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7	Does expansion of the introduced Manila clam Ruditapes philippinarum
8	cause competitive displacement of the European native clam Ruditapes
9	decussatus?
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28 Abstract

29

30 In several estuaries or lagoons of Europe the introduced Manila clam *Ruditapes* philippinarum has supplanted the native grooved carpet shell clam Ruditapes decussatus 31 by occupying almost entirely its ecological niche and relegating it to restricted areas. 32 However, it is not clear if the nonindigenous clam is the direct responsible of this 33 predominance patterns. Within this context, the main goal of the present study was to 34 analyze the competitive interaction between the non indigenous Manila clam and the 35 native carpet shell clam to determine whether this interaction could impact directly 36 growth and mortality of the native clam populations. The effect of exposure to predators 37 on both species was also examined. For this purpose, between May 2010 and May 2011 a 38 field experiment was conducted on an intertidal area in the Bay of Santander (N Spain) 39 where both species coexist without an extreme predominance of the introduced species. 40 Relative density of clam species was manipulated in a randomized block experimental 41 design. The results obtained show that (i) the increased density of Manila clam simulating 42 species expansion scenario does not affect growth or mortality of the native clam; (ii) for 43 densities of Manila clam substantially higher than observed in the field, predation plays 44 an essential role in regulating both species populations, limiting their density increase. 45 Moreover, Manila clam appears to be more negatively impacted by predation compared to 46 47 the native clam. Overall, our results indicate that predation has a more significant effect on both populations when compared to competitive interaction. 48

49

50 *Keywords:* Competition, clams, nonindigenous, native, *Ruditapes decussatus*, *Ruditapes*51 *philippinarum*

53 1. Introduction

Biological invasions are natural and necessary for the persistence of life. However, the 54 introduction and spread of invasive species has been identified as a major ecological 55 threat in coastal marine communities (e.g. Cohen & Carlton 1998; Occhipinti-Amborgi, 56 2001; Dawson et al., 2005; Claudet et al., 2010). This is the result of several mechanisms 57 that can affect the native species such as predation, parasitism, herbivores, vectoring of 58 59 pathogens, modification of critical habitat, hybridization, and competition (Simberloff, 2000; Dudas et al., 2005; Occhipinti-Amborgi, 2007). The 1992 'Rio' Convention on 60 61 Biological Diversity included the problems caused by introduced species as a priority 62 item (Mooney and Neville, 2000). Moreover, Elliot (2003) considered that introduced marine organisms can be treated in the same way as chemical pollutants and incites the 63 use of the term biological pollution. Therefore, the invasion research and managers 64 usually consider that nonindigenous origin of a species is relevant to predict and value 65 their negative impacts (Kueffer and Hadorn, 2008). As a consequence, the use of the 66 origin for judging a species is arguable since it could have close affinity to "xenophobia" 67 or "racism" (e.g., Simberloff, 2003; Warren, 2007). Yet, coastal aliens could also play a 68 beneficial role in ecosystem functioning and can result in a net gain of species (Sax and 69 Gaines, 2003) and in an increase of biomass production (Minchin and Rossental, 2002; 70 FAO, 2009). Nevertheless, a precautionary approach has been proposed for introduced or 71 invasive species risk assessment, which assumes that an alien species is problematic until 72 proven otherwise (Simberloff, 2005; Wittenberg and Cock, 2001). 73

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75 In this context, pressure tends to increase on managers to minimize the impacts of76 nonindigenous species, and on scientists to provide control measures (Bax et al. 2001).

77 Consequently, the study of interactions between the nonindigenous and native marine species has become a focus of conservation biology in order to correct identify potentially 78 injurious nonindigenous species. Moreover, research that contributes to gain insight into 79 the effects of the interaction between introduced commercial species and native 80 communities is of a particular significance from a socioeconomic point of view since it 81 can affect positively or negatively the fisheries yield (e.g. Seijo et al., 1998; Reaser et al., 82 2007). In this regard, several species have been deliberately introduced for aquaculture 83 purposes or in order to improve the fisheries yield: the oysters Crassostrea gigas, 84 85 C.ariakensis, C. rivularis, and Ostrea edulis (Grizel and Héral, 1991; Langdon and Robinson, 1991; Mann, 1983), the hard clam Mercenaria mercenaria and the softshell 86 clam Mya arenaria (Gollasch, 2006; Conde et al., 2010) and the Manila clam Ruditapes 87 philippinarum (Breber, 1985). Moreover, the cultures of the Pacific cupped oyster 88 Crassostrea gigas and Manila clam Ruditapes philippinarum were responsible for the 89 introduction of the largest number (60) of non-native invertebrates and algae, often 90 attached to packaging material, fouling the shell or parasitizing bivalve tissues (Mineur et 91 92 al., 2007; Savini et al., 2010).

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Largely due to overfishing and irregular yields of the European native grooved carpet 94 shell clam, the confamilial Manila clam, native to the western Pacific Ocean (Scarlato, 95 1981, Jensen et al., 2004), has been one of the most widely introduced species for 96 aquaculture purposes in Europe. Moreover, Levings et al. (2002) described Manila clams 97 as an ideal fishery species because of their large stock biomass, ease of capture, strong 98 market demand, and rapid depuration of toxins accumulated from algal blooms. It was 99 first introduced between 1972 and 1975 in France and later in England, Spain and Italy 100 (Flassch and Leborgne, 1992; Humphreys, 2010). This species has shown a high 101

adaptability to various coastal environments and its sustainability for aquaculture (Laing
and Child, 1996; Usero et al., 1997; Humphreys et al. 2007). Currently, it has become a
natural population and one of the most commercially exploited bivalve molluscs along the
European coast (i.e. Atlantic coast, Adriatic and Aegean seas) contributing more than
90% to European yields of the two species (Gosling, 2003).

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108 In several disturbed estuaries or lagoons of Europe as Arcachon Bay (France) or the lagoon of Venice (Italy), this species has supplanted the native clam Ruditapes decussatus 109 110 by occupying almost entirely their ecological niche and relegating them to occupy very 111 restricted areas (Aubby, 1993; Marin, 2003; Mistri, 2004; Caill-Milly et al., 2003, 2006). 112 Occhipinti-Ambrogi (2007) stated that the success of the nonindigenous species after 113 establishement is governed by two different filters: a local dispersal filter, mainly connected to invasion pressure (i.e. introduced repeatedly) and a habitat suitability filter 114 (i.e. environmental and community filter), which determine a species to reach stage of 115 116 invasion where it is widespread and dominant. However, this extreme dominance pattern 117 of the introduced Manila clam has not yet been detected in other estuaries of Europe such as in Bahía de Santander, N Spain (Juanes et al., 2012), where both species are exploited 118 119 regularly. In this estuary, regarding the relative presence, areas where (1) both species coexist, (2) nonindigenous species predominates and (3) the native species predominates 120 121 were found (Juanes et al., 2012). Consequently, considering the differentiated stages of 122 invasion of Manila clam in European estuaries and taking into account the absence of 123 studies investigating the interaction with the native carpet shell clam or other biotic interactions, nowadays, the definition of this nonindigenous species as a direct threat for 124 125 the European native clam is too daring.

127 Allien species can negatively affect native populations through interspecific competition, inter alia. Competition can regulate the distribution patterns and the abundance of 128 129 intertidal communities if one species indirectly inhibits the growth of another species 130 inhabiting the same area (Simberloff 2002). Inter-specific competition is, by definition, a negative interaction between two or more species that utilize the same limiting resource 131 132 (Connell 1983). Resource competition, often measured through density-dependant reductions in growth or survival, has been observed in natural bivalve communities 133 (Peterson 1985; Whitlach et al. 1997). However, interspecific competition is relatively 134 135 difficult to demonstrate unequivocally in invaded communities and, although only half the studies published have been experimental (Byers, 2009), they indicate that competition is 136 137 important and common in marine invasions, so that it is necessary to take it into account. 138

Several authors have suggested that competition might be relatively unimportant in soft-139 bottom marine communities, since the habitat have a third dimension (i.e. sediment depth) 140 141 and hence, it may not be as effective a structuring force as on hard bottom shores 142 (Dayton, 1984; Peterson, 1991 and Wilson, 1991). Regarding the non native species Manila clam, few experimental studies have been designed to test directly whether 143 144 resources of any sort are limiting to other infaunal bivalve species or whether competition 145 for limited resources is an important process. Byers (2005) investigated the interaction 146 between Ruditapes philippinarum and the native bivalve Prothoaca staminea and did not 147 found direct effects of Manila clam on Protothaca. Lee (1996) also studied the interaction between Manila clam and Anomalocardia squamosa and did not find intense interaction 148 effects. Similarly, Lum (2011) found a lack of competition between Manila clam and 149 150 varnish clam (Nuttallia obscurata).

152 Despite the above, the assumption that an alien species is problematic until proven otherwise (Simberloff, 2005; Wittenberg and Cock, 2001) is a central tenant of 153 conservation strategies. Therefore, it is essential to study the interaction between Manila 154 155 clam and the European carpet shell clam in order to adopt appropriate management measures for the conservation and sustainable exploitation of these valuable marine 156 resources. In recognition of both an increasing pressure on managers due to the detection 157 of zones were the introduced Manila clam predominates (Juanes et al., 2012) and to the 158 159 drastic declines of the native carpet shell clam occurred in other estuaries of Europe, a 160 first experimental study was conducted in the Bay of Santander (N Spain) to examine the 161 significance of the competition among these two congeneric species and to explore the 162 impact of the nonindigenous Manila clam on the native carpet shell clam. Moreover, the 163 role of predation in the regulation of the expansion of the nonindigenous species was explored. In order to achieve this goal, this study examines the effects of increasing 164 density of Manila clam and predation on mortality and growth of both clam species. 165 166

167 2. Material and methods

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169 2.1. Experimental site

A field experiment was conducted between May 2010 and May 2011 in an intertidal area near the semiactive Manila clam farming zone in Bahía de Santander (Figure 1), the largest along the North coast of Spain (Gulf of Biscay) (2346 Ha). This estuary is classified as morphologically complex and dominated by intertidal areas and tidal dynamics (Galván et al., 2010). The experimental site was selected because: (i) it is on a large and open area containing populations of both Manila clam and grooved carpet shell clam, (ii) it is located approximately 1 km from the nearest public access, making it

effectively inaccessible to the general public and (iii) the experiment could be best 177 safeguarded by shell fishermans over its duration. The sediment on the experimental site 178 was composed by 25.3 % coarse, 51.2 % sand and 24.5 % silt, contained 3.74 % organic 179 180 matter which represents suitable conditions for both species (Bidegain et al., 2012). 181 Distribution and abundances of clam populations in the estuary obtained from Juanes et 182 al. (2012) were used to determine the experimental densities to simulate both natural abundances and the effect of potential expansion of the nonindigenous species. Further 183 184 details of these estuaries and sand flats can be found elsewhere (e.g., Galván et al., 2010; Puente et al., 2002). 185



187 Figure 1 - Bay of Santander. The grey circle represents the location of the experimental188 site.

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190 2.2. Clam collection and laboratory procedures

191 All clams used in this experiment were collected from the same zone in the bay, near the

- 192 experimental site, to reduce the effects of potential genetic variability. As allometric
- 193 relationships of studied species are slightly different (Bald and Borja, 2001,2004; Caill-

194 Milly et al., 2003,2006; Bradbury et al., 2005), the selected adult clams sizes were higher $(36.8 \pm 2.8 \text{ mm})$ for *R. decussatus* than for *R. philippinarum* $(33.5 \pm 3.0 \text{ mm})$, in order to 195 196 have similar biomass for both species in treatments containing the same density. Bivalves 197 were transported to the laboratory in a tank filled with sea water, and maintained at field temperature (16.5°C) in order to minimize physical and/or chemical stress. In the 198 199 laboratory, they were dried, measured (maximum length) and marked with a nail polish. Marked clams were transported again to the field and were allocated to the experimental 200 201 units. One year after the start of the experiment, individuals collected in each enclosure 202 were measured and counted to determine growth and mortality of clams. Additionally, 203 mortality source was examined (1) periodically, during the year when dead individuals or 204 cracked shells were found during enclosure nets maintenance and (2) meticulously, at the 205 end of the experiment when enclosures were excavated. For the clams that were recovered with the identification markings, from both covered and uncovered enclosures, it was 206 207 discerned the source of mortality (i.e. cracked shells=predation; empty valves intact and 208 blackened=anoxia or disease, unrecovered clams also assumed dead by a predator). 209

210 2.3. Experimental design

211 A factorial experiment was designed with two fixed factors, relative density of clam 212 species and predation, to examine their effect on growth and mortality of both clams. 213 Relative density factor includes three levels as follows: (1) R. decussatus (RD) or R. 214 philippinarum (RP) monoespecific, including 10 individuals (2) Coexistence, with 10 individuals of RD + 10 individuals of RP and (3) predominance of Manila clam, including 215 10 individuals of *RD* and 20 individuals of *RP*. Whilst, predation includes two levels: 216 exposure (uncovered) and no exposure (covered) to predation. Therefore, the experiment 217 examined a total of eight treatments. Density levels were selected to simulate (i) zones 218

219 containing only one species at natural densities, (ii) zones where both species coexist in moderate but higher density levels (augmented by ~ 50 %) than quantified in the field 220 surveys and (iii) zones where R. philippinarum predominate (augmented x 4 natural 221 222 densities observed in the field). The latter two levels, but in particular the Manila clam predominance scenario, simulated the effect of potential expansion of the nonindigenous 223 224 population in the bay due to a hypothetical situation where it forms widespread and 225 dominant populations in the whole Bay after repetitive and intense introductions (see 226 invasive stages, Occhipinti-Amborgi, 2007).

227

228 The experiment was laid out in a randomized block design were the treatments were 229 replicated in three blocks for a total of 24 plots (Figure 2). Assuming that species follow 230the Lotka-Volterra equations three replicas are sufficient to determine if two species compete (Tilman, 1987). Blocks were spaced 10 cm apart to also ensure homogeneity of 231 environmental conditions between them. Although treatments were placed randomly 232 233 regarding density treatments, a covered and uncovered treatments were systematically 234 interspersed to reduce potential biases from predators (i.e. crabs) foraging in uncovered enclosures and spilling over onto an adjacent uncovered enclosure (Figure 2). 235

236

The treatments were examined in plots consisting of nylon net enclosures (0.25 m², height=15 cm), located along a tidal height contour of ~ 1 m above MLLW. For exposure to predator treatments, enclosures were both laterally and top covered with 8.0 mm mesh size nylon net. Enclosures were inserted in to the substrate beyond typical maximum burial depths (~ 2 - 12 cm) for these species (Vilela, 1950; Lee, 1996; Gosling, 2003) to avoid lateral migrations of clams out of the experimental plot. Top nets tried to exclude predators (e.g. fish, crabs and birds) from the enclosure. For no exposure treatments enclosures were not top covered. Prior to initiating the experiment the sediment below
each enclosure was sieved to remove predators and clams. The area was refilled and
clams were placed on the surface until buried in the sand. Top nylon nets of enclosures
were cleaned every two-three weeks to keep the mesh free of algae and other fouling
organisms.



Figure 2 –Layout for the randomized block design (b1, b2, b3) to examine the effect of
increasing density of *R. philippinarum* and predation in growth and mortality of both
species. Covered and uncovered treatments were systematically interspersed within each
block.

254

Additional plots were added in order to demonstrate the absence of effects of both 255 256 enclosures and disturbance of the sediment on clam responses. For this purpose, three replicates for each effect analysis were located ~ 1m from the experimental array. On the 257 one hand, for the enclosures effect analysis, the sediment was sieved from an area of 0.50 258 m^2 , to remove predators and clams as it was done in the original experiment. Then the 259 260area was refilled and 20 measured and marked individuals of each species were placed in 261three replicates until buried. No enclosures were placed above them. This density of 262clams was selected to mirror the coexistence density treatment of uncovered enclosures; 263 although a higher number of clams (x 2) were added on a larger area (x 2). This difference 264 was assumed in order to recover individuals at the end of the experiment, considering that in absence of enclosures a certain lateral migration of clams could be expected. On the 265

266 other hand, for the disturbance effect analysis, 10 marked individuals of each species 267 were added to areas of 0.25 m^2 where no marks of infaunal organisms (e.g. sand crabs, 268 polychaetes, clams, razor clams) were detected. Thus, without disturbing the sediment, 269 clams were placed on the surface, and we waited until sure that they were buried in the 270 sand. Then the enclosures with top nylon net were inserted above.

271

272 2.4. Data analysis

273 Growth and mortality response variables were examined at the end of the experiment (1 274 year). For this purpose, clams were recovered from enclosures, and then counted and 275 measured. The difference between the initial maximum length size and the final size of 276 individuals was used to estimate growth for each species and for each treatment. 277 Mortality was estimated as the proportion of dead individuals in each treatment. Growth 278 and mortality distributions normality was checked for statistical analyses. Additionally, 279 the proportion of dead individuals corresponding to each mortality source (i.e. predation 280 or anoxia or disease) was estimated in order to know the role of predation. Normality of 281 the growth and mortality distributions was checked for statistical analyses. Then, two-282 way ANOVA was used to examine the effect of relative density and predation (fixed 283 factors) on the response variables and, additionally, to test the effect of interaction 284 between the two factors. In addition, t-Test was applied to analyze differences in growth 285 and mortality between species.

286

In order to demonstrate the absence of both enclosures and disturbed sediment on growth and mortality, a one way ANOVA test was used. First, to analyze the effect of the enclosure, growth and mortality were compared between individuals placed in uncovered enclosures and individuals placed in patches with no enclosure. Second, to examine the 291 effect of the sediment disturbance, response variables were compared between individuals

292 placed in covered enclosures with disturbed sediment and individuals placed in covered

293 enclosures with untouched sediment.

294

295 3. Results

296 **3.1. Density effect on growth and mortality of clams**

- 297 Increasing the density of *R. philippinarum* had no effect on the growth of either species
- 298 (Table 1). The nonindigenous species grew significantly more than the native *R*.
- 299 *decussatus* (t = -12.9, df = 126, p<0.0001). Whilst *R. philippinarum* grew 5.7 mm (\pm 1.6
- 300 SD), R. decussatus grew 2.4 mm (± 1.1 SD)(Fig. 3a, Covered). Moreover, R.
- 301 *philippinarum* density also had no effect on the mortality of none of the species (Table 1).

302 Both species showed similar mean values of mortality 0.42 (\pm 0.12 SD) for *R. decussatus*

303 and 0.37 (\pm 0.13 SD) for *R. phillipinarum* (t = 0.93, df = 16, p<0.36) (Fig. 3b, Covered).

304 Similar specific growth patterns and absence of density effects, on both growth and

305 mortality of both species, were also observed on uncovered plots (Figure 3ab,

306 Uncovered).



308 Figure 3 - Growth (a) and mortality (b) of *Ruditapes decussatus (RD)* and *Ruditapes*309 *philippinarum (RP)* (mean ± SD (error bars)) during the experiment (1 year) for three
310 density treatments in covered and uncovered plots: monoescpecific (M), (2) coexistence
311 (C), and (3) predominance of Manila clam (P).

	df	F	р
R. decussatus			
Growth			
Density	2	1.24	0.30
Predation	1	0.15	0.70
Density x			
Predation	2	0.16	0.85
Mortality			
Density	2	0.47	0.65
Predation	1	27.66	0.0001
Density x			
Predation	2	0.35	0.71
R. philippinarum			
Growth			
Density	2	0.57	0.58
Predation	1	3.58	0.06
Density x			
Predation	2	0.05	0.95
Mortality			
Density	2	0.15	0.86
Predation	1	85.76	0.0001
Density x			
Predation	2	0.03	0.97

Table 1 - Effects of varying relative density of *R. philippinarum* and *R. decussatus* and 316 predator exposure on the mortality and shell growth of both species. Differences were 317 considered statistically significant at $p \le 0.01$.

3.2. Predator exposure effects and source of mortality

320 Predator exposure affected *R. decussatus* and *R. philippinarum* differently (Fig. 3b, Uncovered). Whilst mortality did not show differences between species in enclosures 321 322 protected from predators, it was significantly higher for *R. philippinarum* than for *R. decussatus* in uncovered enclosures (t = -4.8, df = 16, p = 0.0002). This augment in 323 percentage of dead individuals was considerably higher for R. philippinarum (from 37% 324 325 to 82 %) than for *R. decussatus* (from 42% to 67 %) on average, considering together all density treatments (Figure 3b). Moreover, exposure to predators had no significant effect 326 on the growth of none of the two species (Table 1). The growth for no top plots was 6.4 327 328 mm (\pm 1.5 SD) for *R. philippinarum* and 2.5 mm (\pm 1.0 SD) for *R. decussatus*. Finally, 329 interaction effects between predation and density were not detected in any of the two 330 response variables for none of the species (Table 1).

331

332 Regarding the source of mortality, in covered enclosures few clams were cracked by

333 predators or missing so that only a low percentage of *R. decussatus* (10%) and *R.*

334 philippinarum (16 %) died due to predation. However, in uncovered enclosures 55% of R.

335 *decussatus* mortality was attributed to crab, fish and other macropredators, compared to a

336 considerably higher 73 % for R. philippinarum (Table 2). Mortality due to anoxia (empty

337 and blackened shells) was higher for *R. decussatus* (75 % of undamaged shells) than for

338 *R. philippinarum* (43 % of undamaged shells).

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	Status of shells (mortality source)		
Species, treatment	Undamaged shells (e.g. anoxia, disease or starvation)	Cracked shells (definitely predator killed)	Missing clams (probably predator killed)
R.decussatus, Covered	34	2	2
R.decussatus, Uncovered	27	8	25
R.philippinarum, Covered	37	4	3
R.philippinarum, Uncovered	26	11	60

348

349 Table 2 - Status of shells of experimental clams and the mortality source discerned following Byers
350 (2005). Initial number of clams was 90 for *R. decussatus* and 120 for *R. philippinarum*.

351 352

353 **3.3. Effect of enclosures and sediment disturbance**

354 None of the species exhibit significant differences between no enclosure and enclosed

355 uncovered plots neither for growth (*R. decussatus*, F = 3.5, df = 1, p = 0.07, *R*.

356 *philippinarum:* F = 0.12, df = 1, p = 0.74) (Fig. 4a) nor for mortality (*R. decussatus*, F =

357 1.0, df = 1, p = 0.37, *R. philippinarum:* F = 0.8, df = 1, p = 0.42) (Fig. 4b). The effect of

358 the disturbance was examined comparing response variables for both species between

359 covered enclosures with disturbed sediment and covered enclosures with untouched

360 sediment free of infaunal invertebrates marks. The 1-way ANOVA test did not show

361 significant differences in any species neither in growth (*R. decussatus*, F = 0.5, df = 1, p =

362 0.48, *R. philippinarum:* F = 0.02, df = 1, p = 0.89) (Fig. 4a) nor in mortality (*R*.

363 *decussatus*, F = 4.0, df = 1, p = 0.12, *R. philippinarum:* F = 0.25, df = 1, p = 0.64) (Fig.

364 4b).



366

Figure 4 - Growth (a) and mortality rate (b) of *Ruditapes decussatus* and *Ruditapes philippinarum* during the field experiment (1 year) in (i) coexistence treatments;
"Uncovered enclosures" (no top enclosures with disturbed sediment) and "Disturbed
sediment" (covered enclosures with disturbed sediment) and (ii) in their respective
"controls"; "No enclosure" to examine effect of enclosure (treatment with no enclosure
and disturbed sediment) and "Undisturbed sediment" to examine effect of sediment
disturbance (treatment with covered enclosure and undisturbed sediment.

374

375 4. Discussion

The results of this study were necessary for two reasons: first, to test for the effects of

377 varying density of the introduced nonindigenous species Ruditapes philippinarum on

378 growth and mortality of the native clam *Ruditapes decussatus*; and second, to explore the 379 effect of predation in the regulation of both populations, with a particular emphasis on the 380 nonindigenous species. For this purpose, the experimental study conducted in the Bay of 381 Santander simulates a scenario of Manila clam expansion, where it would form a widespread and dominant population. Results suggest that, for this potential scenario, the 382 383 nonindigenous clam cannot be defined as an ecological threat regarding competitive 384 interaction and predation has an important effect on the regulation of the expansion of this 385 species.

386

387 The highest density of the introduced species examined (i.e. x 4 natural density observed 388 in the field by Juanes et al., 2012) experiencing no density mitigation by predators 389 (covered enclosures), directly affected neither the growth nor the mortality of the confamilial native carpet shell clam. In fact, no differences were observed in response 390 391 variables of outplanted Manila or carpet shell clams regardless of the density treatments 392 examined (i.e. monoespecific, coexistence and predominance of manila clam). As 393 density-dependant studies are suitable to estimate resource competition between two 394 species (e.g. Peterson 1985, Whitlach et al. 1997; Byers, 2009 and reference therein) the 395 results suggest that clam densities tested in this study did not result in interspecific competition. Results obtained in uncovered plots regarding density effects were similar to 396 397 those found in covered enclosures, yet they should be taken with caution since density 398 they experienced a high density mitigation effect due to predation exposure.

399

Enclosures can modify microenvironmental conditions (Underwood, 1985) and have an
effect on results. However, the enclosures and the sediment disturbance did not show any
substantial effects on response variables studied for none of the species. Byers (2005) also

403 found that the effect of enclosures on mortality and growth of two bivalves were not 404 significant. Although this conclusion permits to extrapolate the results and estimate 405 natural life history properties, some caution should be taken interpreting these results. On 406 the one hand, considering that the experimental site was located near a Manila clam culture area, the location could be affecting the growth rates and survival of clams in the 407 408 neighbour zones (Nizzoli et al., 2005) since high densities of cultivated bivalves are generally considered as "sinks" of oxygen and particulate organic matter (Richard et al., 409 410 2007a,b) and hence, may cause a food shortage and be attractors of macropredators. In 411 this experiment this potential effect was assumed to be similar for different treatments and 412 not intense since observed growth rate for both species was similar to other authors' 413 findings (e.g., Urrutia et al., 1999; Cannas, 2010). On the other hand, the effects of 414 enclosure and sediment disturbance on reproduction or meat content were not estimated and may give additional insight on the short-term dynamics related to food availability. 415 416

417 Competition between both species at Manila clam relatively high densities, simulating a 418 predominance scenario, may not be an essential factor regulating the distribution and 419 abundance of the native species. This is in good agreement with previous studies where 420 there is no evidence that Manila clam has negative effects on native or alien species 421 neither at natural densities (Breber, 2002, Juanes et al., 2012) nor at higher densities than 422 observed in nature where, similarly, a lack of competition between bivalve species has 423 been observed. Black and Peterson (1988) manipulated the densities of large suspension 424 feeding bivalves in three Western Australian sites to test their effect on the density and diversity of smaller bivalves and other invertebrates. The bivalves had no significant 425 426 effect on small bivalves, gastropods, polychaetes, amphipods, other taxa, and all taxa combined, despite maximum treatment densities were x 2 the natural densities. Byers 427

428 (2005) examined the effects of Manila clam densities on the growth, mortality, and fecundity of the native littleneck clam, P. staminea and found that at densities 50% higher 429 than natural ones (up to ~ 200 clams / m²) the Manila clam had no effect on the native 430 431 littleneck clam or on itself. Similarly, Lum (2011) did not observe differences in the growth rates of R. philippinarum and N. obscurata regardless of the relative densities of 432 433 each species (intra-specific competition) or the density of the other species (inter-specific competition), even when clam densities doubled natural densities observed in preliminary 434 435 surveys. Despite using densities larger than natural ones, these authors suggested that the 436 experimental densities used were still not large enough to reach the critical level to trigger 437 competition. Black and Peterson (1988) point out that their one year duration experiment 438 may have been insufficient to permit the small infauna to colonize densely enough for 439 competition to occur. The present experimental study could have a similar result because of densities (80 Manila clam individuals/ m²) insufficient to produce competition. 440 441 However, densities were well above the average found in the Bay of Santander (i.e. 4 times the natural densities of Manila clam found under ambient conditions) and 442 443 considered as sufficient to simulate a scenario of potential expansion of the nonindigenous species. 444

445

Most favoured areas of Venice Lagoon hold densities ~ 1000 individuals / m² (Breber 2002) whilst in Arcachon Bay the maximum density is around ~ 45 individuals / m² (Caill-Milly et al., 2006). Hence, the competition between the two species may not be the main reason for the drastic reductions of the native clam occurred in Arcachon Bay. In the case of the Lagoon of Venice the densities of Manila clam could have been large enough to reach the critical level to trigger competition and drastically impact the native species.

453 cycles altered by the farming of *R. philippinarum* (Bartoli et al., 2001) or invasions by 454 parasites that can caused long lasting or even irreversible consequences (Harvell et al., 455 2002), as in case of *Bonamia ostreae*, a disease of the European native oyster Ostrea 456 edulis, which caused severe decline in their populations and, as a consequence, destruction of native oyster bed ecosystems (Wolff and Reise, 2002). Complementary 457 possible hypotheses have been pointed out to explain the reduction of native species 458 occurred, such as the failure of non specific management measures (e.g. identical 459 460 minimum capture size) adopted during years or decades, leading to an overexploitation of 461 the native species (Juanes et al., 2012) which has some biological disadvantages such as slower growth and less tolerant or resistant species (Spencer, 1991; Mistri, 2004 and 462 463 references therein).

464

Furthermore, the small scale at which we studied the density effects (0.25 m^2) makes us 465 cautious with respect to the interpretation and extrapolation results regarding density 466 467 effects, since large areas with high densities of suspension feeding bivalves are needed for food depletion from the water (Peterson and Black, 1987; Kamermans, 1993). In this 468 respect, Kamermans (1993) found an effect of 10 m² mussel beds on cockle growth while 469 470 no effect could be detected on smaller plots (Kamermans et al., 1992). This indicates that 471 our initial results should be taken with care and that future studies should explore the 472 possibility that density effects might be present by using larger experimental plots or 473 conducting the experiments in proximity of areas (or even better within) with much higher densities (see for example Thrush et al., 1997). 474

475

476 However, on the other hand, three-dimensionality of infaunal species habitat (Peterson,477 1979; Wilson, 1991), different or alternative feeding modes or food sources (Peterson

478 1982; Kamermans et al. 1992), filter feeding and vulnerability to predators (i.e. burial depth) (Peterson and Andre 1980; Byers, 2009) are considered to play important roles 479 making direct interference competition ineffective at relatively high densities. Regarding 480 481 food sources, Bodoy & Plante-Cuny (1984) found that the growth rate of R. decussatus is mainly related to the primary production of the water column, whilst Page and Lastra 482 483 (2003) in their study about diet of intertidal bivalves with stable isotopes proposed that this species use primarily resuspended microbenthos during periods of low phytoplankton 484 concentration, but phytoplankton increases in relative dietary importance during bloom 485 486 events. Meanwhile, also using stable isotopes, Watanabe, et al. (2009) found that R. 487 philippinarum seems to feed mainly on particulated organic matter resuspended from the 488 sediment (i.e. detritus and microphytobenthos).

489

Both species are filter feeders of a food resource that is frequently replenished (i.e., with
tidal cycle and resuspension due to currents) and whose supply is often decoupled from
consumptive pressure by resident organisms. Moreover, burial depth of *R. decussatus* (~
10-12 cm) is considerably higher than that of *R. philippinarum* (~ 2-4 cm) (Vilella, 1950;
Lee, 1996; Gosling 2003). This difference in burial depth is essential in order to interpret
the differences in growth and mortality between species observed in this experiment
which in turns are consistent with current knowledge.

497

Growth of *R. philippinarum* during the experiment (1 year) was ~ 6 mm regardless of the
exposure to predators. For *R. decussatus* it was considerably lower, ~ 2.5 mm. As
commented above, these differences are in line with findings of several authors for the
large clams (~ 35 mm) used in this study (e.g. Spencer et al., 1991; Solidoro et al., 2000;
Urrutia et al., 1999; Cannas, 2010). The energetic efficiency of feeding with a siphon

503 decreases with burial depth (Zaklan and Ydenberg, 1997) and it explains, at least partially, why the more deeply burrowing R. decussatus does not grow as fast as R. 504 philippinarum. With a shallower burial depth Manila clam can filter particles more 505 506 quickly and invests less in the development of its siphons compared to a deeper burrowing clam with a longer siphon. One of the important reasons to introduce the 507 nonindigenous Manila clam worldwide for aquaculture purposes was precisely its fast 508 growth together with the important commercial value (Laing and Child, 1996; Usero et 509 al., 1997). The resulting growth was not significantly different for any species between 510 511 covered and uncovered enclosures. This result is in contradiction to the studies of 512 sublethal effects of predators on clams which confirmed generally that predator presence 513 significantly decreases clam growth (e.g. Irlandi and Peterson, 1991; Nakaoka, 2000).

514

Because of its high vulnerability to predation, it seems that high densities or dramatic 515 expansion of the nonindigenous Manila clam might rarely occur in estuaries with a high 516 517 exposure to macropredators such as the Bay of Santander. Recent findings by Bidegain et al. (2013) suggest that the habitat suitability for this species in the Bay of Santander is 518 also essential in regulating its potential expansion. During the year of the study, each time 519 520 top nylon nets of enclosures were cleaned (i.e. every two-three weeks) the presence of 521 gulls, ovstcathers and crabs around the experimental area was detected at low tides. A 522 considerable abundance of fishes (e.g., gill-head breams, sting rays and mullets), 523 searching for food in the sediment, was also observed during rising tide (Bidegain pers. Obser.). These observations and the results of the predator's exposure experiment are 524 compatible with previous studies where predation on Manila clam was considered to be 525 strong and a great variety of macropredators were described: moonsnails (Euspira lewisi), 526 sea stars (Pisaster spp.), a variety of birds, e.g. diving ducks (Aythya affinis), gulls 527

(Family Laridae), crows (Family Corvidae), oystcachers, (e.g. *Haematopus ostralegus ostralegus*), scoters (Family Anatidae) (Toba et al., 1992; Gillespie et al., 2001; Caldow et al., 2007), *Cancer sp.* crabs (Gillespie et al., 2001; Byers, 2005, Lum, 2011) and bottom fish such as rays (Peterson, 1982, Jamieson et al., 2001). These authors also described the important role of predation on regulation of the nonindigenous species expansion.

The high vulnerability of infaunal species to predation is related with the burial depth 534 535 since clam survival increases with increasing burial depth (Zaklan and Ydenberg, 1997). 536 When clams were exposed to predators the mortality was considerably higher for Manila 537 clam than for carpet shell clam (87 % and 67 % respectively). For R. decussates, 55% of 538 this mortality was attributed to crab, fish and other macropredators, whilst, for R. 539 philippinarum, 73 % of the mortality was attributed to this cause. This higher vulnerability to predation of Manila clam compared with the native carpet shell clam 540 541 agrees with the hypothesis of the authors mentioned above, since the typical burial depths 542 are considerably different between species. Although, the burial depth was not measured 543 meticulously, we noticed that at the end of the experiment, in most cases, the carpet shell clam was found relatively deeper in the sediment than Manila clam. Thus, predators may 544 545 mitigate high R. philippinarum density preying on them more than on R. decussatus, 546 presumably because crabs, birds and fish excavate the shallowly burrowing Manila clam 547 more easily (Seitz et al., 2001). However, while deeper burial depth increases R. 548 *decussatus* protection from predators and humans, it also exposes clams to a more anoxic environment (Byers, 2005). In fact, this species exhibited greater mortality indicative of 549 550 anoxia (i.e. undamaged, blackened shells) than R. philippinarum. 551

554 5. Conclusions

553

To our knowledge this is the first study that has attempted to investigate competitive 555 556 interaction between these species. The results suggest that neither the current distribution of populations (i.e. coexistence) nor a potential spread of the Manila clam in the Bay of 557 Santander can be defined as an ecological threat for the native carpet shell clam regarding 558 the experimental densities. Moreover, predation plays an important role in regulating both 559 560 species populations, but more drastically the expansion of the nonindigenous species and, 561 hence, high densities of this clam species are not expected in this estuary. When 562 extrapolating our experimental densities to other estuaries of Europe, it seems that the 563 competitive interaction between these species is not always the main reason of the drastic 564 decrease of the native clam since our expansion scenario densities were higher than those observed in field (e.g. Bay of Arcachon). However, this finding should be taken with 565 566 caution and it should be recognised that before generalizing from small to estuary scale 567 we should explore competitive mechanisms operating at larger scales. The strength of competitive interactions may be linked to abiotic and biotic processes operating over 568 569 different scales (Schneider, 1994; Thrush et al., 1997), and in fact in other environments 570 changes in density effects have already been shown to be a function of scale (Kamermans et al., 1992; Kamermans, 1993). Moreover, additional experimentation in other estuaries 571 is essential and research including early recruiters and juveniles will help to determine the 572 573 importance of competition in this more critical life phase.

574

575 Conservation strategies of one of the most appreciated resource in many estuaries of
576 Europe, the native carpet shell *Ruditapes decussatus*, requires several quantitative
577 challenges including that of competitive interaction. However, these strategies might be

more focused on aspects other than the potential direct impact of Manila clam, although a dramatic increase in density as it occurred in Venice Lagoon should be avoided. For this purpose, care should be taken in seeding of the nonindigenous species in culture zones i.e. control of pressure of introductions (Carlton, 1996; Lawrence and Cordell, 2010) and specific fishery management measures should be adopted considering the differences in life history attributes.

584

585 With the appropriate conservation and management measures adopted, the nonindigenous 586 species could have beneficial aspects, including (i) enhance of biodiversity and (ii) 587 positive socioeconomic impacts. Moreover, Manila clam could have the potential to be 588 also of considerable benefit to several shorebird populations and other macropredators as 589 suggested by Mieszkowska et al. (2006) and Caldow et al. (2007), since it is more readily 590 consumed due to its shallower burial depth. As a consequence, predation on the native 591 clam may actually decrease. However, this food source could in turn increase predators' 592 abundance and influence nearshore community structure. Thus, since natural systems are highly dynamic and governed by complex nonlinear interactions, conservation measures 593 and strategies may also be re-evaluated over the years to cope with the changes resulting 594 595 from species' spatio-temporal population dynamics.

596

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