Contents lists available at ScienceDirect

Food Hydrocolloids

journal homepage: http://www.elsevier.com/locate/foodhyd

The versatility of collagen and chitosan: From food to biomedical applications

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ARTICLE INFO

Keywords: Chitosan Collagen Food packaging Nutrition Cosmetics Tissue engineering

ABSTRACT

Biodegradable polymers obtained from renewable resources, such as chitosan and collagen, are sustainable alternatives to develop environmentally friendly materials. Due to their abundance, biocompatibility and antimicrobial properties, chitosan and collagen could become a suitable source for food and biomedical applications. In particular, chitosan formulations are used for food packaging purposes to develop intelligent packaging with the aim of providing information about the quality of the packaged product or to prepare active packaging and extend food shelf life. In this regard, chitosan nanoparticles can be used to provide a sustained release of active substances. Regarding collagen, denatured collagen or gelatin is prevalently used in food industry as a food additive, microencapsulating agent or biodegradable packaging material due to its rheological properties and physical versatility. In turn, collagen-derived peptides have revealed antioxidant and antihypertensive activity, among other health beneficial effects for cosmetic and nutraceutical applications. Additionally, collagen is widely used in tissue engineering, also combined with chitosan, to achieve the functional properties required for specific applications in the biomedical field. In this sense, collagen/chitosan scaffolds have been used for bone, cartilage and skin regeneration. This research in the design and processing of materials based on proteins and polysaccharides is leading to great advances in food and biomedical fields.

1. Introduction

Sustainable production has become a critical challenge due to social and environmental concerns together with the growing global population. In particular, valorisation of materials may be a promising solution not only to minimize waste, but also to produce high value-added products (Xiong et al., 2019). In this context, the design and development of innovative products for food, cosmetics and biomedical applications have attracted an increasing interest.

The United Nations defined food waste as the end products of food processing industries that have not been recycled or used for other purposes. These are the non-product flows of raw materials whose economic value is less than the cost of collection and recovery for reuse and, thus, they are discarded as waste (United Nations, 2015). Several initiatives have been implemented worldwide to promote the prevention of food waste in all the life stages. In 2015, the United Nations defined the Sustainable Development Goal (SDG) 12 to ensure sustainable consumption and production patterns with the aim of reducing by half the global per capita food waste at the retail and consumption levels by 2030 and to reduce food losses along production and supply chains, including post-harvest losses (United Nations, 2015). Moreover, the European Commission committed to achieving this objective in the European Action Plan for the Circular Economy (European Commission, 2015), in which industrial ecology concepts, such as "from cradle to cradle" and "zero waste" have been promoted. Therefore, the exploitation of food waste as a source of biopolymers and bioadditives for the creation of a new generation of products has attracted the attention of researchers and industry, promoting the conversion of food industry

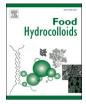
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https://doi.org/10.1016/j.foodhyd.2021.106633

Received 6 December 2020; Received in revised form 24 January 2021; Accepted 25 January 2021 Available online 28 January 2021



Review



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wastes into value-added raw materials (Ong, Kaur, Pensupa, Uisan, & Lin, 2018).

In this context, some opportunities can be generated from byproducts from the slaughter of livestock and the processing of fish. In this regard, industrial processing of livestock generates waste viscera, fat, skin, bones and feet, while industrial fishing waste is mainly composed of muscle, skin, fins, bones, viscera and scales, making these by-products rich sources of various valuable materials, such as proteins, polysaccharides, lipids, minerals, nutrients and flavours (Lee, Patel, Sung, & Kalia, 2020; Martínez-Alvarez, Chamorro, & Brenes, 2015; Meena, Banu, Kannah, Yogalakshmi, & Kumar, 2020). In this sense, natural polymers represent a challenging opportunity for the development of sustainable materials due to their structural and physical characteristics, as well as their safety, availability, biocompatibility and biodegradability (Nasrollahzadeh, Sajjadi, Iravani, & Varma, 2021).

Polysaccharides and proteins provide many possibilities for food and biomedical purposes. Regarding food applications, they can be used as encapsulating agents of flavours, texturizing agents or food thickeners, as well as for food packaging or delivery of bioactives compounds. When employed for biomaterials, they can be used for tissue regeneration or perform as drug carriers; regarding food industry, they show good barrier and mechanical properties for packaging materials (Bealer et al., 2020; Haghighi, Licciardello, Fava, Siesler, & Pulvirenti, 2020). Concerning polysaccharides, chitosan is a copolymer of D-glucosamine and N-acetyl-D-glucosamine (Fig. 1), obtained by the deacetylation of chitin (Rasente et al., 2016). Although it can be obtained from crustaceans, insects, molluscs, fungi or shrimps, the main extraction sources are shrimp and crab exoskeleton residues, producing approximately 2000 tons of chitosan annually (Santos et al., 2020; Sharma & Tiwari, 2020). Chitosan is nontoxic, biocompatible and biodegradable, and it can provide antimicrobial and antioxidant activities (Casadidio et al., 2019).

With regard to proteins, collagen (Fig. 1) can be extracted from skin and bones of animals (Senadheera, Dave, & Shahidi, 2020). The primary sources of commercial collagen are bovine and porcine; however, production has decreased due to bovine spongiform encephalopathy (BSE) and other prions diseases outbreak (Senadheera et al., 2020) and religious considerations for the development of kosher and halal products (Pal & Suresh, 2016). In this sense, seafood by-products are a promising alternative as a collagen source to avoid health, religious and social restrictions (Coppola et al., 2020). The main difference among marine and mammalian collagen is related to the content of hydroxyproline and proline amino acids (Yousefi, Ariffin, & Huda, 2017). Regarding collagen applications, due to its biocompatibility, low antigenicity, high biodegradability and cell growth potential properties, collagen has been widely used in food industry, tissue engineering, pharmaceutical and biomedical industries (Subhan, Ikram, Shehzad, & Ghafoor, 2015). In this context, this work aims to review the latest studies concerning chitosan and collagen biopolymers for food and biomedical applications (Fig. 2).

2. Chitosan for food applications

Chitosan is a suitable material for applications such as food packaging due to its good mechanical properties and capacity of selective permeability to O_2 and CO_2 (Cazón & Vázquez, 2020), which play an important role for protecting food quality during transportation, storage and distribution, when food can be spoiled by chemical and microbiological processes (Sahraee, Milani, Regenstein, & Kafil, 2019). Owing to its antimicrobial activity, chitosan can preserve foods from foodborne pathogens (Shin, Kim, & Shin, 2019). In this regard, the most accepted mechanism of action is the electrostatic interaction between the protonated amine of chitosan and the anionic charges on the microbial surface, which results in a leakage of the cell components and, thus, in the cell necrosis (Amato et al., 2018). Therefore, chitosan is able to prevent microbial spoilage of foods, a major factor that affects shelf life and food quality. With this aim, chitosan can be applied as films or coatings (Fig. 3). Films are preformed layers that can be wrapped around the food (Gudjónsdóttir, Gacutan Jr., Mendes, Chronakis, Jespersen, & Karlsson, 2015) or used as a pouch for foodstuff (Zhang, Liu, et al., 2020), while coatings are thin layers directly formed on food surface by immersing the product in a solution (Yu, Jiang, Xu, & Xia, 2017) or by spraying the solution (Jiang et al., 2020). In this regard, Alemán et al. (2016) studied the shelf life of fish sausages packaged with chitosan-based films and coatings. Results showed that chitosan coatings were imperceptible and able to extend sausage shelf life by 15 days, while sausages packaged with films showed a pickled appearance with lower pH values and water content, and harder texture than coated sausages.

Packaging designed to extend shelf life or provide information about the product's quality is defined as active and/or intelligent packaging (Fig. 4). Active packaging prevents food from deterioration and foodborne pathogens; therefore, it preserves food quality, extends food shelf life, and enhances food safety (Jha, 2020). In this sense, the oxidation of foodstuff is considered a food spoilage factor, which causes discoloration and rancidity affecting food quality negatively (Charles et al., 2021). In food industry, special attention is being paid to natural antioxidant compounds (Talón et al., 2017), such as those containing polyphenols (Zhang, Liang, Li, & Kang, 2020). In this regard, rainbow trout fillets coated with chitosan-Ferulago angulata essential oil reduced the increase of thiobarbituric acid reactive substances (TBARS) during storage at 4 °C, improving fish shelf life up to 16 days (Shokri, Parastouei, Taghdir, & Abbaszadeh, 2020). Bioactive compounds, in spite of enhancing the antioxidant capacity, can also improve other properties of chitosan films. Eucalyptus globulus essential oil incorporated into chitosan films increased the film water resistance and the 2,2-diphenyl-1-picrylhydrazil (DPPH) free radical scavenging capacity up to 43% (Hafsa et al., 2016). Additionally, other natural antioxidants extracted from fruits discards, such as banana peel extracts, reduced water solubility, moisture content and water vapour permeability while increased the antioxidant capacity of chitosan films (Zhang, Li, & Jiang, 2020).

Besides antioxidant capacity, the safety of food is impaired by microbiological activity (Ebadi, Khodanazary, Hosseini, & Zanguee, 2019). Like antioxidants, synthetic antimicrobial compounds, such as sodium nitrite, can generate side effects and health risks for the consumer (Chang, Chen, & Tsai, 2020; De Mey, De Maere, Paelinck, & Fraeye, 2017). Therefore, the food industry seeks substitutes for these compounds from natural sources that do not compromise the sensory properties of the food (Ozaki et al., 2020). In addition to antioxidant activity, essential oils exhibit antimicrobial activity when added to chitosan formulations. For instance, rosemary essential oil inhibited foodborne pathogens, both gram-positive (*Bacillus cereus, Straphylococcus aureus* and *Listeria monocytogenes*) and gram-negative bacteria (*Escherichia coli, Salmonella enterica* and *Pseudomonas aeruginosa*), mainly due to its high content of phenolic compounds (Souza et al.,

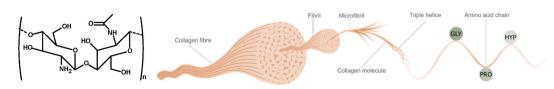


Fig. 1. Structure of chitosan (A) and collagen (B).

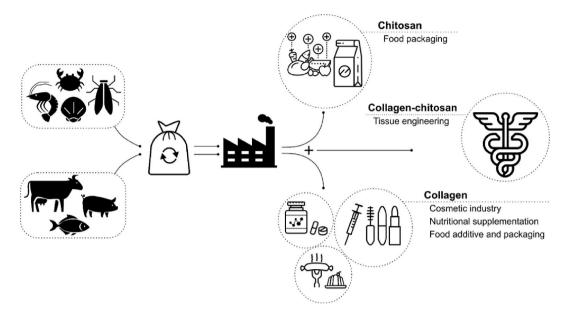


Fig. 2. Food and biomedical applications of chitosan, collagen and their combination.

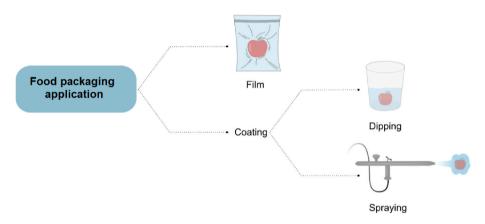


Fig. 3. Schematic representation of chitosan applications for food packaging.

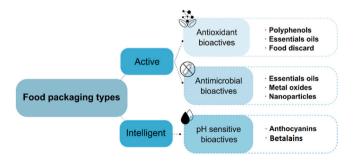


Fig. 4. Schematic representation of different types of chitosan food packaging.

2019). On the other hand, inorganic antimicrobial materials are represented by metal oxides that have higher thermal resistance and broader biocidal spectrum than organic antimicrobials (Al-Tayyar, Youssef, & Al-hindi, 2020). In this regard, zinc oxide (ZnO) nanoparticles decreased initial numbers of *Escherichia coli* by 2.8 log CFU/g and 2.1 log CFU/g in white brined cheese stored at 4 °C and 10 °C, respectively (Al-Nabulsi et al., 2020). In particular, Zn^{2+} ions can attack cell wall, leading to a leakage and, finally, to bacteria death (Yadav, Mehrotra, & Dutta, 2021).

In recent years, chitosan nanoparticles are gaining more attention,

since they can improve functional properties due to their bigger contact surface, which make them more reactive and increase the relative surface of the mass (Istúriz-Zapata, Hernández-López, Correa-Pacheco, & Barrere-Necha, 2020), resulting in the enhancement of antimicrobial activity (Badawy, Lotfy, & Shawir, 2020). Chitosan nanoparticles can be integrated into food formulations, such as those incorporated with cinnamon essential oil for retarding beef patties deterioration during refrigeration storage (Ghaderi-Ghahfarokhi, Barzegar, Sahari, Gavlighi, & Gardini, 2017) or those incorporated with gallic acid and used for Atlantic horse mackerel fillets' coatings (Zarandona et al., 2021). Furthermore, chitosan nanoparticles can be used to provide a sustained release of active substances (Kuai, Liu, Ma, Goff, & Zhong, 2020).

Also recently and due to consumer concerns about the safety and quality of food products, smart packaging has been developed to provide information on food quality of packaged products (Wu, Luo, Liu, Jiang, & Mu, 2019). In general, most smart food packaging has been used to check the freshness of seafood, fish, meat and fruits, using quality indicators such as pH-sensitive colour change films (Merz et al., 2020). The most commonly compounds used as pH indicators are anthocyanins extracted from different food waste. Anthocyanins extracted from purple corn were used in chitosan-silver nanoparticles films, showing pH sensitive properties (Qin, Liu, Yuan, Yong, & Liu, 2019). Additionally, anthocyanins extracted from purple and black eggplant peels were incorporated to chitosan films, resulting effective for the spoilage

control of milk (Yong et al., 2019).

3. Collagen for food and cosmetics industries

Collagen, the main component of the extracellular matrix, is largely used in food and cosmetic industries (Fig. 5) in its native fibrillary form, as well as after denaturation, due to its multiple functional properties and physical versatility to be processed in a variety of products, including gels, porous scaffolds, fibers, films, meshes and micro/nanoparticles (Oliveira et al., 2021). In this context, gelatin, thermally denatured collagen with molecular weight (MW) from 15 to 250 kDa, shows unique rheological properties and is prevalently used in food industry as a food additive, microencapsulating agent, and biodegradable packaging material (Bello, Kim, Kim, Park, & Lee, 2020). Additionally, peptides with biological activity and MW between 300 and 8000 Da can be obtained by several chemical hydrolysis, enzymatic treatment or/and proteolytic fermentation of collagen or gelatin (León-López, Fuentes-Jiménez, Hernández-Fuentes, Camps-Montiel, & Aguirre-Álvarez, 2019). The studies about collagen-derived peptides have revealed their antioxidant and antihypertensive activity, as well as other promising health beneficial effects for cosmetic and nutraceutical applications (Nguyen, Heimann, & Zhang, 2020).

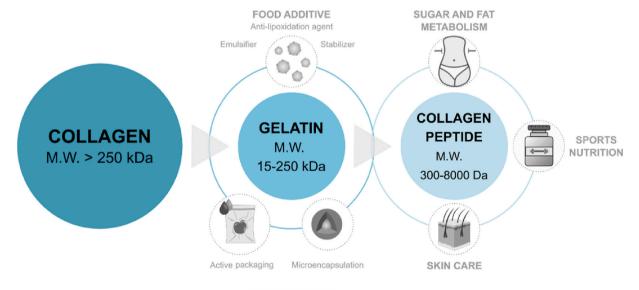
3.1. Gelatin for food industry

Collagen and gelatin have also found wide application in food industry as food additives or packaging materials. Gelatin is incorporated in foods during the food processing to modify colour, texture, flavour and consistence, among others food properties. However, the prevalent applications of gelatin are as food stabilizers and consistence enhancers to form stable gels, emulsions or foaming (Mardani et al., 2019; Yang, Li, Li, Sun, & Guo, 2020). The most popular single use of gelatin as gelling agent may be in water gel desserts, due to its unique melt-in-the-mouth property, but it is also commonly used to form insoluble cross-linked hydrogels that maintain their shape after swelling equilibrium (Huang et al., 2019; Li et al., 2021). Additionally, heat treated collagen fibres have been used as emulsifiers, as natural alternatives to synthetic emulsifiers, especially in acidic products. For example, the incorporation of collagen improves the rheological properties and avoid the fat cap of the oil-in-water emulsions of sausages (Fustier et al., 2015; Huang et al., 2020). In this context, thanks to their antioxidant activity, collagen hydrolysates have been frequently used to inhibit the peroxidation of lipids whose reaction products are dangerous for human health (Bolognesi, Spier, & Rocha Garcia, 2020). This antioxidant activity is generally associated to the radical scavenging capacity of the imidazole group of histidine (Pan et al., 2020).

In recent years, collagen-based films and coatings have played an important role in the development of sustainable packaging materials to protect, maintain, and extend the shelf life of foods (Moreno, Atarés, Chiralt, Cruz-Romero, & Kerry, 2018; Pellá et al., 2020). Generally, food-packaging materials are required to act as a barrier against the migration of oxygen and moisture, as well as to preserve the sensory qualities and prevent fat oxidation, discoloration, and microbial activity (Regubalan, Pandit, Maiti, Nadathur, & Mallick, 2018). In this context, gelatin has been widely studied thanks to its film-forming ability, biodegradability and good gas barrier properties. However, it has poor mechanical strength, and due to its high hygroscopic nature, it tends to swell and dissolve when it is in contact with food with high moisture content, limiting its direct application in food packaging (Liu et al., 2020; Jiang et al., 2020). Therefore, chemical or physical methods, such as crosslinking (Beghetto, Gatto, Conca, Bardella, & Scrivanti, 2019; Tonndorf, Aibibu, & Cherif, 2020; Uranga, Nguyen, Si, Guerrero, & De la Caba, 2020; Wu, Luo, et al., 2019) or combination with other biopolymers (Bhuimbar, Bhagwat, & Dandge, 2019; Hou et al., 2020; Zhuang, Tao, & Cui, 2017), are carried out to improve those properties. Crosslinking reduces the mobility of gelatin chains, improving dimensional stability, water and heat resistance, barrier and mechanical properties (Huang et al., 2019).

The production of sausage casings using coextrusion process has been the best-known industrial application of collagen and gelatin films. However, the strong sensitivity of gelatin to moisture reduces the barrier and mechanical properties of the casings (Chen et al., 2019; Tosati et al., 2017) and multilayered structures can be a strategy to overcome this kind of weakness, where layers with different moisture and oxygen barrier properties are combined in order to comply the required specific package conditions (Figueroa-Lopez, Castro-Mayorga, Andrade-Mahecha, Cabedo, & Lagaron, 2018; Nilsuwan, Guerrero, de la Caba, Benjakul, & Prodpran, 2020; Wang et al., 2020).

In recent years, gelatin has been reported to be one of the first materials used as a carrier of bioactive substances. Gelatin films and coatings can be functionalized with the incorporation of natural antioxidants and antimicrobial components, obtaining active packaging. Many bioactive components have been reported for active packaging but, in the last years, there is a growing interest in using plant extracts like



FOOD PACKAGING

Fig. 5. Collagen application in food and cosmetics industries.

rosemary (Yeddes et al., 2020), grape (Rodrigues, Bertolo, Marangon, Martins, & Plepis, 2020), lemon (Jiang et al., 2020), and oregano (Hernández-Nava, López-Malo, Palou, Ramírez-Corona, Jiménez-Munguía, 2020). Since those extracts may be readily inactivated by exposure to light, heat, and oxygen (Molino, Casanova, Rufián Henares, & Fernandez Miyakawa, 2020), several researchers have tried to embed solid, liquid, or gaseous materials in microcapsules to entrap functional components in a carrier, protect them, and control their release (Mohseni et al., 2019). Moreover, encapsulation may be a useful solution to minimize the taste and odor of some vegetable extracts. Thus, gelatin encapsulation technology could be used to manufacture functional food formulations (Kuai et al., 2020; Paula et al., 2019; Peanparkdee, Yamauchi & Iwamoto, 2018).

3.2. Peptides for cosmetics and nutraceutical industry

Based on the above-mentioned bioactive properties, collagen has also been increasingly utilized for the development of cosmeceutical products, such as skin anti-aging formulations with moisturizing, softening and glowing, antioxidant and UV protective properties (Abuine, Rathnavake, & Byun, 2019; Zamorano-Apodaca et al., 2020). It should be noted that hydrolysed collagen is mainly applied in cosmetic formulations because it offers better effects than native collagen thanks to its superior solubility at neutral pH, water-binding properties, and easy dermis penetration (Alves, Margues, Martins, Silva, & Reis, 2017; Skov, Oxfeldt, Thøgersen, Hansen, & Bertram, 2019). Although hydrolysed collagen can slightly penetrate the epidermis, it cannot replace the lost collagen of the skin (Venkatesan et al., 2017). In this regard, cosmetics industries are interested in subcutaneous injections and oral supplementations. Collagen injections have been popularly used for the repair of dermatological defects, as well as subcutaneous disorders, such as acne scars, and aging symptoms, thanks to collagen biodegradability, price and facility to be produced (Cockerham & Hsu, 2009). When collagen is injected into the dermis, it seems that collagen promotes the biosynthetic capacity of fibroblasts and the formation of an optimal physiologic environment and, thus, increases cell activity, hydration, and the synthesis of collagen (Ganceviciene, Liakou, Theodoridis, Makrantonaki, & Zouboulis, 2012). Alternatively, the positive effect of oral collagen-based supplements on skin appearance has been recently observed (Wang et al., 2018). The fast digestibility of bioactive collagen peptides in the form of liquids, pills or functional foods seems to contribute to the increase of fibroblast density and, hence, to the production of collagen. In that way, several studies have shown the efficacy of the daily bioactive collagen peptides supplementation in the skin hydration, wrinkling, elasticity and density (Genovese, Corbo, & Sibilla, 2017; Ito, Seki, & Ueda, 2019; Kim, Chung, Choi, Sakai, & Lee, 2018).

Besides cosmetics applications, several recent studies have reported the potential of collagen in functional foods and health care applications in form of pills and beverages (Bilek & Bayram, 2015; Guo et al., 2015; Pal & Suresh, 2016). On the one hand, collagen peptides were found to increase the fatty acid metabolism and fats burn, and reduce hypertension and hyperlipidemia. Additionally, they can inhibit the fatty acid synthesis of the liver, decreasing hepatic fat accumulation (Ishak & Sarbon, 2017). On the other hand, they enhanced insulin sensibility and reduced blood sugar levels, which could be attributed to their antioxidant property (Lauritano & Janora, 2016). Therefore, bioactive collagen peptides are used to prevent and treat obesity and type 2 diabetes, inducing the weight loss and reestablishing lipid and blood glucose level (Astre et al., 2018). In this regard, other studies reported that the ingestion of food or drinks enriched with hydrolysed collagen can help on wound healing, bone formation, mineral density and osteoarthritis (Sato, 2017; Suresh, Sugihara, Suzuki, Inoue, & Venkateswarathirukumara, 2015). Collagen is also in demand within the sport nutrition field, nutraceutical industry offers dietary supplements intended to sport nutrition field boost, since it can increase lean muscle, decrease recovery time, reconstruct damaged joint and improve cardiovascular

performance (Oertzen-Hagemann et al., 2019).

4. Chitosan and collagen as materials for biomedical use

Tissue engineering is a multidisciplinary field that combines the knowledge of engineering and biology in order to develop tissue substitutes capable of replace or improve damaged tissues or organs. The methodology of this science is based on the integration of cells, biologically active molecules and materials, recreating in that way the native structure (Langer & Vacanti, 1999). In that context, the material acts as a template to provide structural integrity, define a potential space, guide the restructuring while regeneration, permit diffusion of the nutrients and gas, and provide mechanical characteristics needed for the cell proliferation (Hollister, 2009). In other words, the material attempts to shape the cell microenvironment.

4.1. Characteristics of materials for use in biomedicine

Regardless of the tissue type, a number of key considerations are important when choosing a material for use in tissue engineering (Fig. 6). Therefore, the achievement of these features will determine the suitability of the scaffolds to have the most promising results for the biomaterials that best imitate cells' native physiological environment.

Originally, biomaterials used in the field of biomedicine tended to be inert, such as metals and ceramics, in order to avoid any eventual immune response. However, with the course of time and a better awareness of cellular biology, the use of new materials such as polymers, both natural and artificial, emerged. These materials not only comply with the function of a scaffold, but they are able to interact with the organism, contributing to an active regeneration. A promising approach has been the use of natural polymers, as they have an inherent interaction with the organism, facilitating the development of new tissues while enhancing regeneration. The most widely used among them is collagen, as the major structural component of the native extracellular matrix (O'Brien, 2011). Thus, collagen presents high biocompatibility, biodegradability and malleability. However, collagen by its own forms fiber-like structure scaffolds that exhibit poor mechanical properties, making necessary the modification of the material for its optimal effectiveness (Benavahu et al, 2018, 2020; Pawelec, Best, & Cameron, 2016). One of the strategies employed is the combination of collagen with other natural polymers. The combination of collagen and chitosan

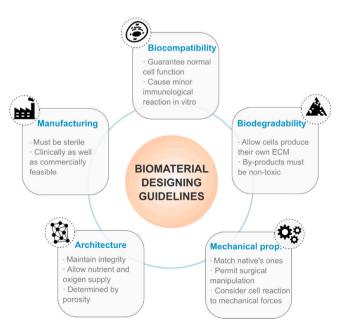


Fig. 6. Criteria to be considered when choosing or designing biomaterials.

is largely employed due to its great potential in tissue engineering applications (Shah, Stodulka, Skopalova, & Saha, 2019). Chitosan offers high biocompatibility, moderate degradation, and antimicrobial effect, among other characteristics. By the addition of chitosan, sheet-like structures with enhanced integrity, mechanical properties and better viability are obtained (Hayashi, Yamada, Guchi, Koyama, & Ikeda, 2012; Hollister, 2009; McBane et al., 2013). According to the literature, the collagen-chitosan construct acquires suitable characteristics when higher concentrations of collagen (>50%) are employed, although a small proportion of chitosan is sufficient to achieve the desired mechanical properties while maintaining cell adhesion and proliferation (Pezeshki-Modaress, Zandi, & Rajabi, 2018).

4.2. Scaffold preparation techniques

The combination of collagen and chitosan is characterized by its high malleability. This feature enables the processing of the materials with a great variety of techniques, allowing a range of forms and structures that can be adapted to the product requirement in each case (Fig. 7).

4.2.1. Films

They are developed by solution casting. The polymer is dissolved in an organic solvent and the solution is poured into a mould. After the casting process, the solvent evaporates. The resulting product shows relatively poor mechanical properties (Shah et al., 2019).

4.2.2. Sponges/porous scaffolds

3D porous structures are mainly obtained by lyophilisation, which produces an interconnected porous structure through the elimination of the ice crystals from the frozen solution. Sponges tend to have poor mechanical properties and low stability in aqueous solutions (Kafi, Aktar, Phanny, & Todo, 2019; Zhang, Zhang, & Wu, 2013).

4.2.3. Hydrogels

These are polymeric networks that absorb and retain large amounts of water. Such formation gives rise to a great capacity of dissolution, so crosslinks are needed to avoid the breakdown of hydrophilic polymer segments in the aqueous phase (Hennink & van Nostrum, 2012).

4.2.4. Fibres

They are based on interconnected structures that resemble the ultrafine fibrous network of the extracellular matrix, thus, they can better promote cell adhesion and proliferation. However, they have poor stability in water due to their fine structure and high surface area. Electrospinning and phase separation are the options for developing this type of material. The latter also provides randomly oriented fibres, which are closer to the 3D structure of the extracellular matrix (Guo et al., 2020).

4.3. Modulation of material properties

Recently, physical and chemical strategies have been implemented in order to ensure that the above-mentioned characteristics are satisfied. On the one hand, mechanical features can be tuned by means of physical or chemical techniques. Those are based on the creation of non-covalent interactions through the use of UV light or temperature, among other agents (Guan et al., 2017). These are inexpensive and easy-to-perform methods that prevent the introduction of possible cytotoxic chemical substances. However, the process is often hard to monitor and fails to achieve a sufficiently high level of cross-linking to meet mechanical requirements (Perez-Puyana, Jiménez-Rosado, Romero, & Guerrero, 2019). This is why the most common approach to overcome the limitations of biomaterials, such as collagen-chitosan scaffolds, is chemical crosslinking. By this method, a chemical compound is added to the material and irreversible covalent bonds are created to interconnect molecules. This approach offers enhanced mechanical properties and an improvement in stability. However, the unreacted crosslinker inside the scaffold can lead to an increase in cytotoxicity together with subsequent processing difficulties caused by the reaction itself (Hennink & van Nostrum, 2012; Reddy, Reddy, & Jiang, 2015; Shah et al., 2019).

There is a great diversity of chemical cross-linking agents according to the degree of improvement desired. In the case of artificial tissues

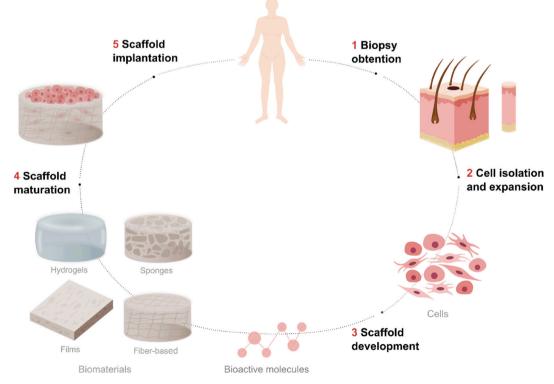


Fig. 7. Schematic representation of the workflow in tissue engineering.

based on collagen and chitosan, one of the most widely used chemical compounds is glutaraldehyde due to its great effectiveness but, even at concentrations higher than 10%, produces cytotoxicity (Liu, Ma, & Gao, 2012; Reddy et al., 2015; Reyna-Urrutia, Mata-Haro, Cauich-Rodriguez, Herrera-Kao, & Cervantes-Uc, 2019). Additionally, tannic acid stands out for its antimicrobial, anti-inflammatory and antioxidant properties (Shah et al., 2019; Sionkowska, Kaczmarek, & Lewandowska, 2014), the combination of N-ethyl-N'-(3-dimethylamino propyl) carbodiimide with N-hydroxysuccinimide (EDC-NHS) enables the maintenance of the porous structure (Martínez, Blanco, Davidenko, & Cameron, 2015; Reddy et al., 2015), and citric acid permits the availability of the binding sites of both biomaterials making feasible further bioconjugations (Reddy et al., 2015; Uranga et al., 2019). Finally, genipin is an aglycone present in the Gardenia jasminoides. This natural compound is effective at low concentrations, creating both intramolecular and intermolecular unions with primary amines of collagen and chitosan, and achieving a totally stable structure that maintains its original porosity (Perez-Puyana et al., 2019; Yan et al., 2010). The properties of collagen-chitosan scaffolds using the abovementioned crosslinking agents are summarized in Table 1.

On the other hand, a major remaining challenge in the design of scaffolds, which directly affect regeneration, is the inadequate interaction between the polymer and the cells, which in vivo may lead to foreign body reactions such as inflammation, infection and implant encapsulation. This problem is correlated with the material surface and the cell recognition motifs that control the interaction between the cells and the substrate. When necessary, cell-binding peptides derived from extracellular matrix proteins can be used to facilitate such interactions between the material and the cell receptors. In this regard, the arginineglycine-aspartic acid (RGD) sequence is currently the most effective and commonly used peptide sequence due to its extensive distribution in the organism, its ability to target more than one cell adhesion receptor, and its biological impact on cell anchorage, behaviour and survival (Hansson et al., 2012; Hersel, Dahmen, & Kessler, 2003). The RGD sequence has been reported to allow the rearrangement of the cellular cytoskeleton, thus facilitating the adhesion and propagation of cells along the collagen-chitosan scaffolds. In vivo, it was demonstrated that a small concentration of RGD is necessary to effectively enhance the cell-material interaction, improving the maturation of the artificial tissue (Hansson et al., 2012; Miklas et al., 2013; Xiao et al., 2013). In many cases these sequences are not accessible in the native conformation of collagen, but collagen denaturalization can lead to conformational changes and to the accessibility of RGD sequences (Pawelec et al., 2016). Along with surface motifs, it has been reported that polarity, surface charge, and surface roughness play a relevant role in the functionality of cells attached to biomaterials.

4.4. Applications of collagen-chitosan scaffolds

Because of their excellent characteristics, particularly their malleability, the combination of collagen and chitosan has been used for the regeneration of a wide variety of tissues. One of the widest applications has been the regeneration of bone. It has been demonstrated that collagen-chitosan membranes have a higher elongation to fracture and lower rate of degradability, critical factors for bone regeneration, than those scaffolds formed only by collagen (Guo et al., 2020). In vitro, the collagen-chitosan combination presents osteoid differentiation capacity, even in the absence of a specific differentiation medium, indicating the capacity of induction to self-differentiation (Georgopoulou et al., 2018; Wang, Wang, Liu, & Zhang, 2016). In vivo, the collagen-chitosan constructs have also demonstrated optimal properties for guided bone regeneration. In this case, electrospun collagen-chitosan membranes allow the formation of new bone in models of calvarial bone defect. By week eight, the cranial defect was completely restored by massive and mature bone tissue (Guo et al., 2020; Lotfi et al., 2016). Similarly, collagen-chitosan scaffolds, crosslinked with carbodiimide, were shown to promote cartilage regeneration in rabbit articular cartilage defects. New cartilage formation was observed as early as at 1 month, and by 6.5 months the cell number, collagen quantity, as well as compressive and storage moduli, approached to the normal cartilage (Whu et al., 2013). Recently, a porous collagen-chitosan scaffold enriched with hydroxyapatite, as a bioactive component, was developed (Campos et al., 2020).

Chitosan-collagen scaffolds approximate very closely the structural hierarchy, organization, biochemical composition, and functional features of native skin extracellular matrix. Furthermore, the combination presents hemostasis and antibacterial properties along with the potential to accelerate the synthesis of extracellular matrix compounds by fibroblast induction, thus, its potential use in skin tissue engineering. The microfiber structures may favour these properties since it has been shown that cross-linked collagen-chitosan microfiber scaffolds can control water loss by evaporation. In addition, the re-epithelization of wounds after 14 days of application of the scaffold, through guided infiltration of fibroblasts and remodelling of collagen in synchrony with the degradation of the scaffold, has been demonstrated (Sarkar, Farrugia, Dargaville, & Dhara, 2013). In skin tissue engineering, the thickness of the graft is crucial, as this influences the effectiveness of the construct implantation. Thinner scaffolds, especially 0.5 mm thick, have been shown to promote ordered fibroblast infiltration and better collagen remodelling. By week 16 after implantation in skin defects, such implants showed total degradation together with complete replacement with new well-arranged host tissue, newly formed vessels, and 75.9% of the tensile strength of native skin (Haifei et al., 2014).

Recently, some attempts have been made to use collagen-chitosan combination in diseases related to the neural tissue such as spinal cord injury. In this case, 3D printed scaffolds could have significant therapeutic effects by bridging axons across the fracture and allowing the mobility of cells, partially re-establishing a microenvironment for axonal regeneration. Once implanted in rat models, nerve-fibre regeneration as well as neurological and locomotor recovery were achieved (Sun et al., 2019).

5. Challenges and new opportunities

Concerning food applications, new opportunities are opening up in the field of nano and intelligent food packaging. The latest studies related to chitosan nanoparticles in the food industry indicate that, due to their bigger contact surface, chitosan nanoparticles can enhance functional properties compared to classical chitosan coatings and,

Table 1

Assessment of different crosslinking agents used in collagen-chitosan scaffolds based on the crosslinking effectivity or level, cytotoxicity, improvement of the biomechanical properties and morphology of the resulting structure. Differences between techniques are shown on a scale of -/+++, with higher crosslinking level, cytotoxicity and biomechanical improvement corresponding to +++.

Agent		Crosslinking level	Cytotoxicity	Biomechanical properties	Remaining structure
Physical	Temperature	_	-	++	No change
	Glutaraldehyde	+++	+	+	Smaller pores
Chemical	Tannic acid	+	-	+	High porosity
	EDC-NHS	+	_	+	No change
	Citric acid	+	-	+	Rougher surface
	Genipin	++	-	+	No change

hence, new methods for food coating are being sought to obtain a greater surface coverage due to a greater penetration of nanoparticles. In this regard, a recent study employed aerosolisation of chitosan nanoparticles for hake fillet coating treatment, which showed that a good coating coverage was achieved with small volumes of solution and the coating had minimal impact on physicochemical parameters (Sullivan et al., 2020). Regarding intelligent food packaging, in addition to anthocyanins, the isolation of new indicators from other natural sources is being analysed. In this sense, betalains have been extracted from vegetable amaranth for application in monitoring of shrimp freshness (Hu, Yao, Qin, Yong, & Liu, 2020). Results suggested that films containing betalains showed good response to the volatile ammonia produced by the shrimp's metamorphism, changing colour when the total volatile basic nitrogen slightly exceeded the limit of the standard (Hu et al., 2020).

In the biomedical field, major efforts are being focused on adapting the properties of collagen and chitosan constructs to clinical demands. In this sense, the development of additive manufacturing techniques has meant a great advance in the field, as they allow the rapid and reproducible manufacture of complex 3D shapes. Recently, the emergence of a novel and innovative 4D printing has allowed the combination of traditional and "smart" materials. These possess features that allow to respond to external stimuli (heat, moisture, light, magnetic field or pH), adapting their properties to the microenvironment (change shape or colour, produce an electrical current, become bioactive, or perform an intended function). 4D printing benefits from the property of smart materials to have dynamic responses and to control the construct spatially and temporally. Moreover, 4D printing eliminates the need for external devices or methods for post-processing. Therefore, 4D printing can cause a disruptive effect in the medical field, since it represents a great potential for non-invasive and remoted-control therapies, such as drug delivery, biosensors or regenerative medicine. Considering that each model in medicine varies from patient to patient, 4D printing could allow the achievement of effective personalized medicine (Lui et al., 2019; Mantha et al., 2019; Piedade, 2019; Shie et al., 2019; Tamay et al., 2019).

Author contributions

Conceptualization, K.C and P.G.; resources, K.C and P.G.; investigation, A.I., I.Z. and M.A.; supervision, K.C and P.G.; writing—original draft preparation, A.I., I.Z. and M.A.; writing—review and editing, K.C and P.G.; funding acquisition, K.C.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgments

Authors thank UPV/EHU (GIU18/154). Ainhoa Irastorza (PRE_2019_1_0031), Iratxe Zarandona (22-2018-00078), and Mireia Andonegi (PRE_2017_1_0025) thank the Basque Government for their fellowships.

References

- Abuine, R., Rathnayake, A. U., & Byun, H.-G. (2019). Biological activity of peptides purified from fish skin hydrolysates. *Fisheries and Aquatic Sciences*, 22, 10. https:// doi.org/10.1186/s41240-019-0125-4
- Al-Nabulsi, A., Osaili, T., Sawalha, A., Olaimat, A. N., Albiss, B. A., Mehyar, G., et al. (2020). Antimicrobial activity of chitosan coating containing ZnO nanoparticles against *E. coli* O157:H7 on the surface of white brined cheese. *International Journal of Food Microbiology*, 334, Article 108838. https://doi.org/10.1016/j. iifoodmicro.2020.108838
- Al-Tayyar, N. A., Youssef, A. M., & Al-hindi, R. (2020). Antimicrobial food packaging based on sustainable bio-based materials for reducing foodborne pathogens: A

review. Food Chemistry, 310, Article 125915. https://doi.org/10.1016/j. foodchem.2019.125915

- Alemán, A., González, F., Arancibia, M. Y., López-Caballero, M. E., Montero, P., & Gómez-Guillén, M. C. (2016). Comparative study between film and coating packaging based on shrimp concentrate obtained from marine industrial waste for fish sausage preservation. *Food Control*, 70, 325–332. https://doi.org/10.1016/j. foodcont.2016.06.007
- Alves, A., Marques, A., Martins, E., Silva, T., & Reis, R. L. (2017). Cosmetic potential of marine fish skin collagen. *Cosmetics*, 4, 39. https://doi.org/10.3390/ cosmetics4040039
- Amato, A., Migneco, L. M., Martinelli, A., Pietrelli, L., Piozzi, A., & Francolini, I. (2018). Antimicrobial activity of catechol functionalized-chitosan versus *Staphylococcus* epidermidis. Carbohydrate Polymers, 179, 273–281. https://doi.org/10.1016/j. carbool.2017.09.073
- Astre, G., Deleruyelle, S., Dortignac, A., Bonnet, C., Valet, P., & Dray, C. (2018). Dietinduced obesity and associated disorders are prevented by natural bioactive type 1 fish collagen peptides (Naticol®) treatment. *Journal of Physiology & Biochemistry*, 74, 647–654. https://doi.org/10.1007/s13105-018-0650-0
- Badawy, M. E. I., Lotfy, T. M. R., & Shawir, S. M. S. (2020). Facile synthesis and characterizations of antibacterial and antioxidant of chitosan monoterpene nanoparticles and their applications in preserving minced meat. *International Journal* of Biological Macromolecules, 156, 127–136. https://doi.org/10.1016/j. iibiomac.2020.04.044
- Bealer, E. J., Onissema-Karimu, S., Rivera-Galletti, A., Francis, M., Wilkowski, J., Salasde la Cruz, D., et al. (2020). Protein-polysaccharide composite materials: Fabrication and applications. *Polymers*, 12, 464. https://doi.org/10.3390/polym12020464
- Beghetto, V., Gatto, V., Conca, S., Bardella, N., & Scrivanti, A. (2019). Polyamidoamide dendrimers and cross-linking agents for stabilized bioenzymatic resistant metal-free bovine collagen. *Molecules*, 24, 3611. https://doi.org/10.3390/molecules24193611
- Bello, A. B., Kim, D., Kim, D., Park, H., & Lee, S.-H. (2020). Engineering and functionalization of gelatin biomaterials: From cell culture to medical applications. *Tissue Engineering Part B Reviews*, 26, 2. https://doi.org/10.1089/ten.TEB.2019.0256
- Benayahu, D., Pomeraniec, L., Shemesh, S., Heller, S., Rosenthal, Y., Rath-Wolfson, L., et al. (2020). Biocompatibility of a marine collagen based scaffold in vitro and in vivo. Marine Drugs, 18, 420. https://doi.org/10.3390/MD18080420
- Benayahu, D., Sharabi, M., Pomeraniec, L., Awad, L., Haj-Ali, R., & Benayahu, Y. (2018). Unique collagen fibers for biomedical applications. *Marine Drugs*, 16, 102. https:// doi.org/10.3390/md16040102
- Bhuimbar, M. V., Bhagwat, P. K., & Dandge, P. B. (2019). Extraction and characterization of acid soluble collagen from fish waste: Development of collagen-chitosan blend as food packaging films. *Journal of Environmental Chemical Engineering*, 7, Article 102983. https://doi.org/10.1016/i.jece.2019.102983
- Bilek, S. E., & Bayram, S. K. (2015). Fruit juice drink production containing hydrolyzed collagen. Journal of Functional Foods, 14, 562–569. https://doi.org/10.1016/j. jff.2015.02.024
- Bolognesi, V. J., Spier, M. R., & Rocha Garcia, C. E. (2020). Brine solution with hydrocolloids used to enhance the properties of sterilized meat. *Food Technology and Biotechnology*, 58, 2. https://doi.org/10.17113/ftb.58.02.20.6336
- Campos, Y., Sola, F. J., Almirall, A., Fuentes, G., Eich, C., Que, I., et al. (2020). Design, construction, and biological testing of an implantable porous trilayer scaffold for repairing osteoarthritic cartilage. *Journal of Tissue Engineering and Regenerative Medicine*, 14, 355–368. https://doi.org/10.1002/term.3001
- Casadidio, C., Peregrina, D. V., Gigliobianco, M. R., Deng, S., Censi, R., & Di Martino, P. (2019). Chitin and chitosans: Characteristics, eco-friendly processes, and applications in cosmetic science. *Marine Drugs*, *17*, 369. https://doi.org/10.3390/ md17060369
- Cazón, P., & Vázquez, M. (2020). Mechanical and barrier properties of chitosan combined with other components as food packaging film. *Environmental Chemistry Letters*, 18, 257–267. https://doi.org/10.1007/s10311-019-00936-3
- Chang, S. H., Chen, C. H., & Tsai, G. J. (2020). Effects of chitosan on *Clostridium perfringens* and application in the preservation of pork sausage. *Marine Drugs*, 18, 70. https://doi.org/10.3390/md18020070
- Charles, A. L., Abdillah, A. A., Saraswati, Y. R., Sridhar, K., Balderamos, C., Masithah, E. D., et al. (2021). Characterization of freeze-dried microencapsulation tuna fish oil with arrowroot starch and maltodextrin. *Food Hydrocolloids*, 112, Article 106281. https://doi.org/10.1016/j.foodhyd.2020.106281
- Chen, X., Zhou, L., Xu, H., Yamamoto, M., Shinoda, M., Tada, I., et al. (2019). The structure and properties of natural sheep casing and artificial films prepared from natural collagen with various crosslinking treatments. *International Journal of Biological Macromolecules*, 135, 959–968. https://doi.org/10.1016/j. ijbiomac.2019.05.182
- Cockerham, K., & Hsu, V. (2009). Collagen-based dermal fillers: Past, present, future. *Facial Plastic Surgery*, 25, 106–113. https://doi.org/10.1055/s-0029-1220650
- Coppola, D., Oliviero, M., Vitale, G. A., Lauritano, C., D'Ambra, I., Iannace, S., et al. (2020). Marine collagen from alternative and sustainable sources: Extraction, processing and applications. *Marine Drugs*, 18, 214. https://doi.org/10.3390/ md18040214
- De Mey, E., De Maere, H., Paelinck, H., & Fraeye, I. (2015). Volatile N-nitrosamines in meat products: Potential precursors, influence of processing and mitigation strategies. *Critical Reviews in Food Science and Nutrition*, 57(13), 2909–2923. https:// doi.org/10.1080/10408398.2015.1078769
- Ebadi, Z., Khodanazary, A., Hosseini, S. M., & Zanguee, N. (2019). The shelf life extension of refrigerated *Nemipterus japonicus* fillets by chitosan coating incorporated with propolis extract. *International Journal of Biological Macromolecules*, 139, 94–102. https://doi.org/10.1016/j.ijbiomac.2019.07.204

- European Commission. (2015). COM/2015/0614 Closing the loop an EU action plan for the Circular Economy. https://eur-lex.europa.eu/legal-content/EN/TXT/?uri=CELEX %3A52015DC0614. (Accessed 6 December 2020).
- Figueroa-Lopez, K. J., Castro-Mayorga, J. L., Andrade-Mahecha, M. M., Cabedo, L., & Lagaron, J. M. (2018). Antibacterial and barrier properties of gelatin coated by electrospun polycaprolactone ultrathin fibers containing black pepper oleoresin of interest in active food biopackaging applications. *Nanomaterials*, 8, 199. https://doi. org/10.3390/nano8040199
- Fustier, P., Achouri, A., Taherian, A. R., Britten, M., Pelletier, M., Sabik, H., et al. (2015). Protein-protein multilayer oil-in-water emulsions for the microencapsulation of flaxseed oil: Effect of whey and fish gelatin concentration. *Journal of Agricultural and Food Chemistry*, 63, 9239–9250. https://doi.org/10.1021/acs.jafc.5b00858
- Ganceviciene, R., Liakou, A. I., Theodoridis, A., Makrantonaki, E., & Zouboulis, C. C. (2012). Skin anti-aging strategies. *Dermato-Endocrinology*, 4(3), 308–319. https:// doi.org/10.4161/derm.22804
- Genovese, L., Corbo, A., & Sibilla, S. (2017). An insight into the changes in skin texture and properties following dietary intervention with a nutricosmeceutical containing a blend of collagen bioactive peptides and antioxidants. *Skin Pharmacology and Physiology*, 30, 146–158. https://doi.org/10.1159/000464470
- Georgopoulou, A., Papadogiannis, F., Batsali, A., Marakis, J., Alpantaki, K., Eliopoulos, A. G., et al. (2018). Chitosan/gelatin scaffolds support bone regeneration. Journal of Materials Science: Materials in Medicine, 29(5), 59. https:// doi.org/10.1007/s10856-018-6064-2
- Ghaderi-Ghahfarokhi, M., Barzegar, M., Sahari, M. A., Gavlighi, H. A., & Gardini, F. (2017). Chitosan-cinnamon essential oil nano-formulation: Application as a novel additive for controlled release and shelf life extension of beef patties. *International Journal of Biological Macromolecules*, 102, 19–28. https://doi.org/10.1016/j. iibiomac.2017.04.002
- Guan, X., Avci-Adali, M., Alarçin, E., Cheng, H., Kashaf, S. S., Li, Y., et al. (2017). Development of hydrogels for regenerative engineering. *Biotechnology Journal*, 12, Article 1600394. https://doi.org/10.1002/biot.201600394
- Gudjónsdóttir, M., Gacutan, M. D., Jr., Mendes, A. C., Chronakis, I. S., Jespersen, L., & Karlsson, A. H. (2015). Effects of electrospun chitosan wrapping for dry-ageing of beef, as studied by microbiological, physicochemical and low-field nuclear magnetic resonance analysis. *Food Chemistry*, 184, 167–175. https://doi.org/10.1016/j. foodchem.2015.03.088
- Guo, L., Harnedy, P. A., Zhang, L., Li, B., Zhang, Z., Hou, H., et al. (2015). In vitro assessment of the multifunctional bioactive potential of Alaska pollock skin collagen following simulated gastrointestinal digestion. *Journal of the Science of Food and Agriculture*, 95, 1514–1520. https://doi.org/10.1002/jsfa.6854
- Guo, S., He, L., Yang, R., Chen, B., Xie, X., Jiang, B., et al. (2020). Enhanced effects of electrospun collagen-chitosan nanofiber membranes on guided bone regeneration. *Journal of Biomaterials Science, Polymer Edition, 31*, 155–168. https://doi.org/ 10.1080/09205063.2019.1680927
- Hafsa, J., ali Smach, M., Khedher, M. R. B., Charfeddine, B., Limem, K., Majdoub, H., et al. (2016). Physical, antioxidant and antimicrobial properties of chitosan films containing *Eucalyptus globulus* essential oil. *Lebensmittel-Wissenschaft und* -*Technologie- Food Science and Technology*, 68, 356–364. https://doi.org/10.1016/j. lwt.2015.12.050
- Haghighi, H., Licciardello, F., Fava, P., Siesler, H. W., & Pulvirenti, A. (2020). Recent advances on chitosan-based films for sustainable food packaging applications. *Food Packaging and Shelf Life*, 26, Article 100551. https://doi.org/10.1016/j. fpsl.2020.100551
- Haifei, S., Xingang, W., Shoucheng, W., Zhengwei, M., Chuangang, Y., & Chunmao, H. (2014). The effect of collagen-chitosan porous scaffold thickness on dermal regeneration in a one-stage grafting procedure. *Journal of the Mechanical Behavior of Biomedical Materials*, 29, 114–125. https://doi.org/10.1016/j.jmbbm.2013.08.031
- Hansson, A., Hashom, N., Falson, F., Rousselle, P., Jordan, O., & Borchard, G. (2012). In vitro evaluation of an RGD-functionalized chitosan derivative for enhanced cell adhesion. Carbohydrate Polymers, 90, 1494–1500. https://doi.org/10.1016/j. carbool.2012.07.020
- Hayashi, Y., Yamada, S., Guchi, K. Y., Koyama, Z., & Ikeda, T. (2012). Chitosan and fish collagen as biomaterials for regenerative medicine. Advances in Food & Nutrition Research. https://doi.org/10.1016/B978-0-12-416003-3.00006-8
- Hennink, W. E., & van Nostrum, C. F. (2012). Novel crosslinking methods to design hydrogels. Advanced Drug Delivery Reviews, 64, 223–236. https://doi.org/10.1016/j. addr.2012.09.009
- Hernández-Nava, R., López-Malo, A., Palou, E., Ramírez-Corona, N., & Jiménez-Munguía, M. T. (2020). Encapsulation of oregano essential oil (Origanum vulgare) by complex coacervation between gelatin and chia mucilage and its properties after spray drying. Food Hydrocolloids, 109, Article 106077. https://doi.org/10.1016/j. foodhyd.2020.106077
- Hersel, U., Dahmen, C., & Kessler, H. (2003). RGD modified polymers: Biomaterials for stimulated cell adhesion and beyond. *Biomaterials*, 24, 4385–4415. https://doi.org/ 10.1016/S0142-9612(03)00343-0
- Hollister, S. J. (2009). Scaffold design and manufacturing: From concept to clinic. Advanced Materials, 21, 3330–3342. https://doi.org/10.1002/adma.200802977
- Hou, C., Gao, L., Wang, Z., Rao, W., Du, M., & Zhang, D. (2019). Mechanical properties, thermal stability and solubility of sheep bone collagen-chitosan films. *Journal of Food Process Engineering*, 43, Article e13086. https://doi.org/10.1111/jfpe.13086
- Huang, T., Tu, Z. C., Zou, Z. Z., Shangguan, X., Sha, X., Wang, H., et al. (2019). Fish gelatin modifications: A comprehensive review. *Trends in Food Science & Technology*, 86, 260–269. https://doi.org/10.1016/j.tifs.2019.02.048
- Huang, T., Tu, Z. C., Zou, Z. Z., Shangguan, X., Wang, H., & Bansal, N. (2020). Gycosylated fish gelatin emulsion: Rheological, tribological properties and its

application as model coffee creamers. *Food Hydrocolloids*, 102, Article 105552. https://doi.org/10.1016/j.foodhyd.2019.105552

- Hu, H., Yao, X., Qin, Y., Yong, H., & Liu, J. (2020). Development of multifunctional food packaging by incorporating betalains from vegetable amaranth (*Amaranthus tricolor L*.) into quaternary ammonium chitosan/fish gelatin blend films. *International Journal of Biological Macromolecules*, 159, 675–684. https://doi.org/10.1016/j. ijbiomac.2020.05.103
- Ishak, N., & Sarbon, N. A. (2018). Review of protein hydrolysates and bioactive peptides deriving from wastes generated by fish processing. *Food and Bioprocess Technology*, 11, 2–16. https://doi.org/10.1007/s11947-017-1940-1
- Istúriz-Zapata, M. A., Hernández-López, M., Correa-Pacheco, Z. N., & Barrera-Necha, L. L. (2020). Quality of cold-stored cucumber as affected by nanostructured coatings of chitosan with cinnamon essential oil and cinnamaldehyde. *Lebensmittel-Wissenschaft* und -Technologie- Food Science and Technology, 123, Article 109089. https://doi.org/ 10.1016/j.lwt.2020.109089
- Ito, N., Seki, S., & Ueda, F. (2019). Effects of composite supplement containing collagen peptide and ornithine on skin conditions and plasma IGF-1 levels-a randomized, double-blind, placebo-controlled trial. *Marine Drugs*, 16, 482. https://doi.org/ 10.3390/md16120482
- Jha, P. (2020). Effect of grapefruit seed extract ratios on functional properties of corn starch-chitosan bionanocomposite films for active packaging. *International Journal of Biological Macromolecules*, 163, 1546–1556. https://doi.org/10.1016/j. iibiomac.2020.07.251
- Jiang, Y., Lan, W., Sameen, D. E., Ahmed, S., Qin, W., Zhang, Q., et al. (2020). Preparation and characterization of grass carp collagen-chitosan-lemon essential oil composite films for application as food packaging. *International Journal of Biological Macromolecules*, 160, 340–351. https://doi.org/10.1016/j.ijbiomac.2020.05.202
- Jiang, Y., Yu, L., Hu, Y., Zhu, Z., Zhuang, C., Zhao, Y., et al. (2020). The preservation performance of chitosan coating with different molecular weight on strawberry using electrostatic spraying technique. *International Journal of Biological Macromolecules*, 151, 278–285. https://doi.org/10.1016/j.ijbiomac.2020.02.169
- Kafi, A., Aktar, K., Phanny, Y., & Todo, M. (2019). Adhesion, proliferation and differentiation of human mesenchymal stem cell on chitosan/collagen composite scaffold. Journal of Materials Science: Materials in Medicine, 30, 131. https://doi.org/ 10.1007/s10856-019-6341-8
- Kim, D. U., Chung, H. C., Choi, J., Sakai, Y., & Lee, B. Y. (2018). Oral intake of lowmolecular weight collagen peptide improves hydration, elasticity, and wrinkling in human skin: A randomized, double-blind, placebo-controlled study. *Nutrients, 10*, 826. https://doi.org/10.3390/nu10070826
- Kuai, L., Liu, F., Ma, Y., Goff, H. D., & Zhong, F. (2020). Regulation of nano-encapsulated tea polyphenol release from gelatin films with different Bloom values. *Food Hydrocolloids*, 108, Article 106045. https://doi.org/10.1016/j. foodhvd.2020.106045
- Langer, R. S., & Vacanti, J. P. (1999). Tissue engineering: The challenges ahead. Scientific American. https://doi.org/10.1038/scientificamerican0499-86
- Lauritano, C., & Ianora, A. (2016). Marine organisms with anti-diabetes properties. Marine Drugs, 14, 220. https://doi.org/10.3390/md14120220
- Lee, J. K., Patel, S. K. S., Sung, B. H., & Kalia, V. C. (2020). Biomolecules from municipal and food industry wastes: An overview. *Bioresource Technology*, 298, Article 122346. https://doi.org/10.1016/j.biortech.2019.122346
- León-López, A., Fuentes-Jiménez, L., Hernández-Fuentes, A. D., Camps-Montiel, R. G., & Aguirre-Álvarez, G. (2019). Hydrolysed collagen from sheepskins as a source of functional petides with antioxidant activity. *International Journal of Molecular Sciences*, 20, 3931. https://doi.org/10.3390/ijms20163931
- Liu, B., Huang, W., Yang, G., An, Y., Yin, Y., Wang, N., et al. (2020). Preparation of gelatin/poly(γ-glutamic acid) hydrogels with stimulated response by hot-pressing preassembly and radiation crosslinking. *Materials Science and Engineering: C, 116*, Article 111259. https://doi.org/10.1016/j.msec.2020.111259
- Liu, Y., Ma, L., & Gao, C. (2012). Facile fabrication of the glutaraldehyde cross-linked collagen/chitosan porous scaffold for skin tissue engineering. *Materials Science and Engineering: C*, 32, 2361–2366. https://doi.org/10.1016/j.msec.2012.07.008
- Li, J., Yu, X., Tang, W., Wan, C., Lu, Y., Dong, N., et al. (2021). Characterization of food gels prepared from the water extract of fish (Cyprinus carpio L.). scales: From molecular components to sensory attributes. *Food Hydrocolloids*, 112, Article 106263. https://doi.org/10.1016/j.foodhyd.2020.106263
- Lotfi, G., Shokrgozar, M. A., Mofid, R., Abbas, F. M., Ghanavati, F., Baghban, A. A., et al. (2016). Biological evaluation (*in vitro* and *in vivo*) of bilayered collagenous coated (nano electrospun and solid wall) chitosan membrane for periodontal guided bone regeneration. Annals of Biomedical Engineering, 44, 2132–2144. https://doi.org/ 10.1007/s10439-015-1516-z
- Lui, Y. S., Sow, W. T., Tan, L. P., Wu, Y., Lai, Y., & Li, H. (2019). 4D printing and stimuliresponsive materials in biomedical aspects. *Acta Biomaterialia*, 92, 19–36. https:// doi.org/10.1016/j.actbio.2019.05.005
- Mantha, S., Pillai, S., Khayambashi, P., Upadhyay, A., Zhang, Y., Tao, O., Pham, H. M., & Tran, S. D. (2019). Smart hydrogels in tissue engineering and regenerative medicine. *Materials*, 12, 3323. https://doi.org/10.3390/ma12203323
- Mardani, M., Yeganehzad, S., Ptichkina, N., Kodatsky, Y., Kliukina, O., Nepovinnykh, N., et al. (2019). Stud on foaming, rheological and thermal properties of gelatin –free marshmallow. Food Hydrocolloids, 93, 335–341. https://doi.org/10.1016/j. foodhyd.2019.02.033
- Martínez-Alvarez, O., Chamorro, S., & Brenes, A. (2015). Protein hydrolysates from animal processing by-products as a source of bioactive molecules with interest in animal feeding: A review. *Food Research International*, 73, 204–212. https://doi.org/ 10.1016/j.foodres.2015.04.005
- Martínez, A., Blanco, M. D., Davidenko, N., & Cameron, R. E. (2015). Tailoring chitosan/ collagen scaffolds for tissue engineering: Effect of composition and different

A. Irastorza et al.

crosslinking agents on scaffold properties. Carbohydrate Polymers, 132, 606–619. https://doi.org/10.1016/j.carbpol.2015.06.084

- McBane, J. E., Vulesevic, B., Padavan, D. T., McEwan, K. A., Korbutt, G. S., & Suuronen, E. J. (2013). Evaluation of a collagen-chitosan hydrogel for potential use as a pro-angiogenic site for islet transplantation. *PloS One*, 8(10), Article e77538. https://doi.org/10.1371/journal.pone.0077538
- Meena, R. A. A., Banu, J. R., Kannah, R. Y., Yogalakshmi, K. N., & Kumar, G. (2020). Biohythane production from food processing wastes - challenges and perspectives. *Bioresource Technology*, 298, Article 122449. https://doi.org/10.1016/j. biortech.2019.122449
- Merz, B., Capello, C., Leandro, G. C., Moritz, D. E., Monteiro, A. R., & Valencia, G. A. (2020). A novel colorimetric indicator film based on chitosan, polyvinyl alcohol and anthocyanins from jambolan (Syzygium cumini) fruit for monitoring shrimp freshness. International Journal of Biological Macromolecules, 153, 625–632. https:// doi.org/10.1016/j.ijbiomac.2020.03.048
- Miklas, J. W., Dallabrida, S. M., Reis, L. A., Ismail, N., Rupnick, M., & Radisic, M. (2013). QHREDGS enhances tube formation, metabolism and survival of endothelial cells in collagen-chitosan hydrogels. *PloS One*, 8(8), Article e72956. https://doi.org/ 10.1371/journal.pone.0072956
- Mohseni, F., & Goli, S. A. H. (2019). Encapsulation of flaxseed oil in the tertiary conjugate of oxidized tannic acid-gelatin and flaxseed (Linum usitatissimum) mucilage. *International Journal of Biological Macromolecules*, 140, 959–964. https:// doi.org/10.1016/j.ijbiomac.2019.08.197
- Molino, S., Casanova, N. A., Rufián Henares, J.Á., & Fernandez Miyakawa, M. E. (2020). Natural tannin wood extracts as a potential food ingredient in the food industry. *Journal of Agricultural and Food Chemistry*, 68, 2836–2848. https://doi.org/10.1021/ acs.jafc.9b00590
- Moreno, O., Atarés, L., Chiralt, A., Cruz-Romero, M. C., & Kerry, J. (2018). Starch-gelatin antimicrobial packaging materials to extend the shelf life of chicken breast fillets. *Lebensmittel-Wissenschaft & Technologie*, 97, 483–490. https://doi.org/10.1016/j. lwt.2018.07.005
- Nasrollahzadeh, M., Sajjadi, M., Iravani, S., & Varma, R. S. (2021). Starch, cellulose, pectin, gum, alginate, chitin and chitosan derived (nano) materials for sustainable water treatment: A review. *Carbohydrate Polymers*, 251, Article 116986. https://doi. org/10.1016/j.carbpol.2020.116986
- Nguyen, T. T., Heimann, K., & Zhang, W. (2020). Protein recovery from underutilised marine bioresources for product development with nutraceutical and pharmaceutical bioactivities. *Marine Drugs*, 18, 391. https://doi.org/10.3390/ md18080391
- Nilsuwan, K., Guerrero, P., de la Caba, K., Benjakul, S., & Prodpran, T. (2020). Properties and application of bilayer films based on poly (lactic acid) and fish gelatin containing epigallocatechin gallate fabricated by thermo-compression molding. *Food Hydrocolloids*, 105, Article 105792. https://doi.org/10.1016/j. foodhvd.2020.105792
- Oertzen-Hagemann, V., Kirmse, M., Eggers, B., Pfeiffer, K., Marcus, K., de Marées, M., et al. (2019). Effects of 12 Weeks of hypertrophy resistance exercise training combined with collagen peptide supplementation on the skeletal muscle proteome in recreationally active men. *Nutrients*, 11, 1072. https://doi.org/10.3390/ nu11051072
- Oliveira, V. D. M., Assis, C. R. D., Costa, B. D. A. M., Neri, R. C. D. A., Monte, F. T., Freitas, H. M. S. D. C. V., et al. (2021). Physical, biochemical, densitometric and spectroscopic techniques for characterization collagen from alternative sources: A review based on the sustainable valorization of aquatic by-products. *Journal of Molecular Structure*, 1224, Article 129023. https://doi.org/10.1016/j. molstruc.2020.129023
- Ong, K. L., Kaur, G., Pensupa, N., Uisan, K., & Lin, C. S. K. (2018). Trends in food waste valorization for the production of chemicals, materials and fuels: Case study South and Southeast Asia. *Bioresource Technology*, 248, 100–112. https://doi.org/10.1016/ j.biortech.2017.06.076
- Ozaki, M. M., Munekata, P. E. S., Lopes, A. S., do Nascimento, M. S., Pateiro, M., Lorenzo, J. M., et al. (2020). Using chitosan and radish powder to improve stability of fermented cooked sausages. *Meat Science*, 167, Article 108165. https://doi.org/ 10.1016/j.meatsci.2020.108165
- O'Brien, F. J. (2011). Biomaterials & scaffolds for tissue engineering. *Materials Today, 14*, 88–95. https://doi.org/10.1016/S1369-7021(11)70058-X
- Pal, G. K., & Suresh, P. V. (2016). Sustainable valorisation of seafood by-products: Recovery of collagen and development of collagen-based novel functional food ingredients. *Innovative Food Science & Emerging Technologies*, 37, 201–215. https:// doi.org/10.1016/j.ifset.2016.03.015
- Pan, M., Liu, K., Yang, J., Liu, S., Wang, S., & Wang, S. (2020). Advances on food-derived peptidic antioxidants- a review. *Antioxidants*, 9, 700. https://doi.org/10.3390/ antiox9090799
- Paula, D. D. A., Martins, E. M. F., Costa, N. D. A., de Oliveira, P. M., de Oliveira, E. B., & Ramos, A. M. (2019). Use of gelatin and gum Arabic for microencapsulation of probiotic cells from Lactobacillus plantarum by a dual process combining double emulsification followed by complex coacervation. *International Journal of Biological Macromolecules*, 133, 722–731. https://doi.org/10.1016/j.ijbiomac.2019.04.110
- Pawelec, K. M., Best, S. M., & Cameron, R. E. (2016). Collagen: A network for regenerative medicine. *Journal of Materials Chemistry B*, 4, 6484–6496. https://doi. org/10.1039/c6tb00807k
- Peanparkdee, M., Yamauchi, R., & Iwamoto, S. (2018). Encapsulation of structured lipids containing medium- and long chain fatty acids by complex coacervation of gelatin and gum Arabic. *Journal of Food Process Engineering*, 41, Article e1290. https://doi. org/10.1111/jfpe.12907
- Pellá, M. C. G., Silva, O. A., Pellá, M. G., Beneton, A. G., Caetano, J., Simões, M. R., et al. (2020). Effect of gelatin and casein additions on starch edible biodegradable films

for fruit surface coating. *Food Chemistry*, 309, Article 125764. https://doi.org/ 10.1016/j.foodchem.2019.125764

- Perez-Puyana, V., Jiménez-Rosado, M., Romero, A., & Guerrero, A. (2019). Crosslinking of hybrid scaffolds produced from collagen and chitosan. *International Journal of Biological Macromolecules*, 139, 262–269. https://doi.org/10.1016/j. ijbiomac.2019.07.198
- Pezeshki-Modaress, M., Zandi, M., & Rajabi, S. (2018). Tailoring the gelatin/chitosan electrospun scaffold for application in skin tissue engineering: An in vitro study. *Progress in Biomaterials, 7,* 207–218. https://doi.org/10.1007/s40204-018-0094-1
 Piedade, A. P. (2019). 4D printing: The shape-morphing in additive manufacturing.
- Journal of Functional Biomaterials, 10(1), 9. https://doi.org/10.3390/jfb10010000
- Qin, Y., Liu, Y., Yuan, L., Yong, H., & Liu, J. (2019). Preparation and characterization of antioxidant, antimicrobial and pH-sensitive films based on chitosan, silver nanoparticles and purple corn extract. *Food Hydrocolloids*, 96, 102–111. https://doi. org/10.1016/j.foodhyd.2019.05.017

Rasente, R. Y., Imperiale, J. C., Lázaro-Martínez, J. M., Gualco, L., Oberkersch, R., Sosnik, A., et al. (2016). Dermatan sulfate/chitosan polyelectrolyte complex with potential application in the treatment and diagnosis of vascular disease. *Carbohydrate Polymers*, 144, 362–370. https://doi.org/10.1016/j. carbopol.2016.02.046

- Reddy, N., Reddy, R., & Jiang, Q. (2015). Crosslinking biopolymers for biomedical applications. *Trends in Biotechnology*, 33(6), 362–369. https://doi.org/10.1016/j. tibtech.2015.03.008
- Regubalan, B., Pandit, P., Maiti, S., Nadathur, G. T., & Mallick, A. (2018). Potential biobased edible films, foams, and hydrogels for food packaging. In S. Ahmed (Ed.), *Biobased materials for food packaging*. Singapore: Springer. https://doi.org/10.1007/ 978-981-13-1909-9 5.
- Reyna-Urrutia, V. A., Mata-Haro, V., Cauich-Rodriguez, J. V., Herrera-Kao, W. A., & Cervantes-Uc, J. M. (2019). Effect of two crosslinking methods on the physicochemical and biological properties of the collagen-chitosan scaffolds. *European Polymer Journal*, 117, 424–433. https://doi.org/10.1016/j. eurpolymj.2019.05.010
- Rodrigues, M.Á. V., Bertolo, M. R. V., Marangon, C. A., Martins, V. D. C. A., & Plepis, A. M. D. G. (2020). Chitosan and gelatin materials incorporated with phenolic extracts of grape seed and jabuticaba peel: Rheological, physicochemical, antioxidant, antimicrobial and barrier properties. *International Journal of Biological Macromolecules*, 160, 769–779. https://doi.org/10.1016/j.ijbiomac.2020.05.240
- Sahraee, S., Milani, J. M., Regenstein, J. M., & Kafil, H. S. (2019). Protection of foods against oxidative deterioration using edible films and coatings: A review. Food Bioscience, 32, Article 100451. https://doi.org/10.1016/j.fbio.2019.100451
- Santos, V. P., Marques, N. S. S., Maia, P. C. S. V., de Lima, M. A. B., Franco, L. O., & de Campos-Takaki, G. M. (2020). Seafood waste as attractive source of chitin and chitosan production and their applications. *International Journal of Molecular Sciences*, 21, 4290. https://doi.org/10.3390/ijms21124290
- Sarkar, S. D., Farrugia, B. L., Dargaville, T. R., & Dhara, S. (2013). Chitosan-collagen scaffolds with nano/microfibrous architecture for skin tissue engineering. *Journal of Biomedical Materials Research Part A*, 101(12), 3482–3492. https://doi.org/10.1002/ ibm.a.34660
- Sato, K. (2017). The presence of food-derived collagen peptides in human boy-structure and biological activity. *Food & Function*, 8, 12. https://doi.org/10.1039/ C7FO01275F
- Senadheera, T. R. L., Dave, D., & Shahidi, F. (2020). Sea cucumber derived type I collagen: A comprehensive review. *Marine Drugs*, 18, 471. https://doi.org/10.3390/ md18090471
- Shah, R., Stodulka, P., Skopalova, K., & Saha, P. (2019). Dual crosslinked collagen/ chitosan film for potential biomedical applications. *Polymers*, 11(12), 2094. https:// doi.org/10.3390/polym11122094
- Sharma, S., & Tiwari, S. (2020). A review on biomacromolecular hydrogel classification and its applications. *International Journal of Biological Macromolecules*, 162, 737–747. https://doi.org/10.1016/j.ijbiomac.2020.06.110
- Shie, M. Y., Shen, Y. F., Astuti, S. D., Kai-Xing Lee, A., Lin, S. H., Dwijaksara, N. L. B., et al. (2019). Review of polymeric materials in 4D printing biomedical applications. *Polymers*, 11(11), 1864. https://doi.org/10.3390/polym11111864
- Shin, C. S., Kim, D. Y., & Shin, W. S. (2019). Characterization of chitosan extracted from Mealworm Beetle (*Tenebrio molitor, Zophobas morio*) and Rhinoceros Beetle (*Allomyrina dichotoma*) and their antibacterial activities. *International Journal of Biological Macromolecules*, 125, 72–77. https://doi.org/10.1016/j. iibiomac.2018.11.242
- Shokri, S., Parastouei, K., Taghdir, M., & Abbaszadeh, S. (2020). Application an edible active coating based on chitosan- *Ferulago angulata* essential oil nanoemulsion to shelf life extension of Rainbow trout fillets stored at 4 °C. *International Journal of Biological Macromolecules*, 153, 846–854. https://doi.org/10.1016/j. ijbiomac.2020.03.080
- Sionkowska, A., Kaczmarek, B., & Lewandowska, K. (2014). Modification of collagen and chitosan mixtures by the addition of tannic acid. *Journal of Molecular Liquids, 199*, 318–323. https://doi.org/10.1016/j.molliq.2014.09.028
- Skov, K., Oxfeldt, M., Thøgersen, R., Hansen, M., & Bertram, H. C. (2019). Enzymatic hydrolysis of a collagen hydrolysate enhances postprandial absorption rate-A randomized controlled trial. *Nutrients*, 11, 1064. https://doi.org/10.3390/ nu11051064
- Souza, V. G. L., Pires, J. R. A., Vieira, E. T., Coelhoso, I. M., Duarte, M. P., & Fernando, A. L. (2019). Activity of chitosan-montmorillonite bionanocomposites incorporated with rosemary essential oil: From *in vitro* assays to application in fresh poultry meat. *Food Hydrocolloids*, 89, 241–252. https://doi.org/10.1016/j. foodhyd.2018.10.049

A. Irastorza et al.

- Subhan, F., Ikram, M., Shehzad, A., & Ghafoor, A. (2015). Marine collagen: An emerging player in biomedical applications. *Journal of Food Science & Technology*, 52(8), 4703–4707. https://doi.org/10.1007/s13197-014-1652-8
- Sullivan, D. J., Cruz-Romero, M. C., Hernandez, A. B., Cummins, E., Kerry, J. P., & Morris, M. A. (2020). A novel method to deliver natural antimicrobial coating materials to extend the shelf-life of European hake (*Merluccius merluccius*) fillets. *Food Packaging and Shelf Life*, 25, Article 100522. https://doi.org/10.1016/j. fpsl.2020.100522
- Sun, Y., Yang, C., Zhu, X., Wang, J. J., Liu, X. Y., Yang, X. P., et al. (2019). 3D printing collagen/chitosan scaffold ameliorated axon regeneration and neurological recovery after spinal cord injury. *Journal of Biomedical Materials Research Part A*, 107(9), 1898–1908. https://doi.org/10.1002/jbm.a.36675
- Suresh, K., Sugihara, F., Suzuki, K., Inoue, N., & Venkateswarathirukumara, S. (2015). A double-blind, placebo-controlled, randomized, clinical study on the effectiveness of collagen peptide on osteoarthritis. *Journal of the Science of Food and Agriculture*, 94 (4), 702–707. https://doi.org/10.1002/jsfa.6752
- Talón, E., Trifkovic, K. T., Nedovic, V. A., Bugarski, B. M., Vargas, M., Chiralt, A., et al. (2017). Antioxidant edible films based on chitosan and starch containing polyphenols from thyme extracts. *Carbohydrate Polymers*, 157, 1153–1161. https:// doi.org/10.1016/j.carbpol.2016.10.080
- Tamay, D. G., Usal, T. D., Alagoz, A. S., Yucel, D., Hasirci, N., & Hasirci, V. (2019). 3D and 4D printing of polymers for tissue engineering applications. *Frontiers in Bioengineering and Biotechnology*, 7, 164. https://doi.org/10.3389/fbioe.2019.00164
- Tondorf, R., Aibibu, D., & Cherif, C. (2020). Collagen multifilament spinning. *Materials Science and Engineering: C, 106*, Article 110105. https://doi.org/10.1016/j. msec 2019 110105
- Tosati, J. V., Messias, V. C., Carvalho, P. I. N., Pollonio, M. A. R., Meireles, M. A. A., & Monteiro, A. R. (2017). Antimicrobial effect of edible coating blend based on turmeric starch residue and gelatin applied onto fresh frankfurter sausage. *Food and Bioprocess Technology*, 10, 2165–2175. https://doi.org/10.1007/s11947-017-1985-1
- United Nations. (2015). A/RES/70/1 transforming our world: The 2030 agenda for sustainable development. https://www.un.org/ga/search/view_doc.asp?symbol=A/ RES/70/1&Lang=E. (Accessed 6 December 2020).
- Uranga, J., Nguyen, B. T., Si, T. T., Guerrero, P., & De la Caba, K. (2020). The effect of cross-linking with citric acid on the properties of agar/fish gelatin films. *Polymers*, 12, 291. https://doi.org/10.3390/polym12020291
- Uranga, J., Puertas, A. I., Etxabide, A., Dueñas, M. T., Guerrero, P., & de la Caba, K. (2019). Citric acid-incorporated fish gelatin/chitosan composite films. *Food Hydrocolloids*, 86, 95–103. https://doi.org/10.1016/j.foodhyd.2018.02.018
- Wang, L., Jiang, Y., Wang, X., Cui, H., Xu, W., He, Y., et al. (2018). Effect of oral administration of collagen hydrolysates from Nile tilapia on the chronologically aged skin. *Journal of Functional Foods*, 44, 112–117. https://doi.org/10.1016/j. iff.2018.03.005
- Wang, P., Li, Y., Zhang, C., Que, F., Weiss, J., & Zhang, H. (2020). Characterization and antioxidant activity of trilayer gelatin/dextran-propyl gallate/gelatin films: Electrospinning versus solvent casting. *Lebensmittel-Wissenschaft & Technologie, 128*, Article 109536. https://doi.org/10.1016/j.lwt.2020.109536
- Wang, X., Wang, G., Liu, L., & Zhang, D. (2016). The mechanism of a chitosan-collagen composite film used as biomaterial support for MC3T3-E1 cell differentiation. *Scientific Reports*, 6(1), 1–8. https://doi.org/10.1038/srep39322
- Whu, S. W., Hung, K. H., Hsieh, K. H., Chen, C. H., Tsai, C. L., & Hsu, S. H. (2013). In vitro and in vivo evaluation of chitosan-gelatin scaffolds for cartilage tissue engineering. *Materials Science and Engineering: C, 33*(5), 2855–2863. https://doi.org/ 10.1016/j.msec.2013.03.003
- Wu, X., Luo, Y., Liu, Q., Jiang, S., & Mu, G. (2019). Improved structure-stability and packaging characters of crosslinked collagen fiber-based film with casein, keratin and SPI. Journal of the Science of Food and Agriculture, 99, 4942–4951. https://doi. org/10.1002/jisfa.9726
- Wu, C., Sun, J., Zheng, P., Kang, X., Chen, M., Li, Y., et al. (2019). Preparation of an intelligent film based on chitosan/oxidized chitin nanocrystals incorporating black rice bran anthocyanins for seafood spoilage monitoring. *Carbohydrate Polymers*, 222, Article 115006. https://doi.org/10.1016/j.carbpol.2019.115006

- Xiao, W., Hu, X. Y., Zeng, W., Huang, J. H., Zhang, Y. G., & Luo, Z. J. (2013). Rapid sciatic nerve regeneration of rats by a surface modified collagen-chitosan scaffold. *Injury*, 44(7), 941–946. https://doi.org/10.1016/j.injury.2013.03.029
- Xiong, X., Yu, I. K. M., Tsang, D. C. W., Bolan, N. S., Ok, Y. S., Igalavithana, A. D., et al. (2019). Value-added chemicals from food supply chain wastes: State-of-the-art review and future prospects. *Chemical Engineering Journal*, 375, Article 121983. https://doi.org/10.1016/j.cej.2019.121983
- Yadav, S., Mehrotra, G. K., & Dutta, P. K. (2021). Chitosan based ZnO nanoparticles loaded gallic-acid films for active food packaging. *Food Chemistry*, 334, Article 127605. https://doi.org/10.1016/j.foodchem.2020.127605
- Yang, X., Li, A., Li, X., Sun, L., & Guo, y. (2020). An overview of classifications, properties of food polysaccharides and their links to applications in improving food textures. *Trends in Food Science & Technology*, 102, 1–15. https://doi.org/10.1016/j. tifs.2020.05.020
- Yan, L. P., Wang, Y. J., Ren, L., Wu, G., Caridade, S. G., Fan, J. B., et al. (2010). Genipincross-linked collagen/chitosan biomimetic scaffolds for articular cartilage tissue engineering applications. *Journal of Biomedical Materials Research Part A*, 95A(2), 465–475. https://doi.org/10.1002/jbm.a.32869
- Yeddes, W., Djebali, K., Aidi Wannes, W., HorchaniNaifer, K., Hammami, M., Younes, I., et al. (2020). Gelatin-chitosan-pectin films incorporated with rosemary essential oil: Optimized formulation using mixture design and response surface methodology. *International Journal of Biological Macromolecules*, 154, 92–103. https://doi.org/ 10.1016/j.ijbiomac.2020.03.092
- Yong, H., Wang, X., Zhang, X., Liu, Y., Qin, Y., & Liu, J. (2019). Effects of anthocyaninrich purple and black eggplant extracts on the physical, antioxidant and pH-sensitive properties of chitosan film. *Food Hydrocolloids*, 94, 93–104. https://doi.org/ 10.1016/j.jfoodhyd.2019.03.012
- Yousefi, M., Ariffin, F., & Huda, N. (2017). An alternative source of type I collagen based on by-product with higher thermal stability. *Food Hydrocolloids*, 63, 372–382. https://doi.org/10.1016/j.foodhyd.2016.09.029
- Yu, D., Jiang, Q., Xu, Y., & Xia, W. (2017). The shelf life extension of refrigerated grass carp (*Ctenopharyngodon idellus*) fillets by chitosan coating combined with glycerol monolaurate. *International Journal of Biological Macromolecules*, 101, 448–454. https://doi.org/10.1016/j.ijbiomac.2017.03.038
- Zamorano-Apodaca, J. C., García-Sifuentes, C. O., Carvajal-Millán, E., Vallejo-Galland, B., Scheuren-Acevedo, S. M., & Lugo-Sánchez, M. E. (2020). Biological and functional properties of peptide fractions obtained from collagen hydrolysate derived from mixed by-product of different fish species. *Food Chemistry*, 331, Article 127350. https://doi.org/10.1016/j.foodchem.2020.127350
- Zarandona, I., López-Caballero, M. E., Montero, M. P., Guerrero, P., de la Caba, K., & Gómez-Guillén, M. C. (2021). Horse mackerel (*Trachurus trachurus*) fillets biopreservation by using gallic acid and chitosan coatings. *Food Control*, 120, Article 107511. https://doi.org/10.1016/j.foodcont.2020.107511
- Zhang, H., Liang, Y., Li, X., & Kang, H. (2020). Effect of chitosan-gelatin coating containing nano-encapsulated tarragon essential oil on the preservation of pork slices. *Meat Science*, 166, Article 108137. https://doi.org/10.1016/j. meatsci.2020.108137
- Zhang, W., Li, X., & Jiang, W. (2020). Development of antioxidant chitosan film with banana peels extract and its application as coating in maintaining the storage quality of apple. *International Journal of Biological Macromolecules*, 154, 1205–1214. https:// doi.org/10.1016/j.ijbiomac.2019.10.275
- Zhang, X., Liu, J., Yong, H., Qin, Y., Liu, J., & Jin, C. (2020). Development of antioxidant and antimicrobial packaging films based on chitosan and mangosteen (*Garcinia* mangostana L.) rind powder. International Journal of Biological Macromolecules, 145, 1129–1139. https://doi.org/10.1016/j.ijbiomac.2019.10.038
- Zhang, H., Zhang, F., & Wu, J. (2013). Physically crosslinked hydrogels from polysaccharides prepared by freeze-thaw technique. *Reactive and Functional Polymers*, 73(7), 923–928. https://doi.org/10.1016/j.reactfunctpolym.2012.12.014
- Zhuang, C., Tao, F., & Cui, Y. (2017). Eco-friendly biorefractory films of gelatin and TEMPO-oxidized cellulose ester for food packaging applications. *Journal of the Science of Food and Agriculture*, 97, 3384–3395. https://doi.org/10.1002/jsfa.8189