **8**, 328.

Seiliez, I., Panserat, S., Skiba-Cassy, S., Fricot, A., Vachot, C., Kaushik, S. & Tesseraud, S. (2008) Feeding Status Regulates the Polyubiquitination Step of the Ubiquitin-Proteasome-Dependent Proteolysis in Rainbow Trout (*Oncorhynchus mykiss*) Muscle. *The Journal of Nutrition* **138**, 487–491.

Seiliez, I., Gutierrez, J., Salmerón, C., Skiba-Cassy, S., Chauvin, C., Dias, K., Kaushik, S., Tesseraud, S. & Panserat, S. (2010) An in Vivo and in Vitro Assessment of Autophagy-Related Gene Expression in Muscle of Rainbow Trout (*Oncorhynchus mykiss*). *Comparative biochemistry and physiology. Part B, Biochemistry & molecular biology* **157**, 258–266.

Seiliez, I., Gabillard, J. C., Riflade, M., Sadoul, B., Dias, K., Avérous, J., Tesseraud, S., Skiba, S. & Panserat, S. (2012) Amino Acids Downregulate the Expression of Several Autophagy-Related Genes in Rainbow Trout Myoblasts. *Autophagy* **8**, 364–375.

Twongo, T. K. & MacCrimmon, H. R. (1976) Significance of the Timing of Initial Feeding in Hatchery Rainbow Trout, *Salmo gairdneri*. *Journal of the Fisheries Research Board of Canada* **33**, 1914–1921.

Ward, D. M., Nislow, K. H. & Folt, C. L. (2009) Increased Population Density and Suppressed Prey Biomass: Relative Impacts on Juvenile Atlantic Salmon Growth. *Transactions of the American Fisheries Society* **138**, 135–143.

Wing, S. S., Haas, A. L. & Goldberg, A. L. (1995) Increase in Ubiquitin-Protein Conjugates Concomitant with the Increase in Proteolysis in Rat Skeletal Muscle during Starvation and Atrophy Denervation. *The Biochemical journal* **307**, 639–645.

Article references of Chapter 3

Adams, C. E., Huntingford, F. A., Krpal, J., Jobling, M., & Burnett, S. J. (1995). Exercise, agonistic behaviour and food acquisition in Arctic charr, *Salvelinus alpinus*. *Environmental Biology of Fishes*, **43**(2), 213-218.

Adams, C. E., Huntingford, F. A., Turnbull, J. F., & Beattie, C. (1998). Alternative competitive strategies and the cost of food acquisition in juvenile Atlantic salmon (*Salmo salar*). *Aquaculture*, **167**(1), 17-26.

Arunachalam, M., Nair, K. M., Vijverberg, J., Kortmulder, K., & Suriyanarayanan, H. (1991). Substrate selection and seasonal variation in densities of invertebrates in stream pools of a tropical river. *Hydrobiologia*, **213**(2), 141-148.

Biro, P. A., Post, J. R., & Parkinson, E. A. (2003). Density-dependent mortality is mediated by foraging activity for prey fish in whole-lake experiments. *Journal of Animal Ecology*, **72**(4), 546-555.

Brännäs, E., Jonsson, S., & Lundqvist, H. (2003). Influence of food abundance on individual behaviour strategy and growth rate in juvenile brown trout (*Salmo trutta*). *Canadian journal of zoology*, **81**(4), 684-692.

Brewin, P. A., Buckton, S. T., & Ormerod, S. J. (2000). The seasonal dynamics and persistence of stream macroinvertebrates in Nepal: do monsoon floods represent disturbance? *Freshwater Biology*, **44**(4), 581-594.

Brown, J. L. (1964). The evolution of diversity in avian territorial systems. *The Wilson Bulletin*, 160-169.

Carpenter, F. L. (1987). Food abundance and territoriality: to defend or not to defend? *American Zoologist*, **27**(2), 387-399.

- Chapman, D. W. (1962). Aggressive behavior in juvenile coho salmon as a cause of emigration. *Journal of the Fisheries Board of Canada*, **19**(6), 1047-1080.
- Cushing, D. H. (1972). The production cycle and the numbers of marine fish. *Conservation and productivity of natural waters*, **29**, 213-232.
- Cutts, C. J., Brembs, B., Metcalfe, N. B., & Taylor, A. C. (1999). Prior residence, territory quality and life-history strategies in juvenile Atlantic salmon (*Salmo salar* L.). *Journal of Fish Biology*, **55**(4), 784-794.
- Einum, S., & Fleming, I. A. (1999). Maternal effects of egg size in brown trout (*Salmo trutta*): norms of reaction to environmental quality. *Proceedings of the Royal Society of London B: Biological Sciences*, **266**(1433), 2095-2100.
- Garcia de Leaniz, C., Fraser, N., Mikheev, V., & Huntingford, F. (1994). Individual recognition of juvenile salmonids using melanophore patterns. *Journal of fish biology*, **45**(3), 417-422.
- Grant, J. W. A. (1997). Territoriality. *Behavioural ecology of teleost fishes*, 81-103.
- Grant, J. W., & Kramer, D. L. (1990). Territory size as a predictor of the upper limit to population density of juvenile salmonids in streams. *Canadian Journal of Fisheries and Aquatic Sciences*, **47**(9), 1724-1737.
- Grant, J. W., Girard, I. L., Breau, C., & Weir, L. K. (2002). Influence of food abundance on competitive aggression in juvenile convict cichlids. *Animal Behaviour*, **63**(2), 323-330.
- Grant, J. W., Steingrímsson, S. Ó., Keeley, E. R., & Cunjak, R. A. (1998). Implications of territory size for the measurement and prediction of salmonid abundance in streams. *Canadian Journal of Fisheries and Aquatic Sciences*, **55**(S1), 181-190.
- Grant, J. W., Steingrímsson, S. Ó., Keeley, E. R., & Cunjak, R. A. (1998). Implications of territory size for the measurement and prediction of salmonid abundance in streams. *Canadian Journal of Fisheries and Aquatic Sciences*, **55**(S1), 181-190.
- Grosholz, E., & Gallo, E. (2006). The influence of flood cycle and fish predation on invertebrate production on a restored California floodplain. *Hydrobiologia*, **568**(1), 91-109.
- Harwood, A. J., Armstrong, J. D., Metcalfe, N. B., & Griffiths, S. W. (2003). Does dominance status correlate with growth in wild stream-dwelling Atlantic salmon (*Salmo salar*)? *Behavioral Ecology*, **14**(6), 902-908.

- Hildebrand, S. G. (1974). The relation of drift to benthos density and food level in an artificial stream. *Limnology and Oceanography*, **19**(6), 951-957.
- Höjesjö, J., Johnsson, J. I., & Bohlin, T. (2002). Can laboratory studies on dominance predict fitness of young brown trout in the wild? *Behavioral Ecology and Sociobiology*, **52**(2), 102-108.
- Huntigford, F. A., & Leaniz, C. G. (1997). Social dominance, prior residence and the acquisition of profitable feeding sites in juvenile Atlantic salmon. *Journal of Fish Biology*, **51**(5), 1009-1014.
- Huntingford, F. A., Metcalfe, N. B., & Thorpe, J. E. (1993). Social status and feeding in Atlantic salmon *Salmo salar* parr: the effect of visual exposure to a dominant. *Ethology*, **94**(3), 201-206.
- Imre, I., Grant, J. W. A., & Keeley, E. R. (2004). The effect of food abundance on territory size and population density of juvenile steelhead trout (*Oncorhynchus mykiss*). *Oecologia*, **138**(3), 371-378.
- Imre, I., Grant, J. W., & Keeley, E. R. (2002). The effect of visual isolation on territory size and population density of juvenile rainbow trout (*Oncorhynchus mykiss*). *Canadian Journal of Fisheries and Aquatic Sciences*, **59**(2), 303-309.
- IPCC (2013) Climate change 2013: the physical science basis. In: Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change (Eds T.F. Stocker, D. Qin, G.-K. Plattner, M. Tignor, S.K. Allen, J. Boschung et al.), p. 1265. IPCC, Cambridge, UK and New York, NY.
- Johnsson, J. I., Nöbbelin, F., & Bohlin, T. (1999). Territorial competition among wild brown trout fry: effects of ownership and body size. *Journal of Fish Biology*, **54**(2), 469-472.
- Kalleberg, H. (1958). Observations in a stream tank of territoriality and competition in juvenile salmon and trout. *Inst. Freshw. Res. Drottningholm Rep.*, **39**, 55-98.
- Katano, O. (1985). Aggressive behaviour and dominance relationships of the dark chub, *Zacco temmincki* with special reference to their individual recognition. *Japanese Journal of Ichthyology*, **32**(2), 225-238.
- Keeley, E. R. (2000). An experimental analysis of territory size in juvenile steelhead trout. *Animal Behaviour*, **59**(3), 477-490.

- Kennedy, T. A., Yackulic, C. B., Cross, W. F., Grams, P. E., Yard, M. D., & Copp, A. J. (2014). The relation between invertebrate drift and two primary controls, discharge and benthic densities, in a large regulated river. *Freshwater Biology*, **59**(3), 557-572.
- Maclean, A., & Metcalfe, N. B. (2001). Social status, access to food, and compensatory growth in juvenile Atlantic salmon. *Journal of Fish Biology*, **58**(5), 1331-1346.
- McLay, C. L. (1968). A study of drift in the Kakanui River, New Zealand. *Marine and Freshwater Research*, **19**(2), 139-150.
- McMullen, L. E., & Lytle, D. A. (2012). Quantifying invertebrate resistance to floods: a global-scale meta-analysis. *Ecological Applications*, **22**(8), 2164-2175.
- Metcalfe, N. B. (1986). Intraspecific variation in competitive ability and food intake in salmonids: consequences for energy budgets and growth rates. *Journal of Fish Biology*, **28**(5), 525-531.
- Metcalfe, N. B., & Thorpe, J. E. (1992). Early predictors of life-history events: the link between first feeding date, dominance and seaward migration in Atlantic salmon, *Salmo salar* L. *Journal of Fish Biology*, **41**, 93-99.
- Miller, A. M., & Golladay, S. W. (1996). Effects of spates and drying on macroinvertebrate assemblages of an intermittent and a perennial prairie stream. *Journal of the North American Benthological Society*, **15**(4), 670-689.
- Nakano, S., & Furukawa-Tanaka, T. (1994). Intra- and interspecific dominance hierarchies and variation in foraging tactics of two species of stream-dwelling charrs. *Ecological Research*, **9**(1), 9-20.
- Nakano, S., & Murakami, M. (2001). Reciprocal subsidies: dynamic interdependence between terrestrial and aquatic food webs. *Proceedings of the National Academy of Sciences*, **98**(1), 166-170.
- Nislow, K. H., Magilligan, F. J., Folt, C. L., & Kennedy, B. P. (2002). Within-basin variation in the short-term effects of a major flood on stream fishes and invertebrates. *Journal of Freshwater Ecology*, **17**(2), 305-318.
- Olsen, E. M., & Vøllestad, L. A. (2001). An evaluation of visible implant elastomer for marking age-0 brown trout. *North American Journal of Fisheries Management*, **21**(4), 967-970.

Skoglund, H., & Barlaup, B. T. (2006). Feeding pattern and diet of first feeding brown trout fry under natural conditions. *Journal of Fish Biology*, **68**(2), 507-521.

Slaney, P. A., & Northcote, T. G. (1974). Effects of prey abundance on density and territorial behavior of young rainbow trout (*Salmo gairdneri*) in laboratory stream channels. *Journal of the Fisheries Board of Canada*, **31**(7), 1201-1209.

Sloman, K. A., Gilmour, K. M., Taylor, A. C., & Metcalfe, N. B. (2000). Physiological effects of dominance hierarchies within groups of brown trout, *Salmo trutta*, held under simulated natural conditions. *Fish Physiology and Biochemistry*, **22**(1), 11-20.

Sumner, W. T., & Fisher, S. G. (1979). Periphyton production in Fort River, Massachusetts. *Freshwater Biology*, **9**(3), 205-212.

Symons, P. E. (1968). Increase in aggression and in strength of the social hierarchy among juvenile Atlantic salmon deprived of food. *Journal of the Fisheries Board of Canada*, **25**(11), 2387-2401.

Titus, R. G. (1990). Territorial behavior and its role in population regulation of young brown trout (*Salmo trutta*): new perspectives. In *Annales Zoologici Fennici* (pp. 119-130). Finnish Zoological Publishing Board, formed by the Finnish Academy of Sciences, Societas Scientiarum Fennica, Societas Biologica Fennica Vanamo and Societas pro Fauna et Flora Fennica.

Titus, R. G., & Mosegaard, H. (1991). Selection for growth potential among migratory brown trout (*Salmo trutta*) fry competing for territories: evidence from otoliths. *Canadian Journal of Fisheries and Aquatic Sciences*, **48**(1), 19-27.

Toobaie, A., & Grant, J. W. (2013). Effect of food abundance on aggressiveness and territory size of juvenile rainbow trout, *Oncorhynchus mykiss*. *Animal behaviour*, **85**(1), 241-246.

Van Leeuwen, T. E., Hughes, M. R., Dodd, J. A., Adams, C. E., & Metcalfe, N. B. (2015). Resource availability and life-history origin affect competitive behavior in territorial disputes. *Behavioral Ecology*, **27**(2), 385-392.

Photo credit: Stéphane Glise (INRA) & Elorri Arevalo