

Neurolipid systems: A new target for the treatment of dementia

Gorka Pereira-Castelo¹ | Iker Bengoetxea de Tena¹  |
 Jonatan Martínez-Gardeazabal¹ | Marta Moreno-Rodríguez² |
 Estibaliz González de San Román¹ | Iván Manuel^{1,3}  | Rafael Rodríguez-Puertas^{1,3}

¹Department of Pharmacology, University of the Basque Country (UPV/EHU), Leioa, Spain

²Department of Translational Neuroscience, Barrow Neurological Institute, Phoenix, Arizona, USA

³Neurodegenerative Diseases, BioBizkaia Health Research Institute, Barakaldo, Spain

Correspondence

Iván Manuel, Department of Pharmacology, University of the Basque Country (UPV/EHU), Leioa, Spain.
 Email: ivan.manuel@ehu.eus

Funding information

Basque Government, Grant/Award Number: IT1454-22; Instituto de Salud Carlos III, Grant/Award Number: PI20/00153; Eitb Maratoia, Grant/Award Number: BIO22/ALZ/010

[Correction added on 26 July 2024, after first online publication: The 5th author's name has been corrected from "Esibaliz" to "Estibaliz" in this version.]

Abstract

Neurolipids comprise a diverse class of bioactive lipids that include molecules capable of activating G protein-coupled receptors, thereby inducing systemic effects that contribute to the maintenance of homeostasis. Dementia, a non-specific brain disorder characterized by a common set of signs and symptoms, usually arises subsequent to brain injuries or diseases and is often associated with the aging process. Individuals affected by dementia suffer from the disruption of several neurotransmitter and neuromodulatory systems, among which neurolipids play an important role, including the endocannabinoid, lysophosphatidic acid and sphingosine 1-phosphate systems. In this review, we present an overview of the most recent and pertinent findings regarding the involvement of these neurolipidic systems in dementia, including data from a wide range of both in vitro and in vivo experiments as well as clinical trials.

KEYWORDS

dementia, endocannabinoid, lysophosphatidic acid, neurolipid, sphingosine 1-phosphate

Plain English Summary

Lipids, apart from the classical functions attributed to them, also have a function as molecules with neurotransmitter activity; these are called *neurolipids*. *Neurolipids* exert their function through G protein-coupled receptors and are involved in many physiological processes. In this review, we approach possible treatments involving the modulation of neurotransmitter systems by *neurolipids*, focusing on the cannabinoid, lysophosphatidic acid and sphingosine 1-phosphate systems in diseases in which dementia appears, such as Alzheimer's disease, Parkinson's disease and Huntington's disease. Given the lack of effective treatments for these diseases, *neurolipids* emerge as molecules with great therapeutic potential.

1 | INTRODUCTION

Dementia is a nonspecific brain disorder characterized by a set of common signs and symptoms that frequently appears after brain damage caused by injury or disease and is heavily related to aging. Along with neurodegenerative signs and neurological symptoms, such as impairments in progressive learning and memory, other symptoms are usually observed related to difficulties in behavioural, motor or emotional aspects, including language problems and depression. These signs may vary based on the type of dementia and the characteristics of each patient, which generates a large and heterogeneous group of different clinical profiles requiring individualized treatments.¹ Several neurodegenerative disorders are accompanied by different symptoms of dementia, including Alzheimer's disease (AD), Parkinson's disease (PD) and Huntington's disease (HD). Currently, there are few effective pharmacological treatments indicated for these diseases, and they are generally focused on symptomatic relief rather than targeting the neurodegenerative process. Moreover, there is a lack of early diagnostic biomarkers for early detection.

AD, the most common dementia worldwide, is characterized by a progressive cognitive impairment of learning and memory, with fatal consequences for the patient. While the classical histopathological hallmarks of AD include extracellular amyloid- β plaques and neurofibrillary tangles composed of hyperphosphorylated tau protein, synaptic loss is the parameter that best correlates with disease progression.² Neurotransmitter systems

specifically affected in AD include cholinergic pathways and also lipid-based neurotransmitter systems such as the endocannabinoid (eCB).³ Currently, the only approved treatments for the disease are inhibitors of acetylcholinesterase, antagonists of NMDA receptors, and more recently, anti-amyloid- β peptide antibodies; the last one is not exempt from controversy, and all of them show very limited improvements.

PD is principally a motor disorder that often courses with cognitive impairment. Recently, some types of PD have been defined neuropathologically as an alpha synucleinopathy showing accumulation of alpha synuclein protein in dopaminergic neurons of the *substantia nigra* that may lead to neuronal death and progressive impairment of the motor cognition of the patient. PD patients can develop a specific type of dementia, which is characterized by cognitive impairment, visual hallucinations and, more importantly, parkinsonism, defined by bradykinesia, gait impairment and rest tremor, among other symptoms.⁴ As is the case with other neurodegenerative diseases, it has no healing treatment, being L-Dopa the most effective drug to mitigate the motor symptoms.

Unlike AD and PD, HD is a hereditary neurodegenerative disease, being the most common form of genetic-associated dementia. It is caused by a cytosine-adenine-guanine (CAG) trinucleotide repetition in the gene that encodes the huntingtin protein on chromosome 4.⁵ Chorea, characterized by brief, semi-directed and irregular movements, is the most characteristic symptom of HD, and the treatment lies in antipsychotics and benzodiazepines, which do not reverse nor protect the patient from neurodegeneration.⁶

Therefore, new and innovative treatments are necessary for these three neurodegenerative diseases.

2 | NEUROLIPIDS AND THEIR ROLE IN THE MOST COMMON DEMENTIAS

Lipids are widely recognized for their roles in providing structure, energy storage and thermoregulation to the organism. However, it is important to broaden our perspective on lipids, as some of them also possess signalling capabilities in the CNS through binding and activation to G protein-coupled receptors (GPCRs).⁷ There is a large variety of lipids in the CNS classified according to their chemical structures, such as sphingolipids, phospholipids, cholesterol and glycosphingolipids.⁸ Few of them can be considered integral members of the extensive family of neurotransmitters and/or neuromodulators. Since peptides with these properties are called 'neuropeptides', lipids could be named as 'neurolipids'.⁹ The discovery of

new lipid-based neurotransmitter systems, such as eCBs, lysophosphatidic acid (LPA) and sphingosine 1-phosphate (S1P), shifts the focus to the relevance of these systems in both neurophysiological status and neurological disorders.

Neurolipids are typically synthesized in a calcium-dependent manner, as needed. This synthesis occurs through the enzymatic processing from membrane lipid precursors by phospholipases, sphingomyelinases or cytochrome P450 hydroxylases.¹⁰ Precursors are located mainly at the cell membranes of the pre- or post-synaptic compartments. Endogenous ligands for neurolipids have the capability to interact with their respective receptors either in an autocrine manner, where they directly engage with their specific receptor within the lipid bilayer, or in a paracrine way. Their amphipathic properties allow them to easily cross the lipid bilayer, allowing them to function also in the cytoplasm¹¹ (Figure 1). Regarding their physiological roles, neurolipids have been involved in multiple processes in both peripheral and central nervous systems (CNSs), as well as regulating immune responses. These neurolipid systems are also regulated during the abovementioned neurodegenerative diseases. Thus, receptors and enzymes involved in the synthesis and degradation of the endogenous neurotransmitters of these systems could be used not only as biomarkers for early diagnosis of the mentioned diseases but also as therapeutic targets.¹²

Therefore, we have reviewed three of the most studied lipid-based or neurolipid neurotransmitter systems, namely, eCB, LPA and S1P systems, and their roles in the context of dementia linked to AD, PD and HD (Table 1, Figure 2).

2.1 | Endocannabinoid system

The eCB system has a widespread distribution in the CNS, modulating multiple neural processes, including neurodevelopment, synaptic plasticity and adaptive responses.³⁹ The main eCB neurotransmitters are N-arachidonylethanolamine (anandamide or AEA) and 2-arachidonoylglycerol (2-AG), which activate two different subtypes of G_{i/o}-proteins-coupled cannabinoid receptors, CB₁ and CB₂. Some of the enzymes involved in the synthesis of the eCB include N-acylphosphatidylethanolamine-specific phospholipase D (NAPE-PLD) for AEA and diacylglycerol lipase (DAGL) for 2-AG, and for degradation, these include fatty acid amide hydrolase-1 (FAAH) for AEA and monoacylglycerol lipase (MAGL) for 2-AG.⁴⁰

The eCB system is involved in different neurodevelopmental, psychiatric and neurodegenerative disorders. Thus, cannabimimetic and cannabinoid-based drugs have been proposed for the treatment of anxiety, schizophrenia, epilepsy and many neurological diseases, including some genetic syndromes or others accompanied by dementia, such as AD, PD and HD.

The activity of CB₁ receptors is higher in brain regions such as the hippocampus and the frontal cortex, which are involved in learning and memory processes at initial phases of the AD, gradually decreasing in advanced stages.⁴¹ In frontal cortex, the same tendency was observed using the [¹²⁵I]SD-7015 radioligand to measure CB₁ receptor density and, importantly, this parameter inversely correlates with Braak's tau-based classification.⁴² A study performed using brain samples from late stages of AD (Braak V/VI) analysed both CB

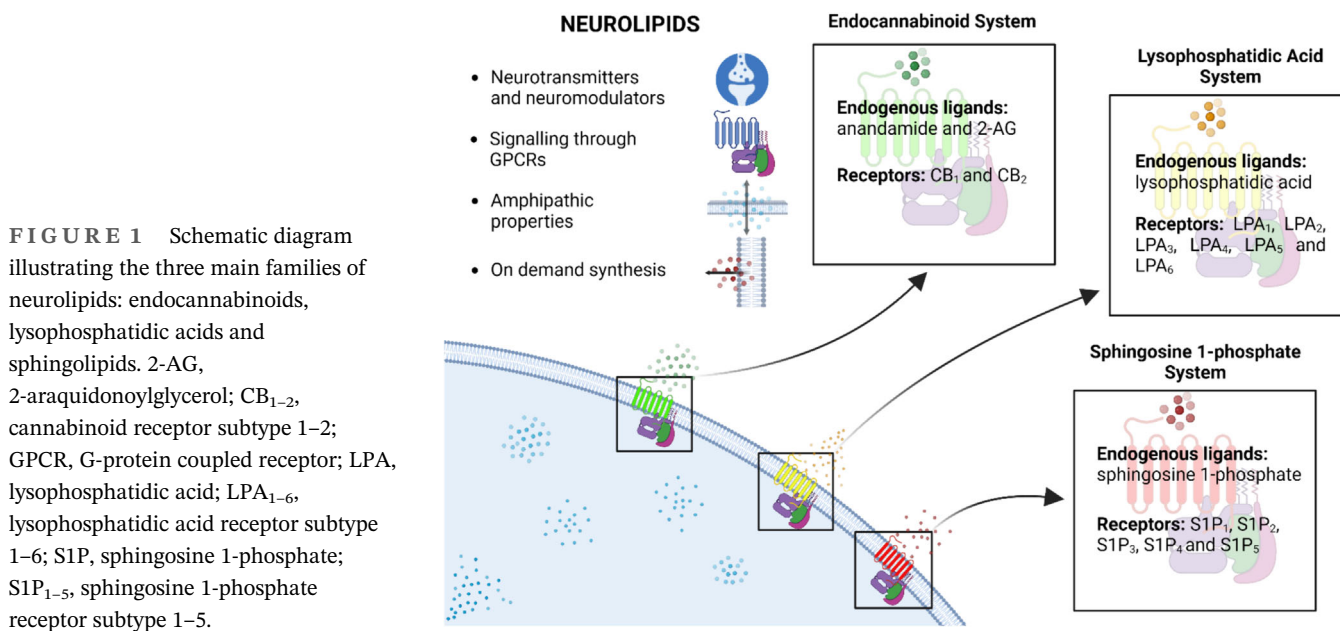


FIGURE 1 Schematic diagram illustrating the three main families of neurolipids: endocannabinoids, lysophosphatidic acids and sphingolipids. 2-AG, 2-arachidonoylglycerol; CB₁₋₂, cannabinoid receptor subtype 1–2; GPCR, G-protein coupled receptor; LPA, lysophosphatidic acid; LPA₁₋₆, lysophosphatidic acid receptor subtype 1–6; S1P, sphingosine 1-phosphate; S1P₁₋₅, sphingosine 1-phosphate receptor subtype 1–5.

TABLE 1 Novel neuropilidic treatments for Alzheimer's, Parkinson's and Huntington's diseases in different preclinical models and clinical trials.

Target disease	Study type	Treatment	Method	Findings	References
Alzheimer's disease	Patients	Cannabinoid extract	Mnemonic test	Improved mnemonic ability	13
	Experimental models	Fingolimod; SPHK1	MWM	Improved cognitive performance	14–16
		Botanical extracts (THC + CBD)	AAT, ELISA, immunohistochemistry, PCR, TORT and WB	Improved cognitive performance and decreased astrogliosis, microgliosis and neuroinflammation	17
		WIN55,212-2; JZL184	Autoradiographic studies and Immunohistochemistry	Increased CB ₁ activity and density	18
	Gintonin	ELISA, immunohistochemistry, MWM, WB and YM	Improved cognitive performance and reduced β -amyloid levels	19	
Parkinson's disease	Patients	CBD	Randomized, double-blinded, placebo-controlled, crossover clinical trial	Reduced anxiety and tremor amplitude	20
		Nabilone	Phase II placebo-controlled, double-blind, parallel-group, enriched enrolment randomized withdrawal trial	Improvements in anxiety and sleeping problems	21
	Experimental models	Ponesimod	Immunohistochemistry, WB and YM	Spatial memory improvement, reduced TNF- α and CXCL10 levels and increased IL-33 levels	22
		S1P	Immunohistochemistry, PCR and WB	Increased SPHK1 levels and cell viability restoration	23
		Fingolimod	Actimeter, beam test, immunohistochemistry, pole test, rotarod, RT-PCR and WB	Motor improvement and increased BDNF levels.	24–26
		2-AG	Immunohistochemistry and HPLC	Neuroprotection	27
		URB597	Immunohistochemistry and rotarod	Motor improvement and increased anandamide levels	28
		Δ 9-tetrahydrocannabivarin	CAA	Motor improvement	29
		VCE-004.8	CRT, immunohistochemistry and pole test	Increased dopaminergic neuron survival	30
Gintonin	PCR, pole test, immunohistochemistry, rotarod and WB	Increased dopaminergic neuron survival and cognitive performance. Decreased inflammation	31		

TABLE 1 (Continued)

Target disease	Study type	Treatment	Method	Findings	References
Huntington's disease	Patients	Sativex (THC + CBD)	Cohort study	No effect	32
	Experimental models	A-971432	Horizontal ladder task and rotarod	Motor improvement and increased lifespan	33
		K6PC-5	Clasping analysis, horizontal ladder task, PCR and rotarod	Motor improvement and increased S1P, S1P ₁ , S1P ₅ and BDNF levels	34
		THI	Clasping analysis, horizontal ladder task, LC-MS/MS, negative geotaxic assay, rotarod, TEM and WB	Motor improvement, activation of pro-survival pathways, normalization of brain-derived neurotrophic factor levels, preservation of white matter integrity and stimulation synaptic functions	35
		CBD; CB ₂ agonists	Immunohistochemistry, in situ hybridation, HPLC and PCR	Neuroprotection	36,37
Gintonin	Immunohistochemistry and RT-PCR	Neuroprotection	38		

Note: The articles included were selected on the basis of their scientific impact, novelty and relevance.

Abbreviations: 2-AG, 2-arachidonoylglycerol; AAT, active avoidance test; AD, Alzheimer's disease; BDNF, brain derived neurotrophic factor; CAA, computer aided acidimetry; CBD, cannabidiol; CRT, cylinder rearing test; ELISA, enzyme-linked immunosorbent assay; HPLC, high performance liquid chromatography; MWM, Morris water maze; PCR, polymerase chain reaction; PD, Parkinson's disease; S1P, sphingosine 1-phosphate; SPHK1, sphingosine kinase 1; TEM, transmission electron microscopy; THC, tetrahydrocannabinol; THI, 2-acetyl-5-tetrahydroxybutyl imidazole; TORT, two object recognition test; WB, western blot; YM, Y maze.

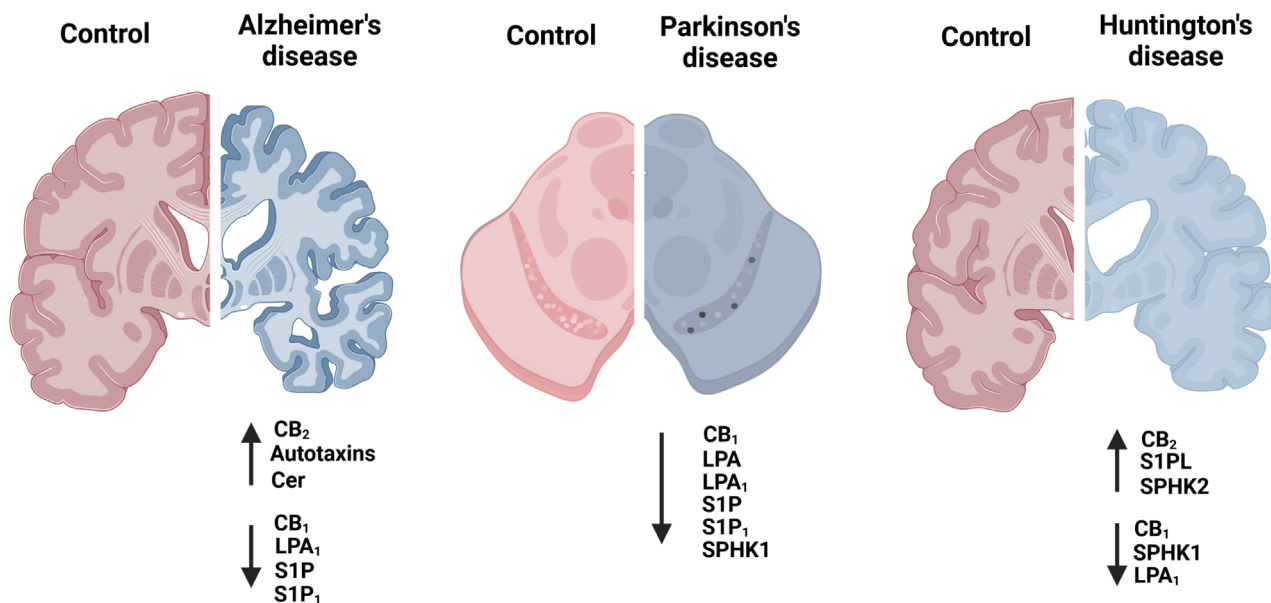


FIGURE 2 A schematic display of the major changes of the sphingosine 1-phosphate, endocannabinoid and lysophosphatidic acid systems in three of the most prominent neurodegenerative diseases, Alzheimer's disease, Parkinson's disease and Huntington's disease. CB₁, cannabinoid receptor subtype 1; CB₂, cannabinoid receptor subtype 2; Cer, ceramide; LPA, lysophosphatidic acid; LPA₁, lysophosphatidic acid receptor subtype 1; S1P, sphingosine 1-phosphate; S1P₁, sphingosine 1-phosphate receptor subtype 1; S1PL, sphingosine 1-phosphate lyase; SPHK, sphingosine kinase.

receptor subtypes using immunoblotting and reported a decrease in CB₁ receptor, in line with previous studies, while CB₂ density augmented probably as a consequence of inflammatory processes.⁴³

Studies using a rat lesion model of basal forebrain cholinergic neurons (BFCNs) degeneration, which are selectively deleted in AD patients,⁴⁴ show increased activity of CB₁ receptors in cortical projection areas.^{45,46} Similar alterations were also found in genetic AD models, such as the APP/PS1 double transgenic mice, in which CB₁ and CB₂ genes expression are altered in presymptomatic stages in the hippocampus and prefrontal cortex.⁴⁷ Importantly, a chronic administration of botanical extracts containing both Δ -9-tetrahydrocannabinol (THC) and cannabidiol (CBD) in early disease stages of these same transgenic mice, reduced astrogliosis, microgliosis and neuroinflammation-related molecules, as well as improved cognitive performance.¹⁷ In a subsequent study using older APP/PS1 mice, reduced memory impairment was equally observed following the treatment, but unlike mice treated at prodromal stages, there was no modulation of glial reactivity nor inflammatory markers. In contrast, cannabinoids altered the cortical excitatory/inhibitory balance only in aged mice.⁴⁸ In other transgenic models, like 5xFAD mice, CB₁ expression in the hippocampus was reduced, and this decrease was accompanied by cognitive impairment.⁴⁹ More recently, in a 5xFAD/CB₂^{EGFP/f/f} mice model, microglial CB₁ receptor increase has been described.⁵⁰ In the 3xTg-AD model, alterations in CB₁ density and activity have been described. In line with the findings obtained from human AD samples, 4-month-old 3xTg-AD mice (prodromal stages) show upregulated CB₁ activity in areas such as the thalamus, which is decreased in areas such as the basal forebrain in older mice (15-month-old, advanced stages).⁵¹ Seven-month-old mice (moderate pathological stage) express increased CB₁ receptor density and activity in various brain areas, particularly in the basolateral amygdala. This increase was reversed following the sub-chronic stimulation of the eCB system with either CB₁/CB₂ agonist WIN55,212-2 or JZL184, an inhibitor of MAGL, the main enzyme that catabolizes 2-AG.¹⁸

Together, these results indicate that CB₁ density and activity in AD are stage-dependent, up-regulated in the early stages of the disease and progressively reduced during neurodegeneration. This biphasic modulation raises controversy about the benefits and potential risks of CB treatments in the early *versus* late stages of the disease. Importantly, a case report published in 2022 described mnemonic improvement following a 22-month-long administration of cannabinoid extract microdoses to a 75-year-old patient presenting memory deficit, spatial and temporal disorientation, and impaired daily

activity.¹³ This report further highlights those biphasic effects of cannabinoid agonism on cognition and the beneficial effects of low as opposed to high doses, as well as the importance of the previous cognitive status of the patient receiving the drug.

In PD, receptors and enzymes, as well as endogenous ligands of the eCB system, are also altered⁵² (Figure 2). Preclinical studies indicate that a modulation of this system exerts neuroprotective actions. A study using the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) neurotoxin mouse model reported neuroprotective effects of increasing 2-AG levels.²⁷ Additionally, URB597, a FAAH inhibitor, effectively prevented motor impairment induced by MPTP. This protective effect was mediated through CB₁ and CB₂ signalling pathways by increasing anandamide levels. However, it did not maintain dopamine levels in the nigrostriatal pathway.²⁸ Δ 9-tetrahydrocannabivarin, a non-psychoactive homologue of THC, demonstrates a dual mechanism of action in PD; it exerts symptoms of palliation by activating CB₂ receptors while simultaneously blocking CB₁ receptors in the 6-hydroxydopamine (6-OHDA) PD model. Additionally, it provides neuroprotection through antioxidant effects.²⁹ More recently, in the PD rat model of 6-OHDA, a CBD derivative, VCE-004.8, an agonist of the CB₂ receptor, was able to attenuate the toxin-induced loss of dopaminergic neurons in the *substantia nigra*.³⁰ Several clinical trials involving various cannabinoid-based drugs have been conducted, resulting in diverse outcomes. Some of these drugs exhibit motor effects, while others focus on neuropsychiatric improvement. The variability in results appears to depend on both the specific drug used and, more significantly, the methodology employed to assess these outcomes.⁵³ Some of the latest clinical trials include a randomized, double-blinded, placebo-controlled, crossover clinical trial, which concluded that CBD administration is associated with reduced anxiety and tremor amplitude in PD patients,⁵⁴ as well as a phase II placebo-controlled, double-blind, parallel-group, enriched enrolment randomized withdrawal trial conducted with nabilone (a CB₁ receptor agonist), which demonstrated improvements in anxiety and sleeping problems among PD patients.⁵⁵

In HD, cannabinoid compounds could act as neuroprotective agents, at least in animal models. The mechanisms by which they exert neuroprotection vary depending on the animal model used. A study using a 3-nitropropionic acid (3-NPA) model reported neuroprotection of striatal projection neurons following CBD administration in a CB₁-independent manner, probably based on the antioxidant properties of CBD.⁵⁶ In a subsequent study in which an intra-striatal injection of the mitochondrial complex II inhibitor malonate was

performed, only CB₂ agonists protected striatal projection neurons from toxin-induced death.⁵⁷ In contrast with preclinical evidence, a double-blind study conducted with 24 HD patients indicated that Sativex, which contains both CBD and THC, did not affect disease progression.³²

2.2 | Lysophosphatidic acid system

LPA is a metabolite of phospholipids biosynthesis in the cell membrane. The generic form of LPA (1- or 2-acyl-sn-glycero-3-phosphate) is composed of phosphate, a glycerol and a fatty acid, but there are different LPA species depending on the length of the fatty acid chain, saturation or position.⁵⁸ The actions of LPA are numerous and different. For instance, LPA produced by platelets is essential as a mediator of tissue regeneration.⁵⁹ LPA also allows different types of cellular responses, such as migration,⁶⁰ neurite contraction,²⁰ stimulation or inhibition of adenylyl cyclase,²¹ or prevention of apoptosis.⁶¹

While the biosynthesis of LPA is not fully known, some enzymatic pathways have been described. The most important is the autotaxin enzymatic pathway, the second one, the production by phospholipase from membrane phospholipids, and, finally, the acyltransferase pathway.⁶² LPA acts as a neurotransmitter through six different receptors, LPA₁-LPA₆ (Figure 1), and intracellularly these can be coupled to four different G α subunits (G α_i , G α_q , G α_{12} , G α_s), and, consequently, different signalling routes can be activated.⁶³

The extent of the involvement of LPA in neurodegenerative diseases is not precisely known, but a number of studies have established the relationship in one way or another between this neurolipid system and AD (Figure 2). Evidences show an increased expression of autotaxin, which in addition to being involved in synthesis, has an antioxidant effect in the prefrontal cortex of AD patients.⁶⁴ LPA levels are also found increased in patients suffering mild cognitive impairment (MCI).⁶⁵ In this context, there seems to be an association between various forms of LPA and A β ₁₋₄₂, as LPA upregulates β -secretase activity.⁶⁶ Still, other authors have shown that administration of LPA agonists, such as gintonin, in transgenic models of AD reduces amyloid plaque deposition and improves memory.¹⁹ Similarly, LPAs participate in the upregulation of glycogen synthase kinase-3 (GSK-3), an enzyme involved in tau phosphorylation, another hallmark of AD.⁶⁷ LPA appears to play a critical role in neurogenesis by modulating neuronal progenitors and in this regard, deficits in neurogenesis have been described.⁶⁸ Indeed, exogenous administration of LPA substantially improves memory in animal models, and

this correlates with increased neurogenesis.⁶⁹ Regarding behaviour, LPA₁ receptor depletion causes symptoms in animals that mimic some of the symptoms that appear in AD type dementias, such as anxiety, memory impairment and motor disturbances.⁷⁰ These evidences, although inconclusive, seem to indicate the relationship between LPA and the development of AD and other dementias.

Regarding PD models in experimental animals, 6-OHDA-injured rats showed a significant reduction of LPA₁ density in the *substantia nigra* and differentiation of mesenchymal stem cells (MSCs) into dopamine neurons following LPA treatment, suggesting that LPA/LPA₁ signalling holds potential importance in the development and maintenance of dopaminergic neurons.⁷¹ Furthermore, the oral administration of gintonin prevented the loss of dopaminergic cells in a PD model induced by MPTP injection. This treatment also exhibited anti-inflammatory and antioxidant properties and supported the preservation of the blood-brain barrier integrity, ultimately enhancing cognitive function. Importantly, these effects of gintonin were substantiated through co-administration with Ki16425, an antagonist targeting LPA_{1/3} receptors, which completely nullified the benefits of gintonin.³¹

Regarding HD, elevated levels of LPA species were observed in the striatum of the Q140/Q140 HD mice model.⁷² A treatment with gintonin exerted substantial protective effects in a 3-NPA model through the activation of LPA receptors and the nuclear factor erythroid 2-related factor 2 (Nrf-2) signalling pathway, along with the inhibition of the mitogen-activated protein kinases (MAPKs) and the nuclear factor- κ B (NF- κ B) signalling pathways. Moreover, gintonin demonstrates protective effects in STHdh cells and in an adeno-associated viral (AAV) vector-infected model of HD.³⁸

2.3 | Sphingosine 1-phosphate system

The S1P system is a lipid-based neurotransmitter system that is located all along the organism. Five different transmembrane receptors, S1P₁₋₅, which are coupled to different G proteins (G $\alpha_{i/o}$, G $\alpha_{12/13}$, G α_s and/or G α_q), have been described.⁷³ The endogenous ligand is S1P (Figure 1), a lipid signalling molecule that belongs to the family of sphingolipids. S1P is in constant balance with its precursor, ceramide, regulated by enzymatic processes. Ceramide is catabolized by ceramidases into sphingosine, which in turn is phosphorylated by sphingosine kinase enzymes (SPHK1/SPHK2) into S1P, and it can be catabolized by S1P lyase (S1PL). On the contrary, S1P can be dephosphorylated by sphingosine

phosphatases into sphingosine, to be later anabolized by ceramide synthases into ceramide.³⁶ Both neurolipids, S1P and ceramide, play an important role in cellular processes involved with survival, proliferation and differentiation; nevertheless, their effects are opposite. That is, while S1P acts as a positive modulator of cellular survival, displaying anti-apoptotic, proliferative and cell-trafficking effects, ceramide acts as a negative modulator, inducing apoptosis and cellular growth arrest.³⁷

Aging, the main risk factor for AD development, has been linked with a disbalance in several sphingolipid molecules (Figure 2), highlighting S1P/sphingosine balance disruption in elderly people.⁷⁴ In a plasma samples study conducted within a longitudinal cohort involving healthy individuals, cognitively impaired non-demented individuals, AD patients and vascular dementia (VD) patients, it was observed that an increase in the ratio of d18:1 to d16:1 S1P correlated with cognitive deficits. Interestingly, despite these observed changes in the S1P ratios, the overall levels of S1P in plasma did not exhibit significant reductions.⁷⁵ In a subsequent study, a lipidomic analysis of plasma samples from control and both PD and AD patients showed a decrease in S1P levels and an increase in monoheylceramide and lactosylceramide levels in AD and PD groups compared with control groups.⁷⁶ Cerebrospinal fluid (CSF) lipid alterations have also been reported in PD and AD; sphingomyelins d18:1/18:0, which are precursors of ceramides and S1P, are increased in the CSF of patients that display pathological levels of A β ₄₂, Tau and P-tau-181.⁷⁷

A recent study indicates S1P metabolism dysregulation in the 5xFAD mouse model of AD, revealing a decrease in SPHK1/SPHK2 levels and an increase of S1PL levels, together with the increase of the S1P₁ receptor expression.⁷⁸ Strikingly, using the same model, a treatment of ponesimod, an agonist of S1P receptors, was able to offer neuroprotection by reducing tumor necrosis factor-alpha (TNF- α) and C-X-C motif chemokine ligand 10 (CXCL10) levels while increasing anti-inflammatory interleukin 33 (IL-33) levels and improving spatial memory.²² Previous studies suggested that S1P₁ receptor activity is downregulated in several brain areas, including the hippocampus, in the 3xTg-AD mouse model.⁷ Several studies also suggest that FTY720, commonly known as fingolimod, a functional antagonist of S1P₁ receptors, can provide neuroprotection against learning and memory deterioration in mouse and rat AD models. As an example, a subchronic treatment with fingolimod restored cognitive impairment as measured in the Morris water maze (MWM), using an AD-like rat model.¹⁴ Moreover, a more recent study demonstrated that fingolimod exerted neuroprotection against cognitive impairment in 3xTg-AD mice in both novel object localization test and MWM.¹⁵ Synthesis and

degradation enzymes of the S1P system have also been reported to display an important role in neuroinflammatory processes; indeed, an elevation of SPHK1 improved AD-like pathology in the APP/PS1 model.¹⁶

Regarding PD, a clinical study showed that serum S1P levels were significantly depleted in PD patients compared to the control group. A sub-cohort of the same study demonstrated an association between decreased motor functions and decreased S1P serum levels.⁷⁹ In a 1-methyl-4-phenylpyridinium (MPP+) in vitro model, SPHK1 depletion diminished cell viability and could be restored by addition of S1P to the medium.²³ In the same way, using the aforementioned model, SPHK1 inhibition via targeting microRNA-6862 increases SPHK1 expression and protects neurons.⁸⁰ In various different in vivo models of PD, including models by intraparenchymal injection of rotenone, 6-OHDA or MPTP, as well as in GM2+/- transgenic mice, fingolimod or SEW-2871, a selective agonist of the S1P₁ receptor, improved motor function and provided neuroprotection through augmented brain-derived neurotrophic factor (BDNF)^{24,25} and through the S1PR1/AKT/BNIP3/PINK1/Parkin axis.²⁶

Concerning HD, *postmortem* HD patients show an aberrant metabolism of S1P where there is an increase in the expression and activity of S1PL and reduction of SPHK1.⁸¹ In vivo experiments have described that the augmentation of SPHK1 or the inhibition of SPHK2 as well as S1PL has neuroprotective effects against huntingtin-evoked neurotoxicity.⁸² A treatment with a specific agonist of S1P₅ receptors, A-971432, not only ameliorated motor dysfunction in the R6/2 mice model but also increased their lifespan.³³ In a further experiment, a treatment with an enhancer of SPHK1, K6PC-5, was able to increase S1P, BDNF, S1P₁ and S1P₅ expression while preventing motor dysfunction.³⁴ Interestingly, a study using both R6/2 mice model and fly models of HD treated with 2-acetyl-5-tetrahydroxybutyl imidazole, an SGLP1 inhibitor, recovered motor functions, BDNF levels and white matter integrity.³⁵ Nevertheless, no clinical trials have yet been made involving S1P drugs for the treatment of HD.

3 | CONCLUSIONS AND FUTURE PERSPECTIVES

Neurolipids are important signalling molecules with potential therapeutic effects against the most common dementias associated with the main neurodegenerative diseases, as has been demonstrated not only with preclinical in vitro and in vivo experiments but also with data from patients (Table 1). eCB and S1P drugs have shown relevant and interesting results in several animal models

of dementias from different origins, paving the way for clinical trials, although further data acquisition is needed in this regard. Despite the limited evidence regarding the involvement of LPA, it also plays an important role in the development of dementia and could be a potential coadjutant for future therapeutic approaches.

In summary, neurolipids represent a novel approach for the advancement of innovative therapies targeting the most prevalent forms of dementia, including AD, PD and HD. A deeper comprehension of these lipid systems holds the promise of enhancing our understanding of the aetiology, progression and potential treatments for these neurological conditions.

4 | LIMITATIONS OF THE REVIEW

The present review focuses on the description of the relationship of neurolipid systems in three specific diseases with dementia, such as AD, PD and HD. Among all the diseases that lead to dementia, these three are those that have attracted the most interest in the field of neurolipids. Although research on lipids that have neurotransmitter activity is a field that is expanding thanks to new techniques in the field of lipids that have been developed in recent years, there are currently not a large number of specific studies in the context of dementias, which may hinder a complete and thorough view of the subject. Much of the research in this field is conducted in animal models, limiting the direct extrapolation to human pathophysiology and its clinical relevance in dementias. A high heterogeneity has been observed in the design of the studies used for this review, and this may complicate the comparison and summarizing of the results, preventing a consensus on the usefulness of neurolipid treatments in these diseases. Due to the exponential growth of knowledge in this field, it is challenging to include every new publication within this article. Consequently, we have selectively included a limited number of the most pertinent, recent and innovative studies.

ACKNOWLEDGEMENTS

Technical and human support provided by University of the Basque Country (UPV/EHU), Ministry of Economy and Competitiveness (MINECO), Basque Government (GV/EJ), European Regional Development Fund (ERDF), and European Social Fund (ESF) is gratefully acknowledged. G.P.-C. is the recipient of a University of the Basque Country predoctoral fellowship. I.B.d.T. is the recipient of an Investigo fellowship funded by the European Union-Next Generation EU. J.M.-G. is the recipient of Margarita Salas fellowship funded by the European Union-Next Generation EU.

This work was supported by grants from Basque Government IT1454-22 to the ‘Neurochemistry and Neurodegeneration’ consolidated research group and by Instituto de Salud Carlos III through the project ‘PI20/00153’ (co-funded by European Regional Development Fund ‘A way to make Europe’) and by BIOEF project BIO22/ALZ/010 funded by Eitb Maratoia.

Both figures of this work were created with [BioRender.com](https://www.biorender.com).

CONFLICT OF INTEREST STATEMENT

The authors have no conflicts of interest to declare.

DATA AVAILABILITY STATEMENT

Data sharing not applicable to this article as no datasets were generated or analysed during the current study.

ORCID

Iker Bengoetxea de Tena  <https://orcid.org/0000-0003-0586-1857>

Iván Manuel  <https://orcid.org/0000-0003-0958-4738>

REFERENCES

- Mehta RI, Schneider JA. Neuropathology of the common forms of dementia. *Clin Geriatr Med*. 2023;39(1):91-107. doi:10.1016/j.cger.2022.07.005
- Mufson EJ, Ikonovic MD, Counts SE, et al. Molecular and cellular pathophysiology of preclinical Alzheimer's disease. *Behav Brain Res*. 2016;311:54-69. doi:10.1016/j.bbr.2016.05.030
- Mufson EJ, Counts SE, Perez SE, Ginsberg SD. Cholinergic system during the progression of Alzheimer's disease: therapeutic implications. *Expert Rev Neurother*. 2008;8(11):1703-1718. doi:10.1586/14737175.8.11.1703
- Walker L, Stefanis L, Attems J. Clinical and neuropathological differences between Parkinson's disease, Parkinson's disease dementia and dementia with Lewy bodies—current issues and future directions. *J Neurochem*. 2019;150(5):467-474. doi:10.1111/jnc.14698
- Palaiogeorgou A, Papakonstantinou E, Goufinaidou R, et al. Recent approaches on Huntington's disease (Review). *Biomedical Reports*. 2022;18(1):5. doi:10.3892/br.2022.1587
- Coppen EM, Roos RAC. Current pharmacological approaches to reduce chorea in Huntington's disease. *Drugs*. 2017;77(1):29-46. doi:10.1007/s40265-016-0670-4
- González de San Román E, Llorente-Ovejero A, Martínez-Gardeazabal J, et al. Modulation of neurolipid signaling and specific lipid species in the triple transgenic mouse model of Alzheimer's disease. *Int J Mol Sci*. 2021;22(22):12256. doi:10.3390/ijms222212256
- Bozek K, Wei Y, Yan Z, et al. Organization and evolution of brain lipidome revealed by large-scale analysis of human, chimpanzee, macaque, and mouse tissues. *Neuron*. 2015;85(4):695-702. doi:10.1016/j.neuron.2015.01.003
- Veloso A, Fernández R, Astigarraga E, et al. Distribution of lipids in human brain. *Anal Bioanal Chem*. 2011;401(1):89-101. doi:10.1007/s00216-011-4882-x

10. Manuel I, Lombardero L, Llorente-Ovejero A, Rodríguez-Puertas R. Neuropeptides and neurolipids. In: *Genetics, Neurology, Behavior, and Diet in Dementia*. Elsevier; 2020:423-439. doi:[10.1016/b978-0-12-815868-5.00027-x](https://doi.org/10.1016/b978-0-12-815868-5.00027-x)
11. Piomelli D, Astarita G, Rapaka R. A neuroscientist's guide to lipidomics. *Nat Rev Neurosci*. 2007;8(10):743-754. doi:[10.1038/nrn2233](https://doi.org/10.1038/nrn2233)
12. Sinclair E, Trivedi DK, Sarkar D, et al. Metabolomics of sebum reveals lipid dysregulation in Parkinson's disease. *Nat Commun*. 2021;12(1):1592. doi:[10.1038/s41467-021-21669-4](https://doi.org/10.1038/s41467-021-21669-4)
13. Ruver-Martins AC, Bicca MA, de Araujo FS, et al. Cannabinoid extract in microdoses ameliorates mnemonic and non-mnemonic Alzheimer's disease symptoms: a case report. *J Med Case Reports*. 2022;16(1):277. doi:[10.1186/s13256-022-03457-w](https://doi.org/10.1186/s13256-022-03457-w)
14. Asle-Rousta M, Kolahdooz Z, Dargahi L, Ahmadiani A, Nasoohi S. Prominence of central sphingosine-1-phosphate receptor-1 in attenuating A β -induced injury by fingolimod. *J Mol Neurosci*. 2014;54(4):698-703. doi:[10.1007/s12031-014-0423-3](https://doi.org/10.1007/s12031-014-0423-3)
15. Fagan SG, Bechet S, Dev KK. Fingolimod rescues memory and improves pathological hallmarks in the 3xTg-AD model of Alzheimer's disease. *Mol Neurobiol*. 2022;59(3):1882-1895. doi:[10.1007/s12035-021-02613-5](https://doi.org/10.1007/s12035-021-02613-5)
16. Lee JY, Han SH, Park MH, et al. Neuronal SphK1 acetylates COX2 and contributes to pathogenesis in a model of Alzheimer's disease. *Nat Commun*. 2018;9(1):1479. doi:[10.1038/s41467-018-03674-2](https://doi.org/10.1038/s41467-018-03674-2)
17. Aso E, Sánchez-Pla A, Vegas-Lozano E, Maldonado R, Ferrer I. Cannabis-based medicine reduces multiple pathological processes in A β PP/PS1 mice. *J Alzheimers Dis*. 2015;43(3):977-991. doi:[10.3233/JAD-141014](https://doi.org/10.3233/JAD-141014)
18. Llorente-Ovejero A, Manuel I, Lombardero L, et al. Endocannabinoid and muscarinic signaling crosstalk in the 3xTg-AD mouse model of Alzheimer's disease. *J Alzheimers Dis*. 2018;64(1):117-136. doi:[10.3233/JAD-180137](https://doi.org/10.3233/JAD-180137)
19. Hwang SH, Shin EJ, Shin TJ, et al. Gintonin, a ginseng-derived lysophosphatidic acid receptor ligand, attenuates Alzheimer's disease-related neuropathies: involvement of non-amyloidogenic processing. *J Alzheimers Dis*. 2012;31(1):207-223. doi:[10.3233/JAD-2012-120439](https://doi.org/10.3233/JAD-2012-120439)
20. Sun Y, Kim NH, Yang H, Kim SH, Huh SO. Lysophosphatidic acid induces neurite retraction in differentiated neuroblastoma cells via GSK-3 β activation. *Mol Cells*. 2011;31(5):483-490. doi:[10.1007/s10059-011-1036-0](https://doi.org/10.1007/s10059-011-1036-0)
21. Thomas A, Ramananda Y, Mun KS, Naren AP, Arora K. AC6 is the major adenylate cyclase forming a diarrheagenic protein complex with cystic fibrosis transmembrane conductance regulator in cholera. *J Biol Chem*. 2018;293(33):12949-12959. doi:[10.1074/jbc.RA118.003378](https://doi.org/10.1074/jbc.RA118.003378)
22. Zhu Z, Zhang L, Elsherbini A, et al. The S1P receptor 1 antagonist Ponesimod reduces TLR4-induced neuroinflammation and increases A β clearance in 5XFAD mice. *EBioMedicine*. 2023;94:104713. doi:[10.1016/j.ebiom.2023.104713](https://doi.org/10.1016/j.ebiom.2023.104713)
23. Pyszko J, Strosznajder JB. Sphingosine kinase 1 and sphingosine-1-phosphate in oxidative stress evoked by 1-methyl-4-phenylpyridinium (MPP⁺) in human dopaminergic neuronal cells. *Mol Neurobiol*. 2014;50(1):38-48. doi:[10.1007/s12035-013-8622-4](https://doi.org/10.1007/s12035-013-8622-4)
24. Vidal-Martinez G, Najera K, Miranda JD, et al. FTY720 improves behavior, increases brain derived neurotrophic factor levels and reduces α -synuclein pathology in parkinsonian GM2 +/- mice. *Neuroscience*. 2019;411:1-10. doi:[10.1016/j.neuroscience.2019.05.029](https://doi.org/10.1016/j.neuroscience.2019.05.029)
25. Pépin É, Jalinier T, Lemieux GL, Massicotte G, Cyr M. Sphingosine-1-phosphate receptors modulators decrease signs of neuroinflammation and prevent Parkinson's disease symptoms in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model. *Front Pharmacol*. 2020;11. doi:[10.3389/fphar.2020.00077](https://doi.org/10.3389/fphar.2020.00077)
26. Rajan S, Sood A, Jain R, Kamatham PT, Khatri DK. Fingolimod exerts neuroprotection by regulating S1PR1 mediated BNIP3-PINK1-Parkin dependent mitophagy in rotenone induced mouse model of Parkinson's disease. *Neurosci Lett*. 2024;820:137596. doi:[10.1016/j.neulet.2023.137596](https://doi.org/10.1016/j.neulet.2023.137596)
27. Mounsey RB, Mustafa S, Robinson L, et al. Increasing levels of the endocannabinoid 2-AG is neuroprotective in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease. *Exp Neurol*. 2015;273:36-44. doi:[10.1016/j.expneurol.2015.07.024](https://doi.org/10.1016/j.expneurol.2015.07.024)
28. Celorrio M, Fernández-Suárez D, Rojo-Bustamante E, et al. Fatty acid amide hydrolase inhibition for the symptomatic relief of Parkinson's disease. *Brain Behav Immun*. 2016;57:94-105. doi:[10.1016/j.bbi.2016.06.010](https://doi.org/10.1016/j.bbi.2016.06.010)
29. García C, Palomo-Garo C, García-Arencibia M, Ramos J, Pertwee R, Fernández-Ruiz J. Symptom-relieving and neuroprotective effects of the phytocannabinoid Δ^9 -THCV in animal models of Parkinson's disease. *Br J Pharmacol*. 2011;163(7):1495-1506. doi:[10.1111/j.1476-5381.2011.01278.x](https://doi.org/10.1111/j.1476-5381.2011.01278.x)
30. Burgaz S, García C, Gómez-Cañas M, Rolland A, Muñoz E, Fernández-Ruiz J. Neuroprotection with the cannabidiol quinone derivative VCE-004.8 (EHP-101) against 6-hydroxydopamine in cell and murine models of Parkinson's disease. *Molecules*. 2021;26(11):3245. doi:[10.3390/molecules26113245](https://doi.org/10.3390/molecules26113245)
31. Choi JH, Jang M, Oh S, Nah SY, Cho IH. Multi-target protective effects of gintonin in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-mediated model of Parkinson's disease via lysophosphatidic acid receptors. *Front Pharmacol*. 2018;9(MAY):515. doi:[10.3389/fphar.2018.00515](https://doi.org/10.3389/fphar.2018.00515)
32. López-Sendón Moreno JL, García Caldentey J, Trigo Cubillo P, et al. A double-blind, randomized, cross-over, placebo-controlled, pilot trial with Sativex in Huntington's disease. *J Neurol*. 2016;263(7):1390-1400. doi:[10.1007/s00415-016-8145-9](https://doi.org/10.1007/s00415-016-8145-9)
33. di Pardo A, Castaldo S, Amico E, et al. Stimulation of S1PR5 with A-971432, a selective agonist, preserves blood-brain barrier integrity and exerts therapeutic effect in an animal model of Huntington's disease. *Hum Mol Genet*. 2018;27(14):2490-2501. doi:[10.1093/hmg/ddy153](https://doi.org/10.1093/hmg/ddy153)
34. di Pardo A, Pepe G, Castaldo S, et al. Stimulation of sphingosine kinase 1 (SPHK1) is beneficial in a Huntington's disease pre-clinical model. *Front Mol Neurosci*. 2019;12. doi:[10.3389/fnmol.2019.00100](https://doi.org/10.3389/fnmol.2019.00100)
35. Pepe G, Capocci L, Marracino F, et al. Treatment with THI, an inhibitor of sphingosine-1-phosphate lyase, modulates glycosphingolipid metabolism and results therapeutically effective

- in experimental models of Huntington's disease. *Mol Ther*. 2023;31(1):282-299. doi:10.1016/j.yymthe.2022.09.004
36. Hannun YA, Obeid LM. Sphingolipids and their metabolism in physiology and disease. *Nat Rev Mol Cell Biol*. 2018;19(3):175-191. doi:10.1038/nrm.2017.107
 37. Tringali C, Giussani P. Ceramide and sphingosine-1-phosphate in neurodegenerative disorders and their potential involvement in therapy. *Int J Mol Sci*. 2022;23(14):7806. doi:10.3390/ijms23147806
 38. Jang M, Choi JH, Chang Y, Lee SJ, Nah SY, Cho IH. Gintonin, a ginseng-derived ingredient, as a novel therapeutic strategy for Huntington's disease: activation of the Nrf2 pathway through lysophosphatidic acid receptors. *Brain Behav Immun*. 2019;80:146-162. doi:10.1016/j.bbi.2019.03.001
 39. Lu HC, Mackie K. Review of the endocannabinoid system. *Biological Psychiatry Cognitive Neuroscience and Neuroimaging*. 2021;6(6):607-615. doi:10.1016/j.bpsc.2020.07.016
 40. Rezende B, Alencar AKN, de Bem GF, Fontes-Dantas FL, Montes GC. Endocannabinoid system: chemical characteristics and biological activity. *Pharmaceuticals*. 2023;16(2):148. doi:10.3390/ph16020148
 41. Manuel I, de San Román EG, Giralt MT, Ferrer I, Rodríguez-Puertas R. Type-1 cannabinoid receptor activity during Alzheimer's disease progression. *J Alzheimers Dis*. 2014;42(3):761-766. doi:10.3233/JAD-140492
 42. Farkas S, Nagy K, Palkovits M, et al. [¹²⁵I]SD-7015 reveals fine modalities of CB₁ cannabinoid receptor density in the prefrontal cortex during progression of Alzheimer's disease. *Neurochem Int*. 2012;60(3):286-291. doi:10.1016/j.neuint.2011.11.004
 43. Solas M, Francis PT, Franco R, Ramirez MJ. CB2 receptor and amyloid pathology in frontal cortex of Alzheimer's disease patients. *Neurobiol Aging*. 2013;34(3):805-808. doi:10.1016/j.neurobiolaging.2012.06.005
 44. Coyle JT, Price DL, DeLong MR. Alzheimer's disease: a disorder of cortical cholinergic innervation. *Science*. 1983;219(4589):1184-1190. doi:10.1126/science.6338589
 45. Llorente-Ovejero A, Manuel I, Giralt MT, Rodríguez-Puertas R. Increase in cortical endocannabinoid signaling in a rat model of basal forebrain cholinergic dysfunction. *Neuroscience*. 2017;362:206-218. doi:10.1016/j.neuroscience.2017.08.008
 46. Llorente-Ovejero A, Bengoetxea de Tena I, Martínez-Gardeazabal J, et al. Cannabinoid receptors and glial response following a basal forebrain cholinergic lesion. *ACS Pharmacol Transl Sci*. 2022;5(9):791-802. doi:10.1021/acspsci.2c00069
 47. Vidal-Palencia L, Ramon-Duaso C, González-Parra JA, Busquets-García A. Gene expression analysis of the endocannabinoid system in presymptomatic APP/PS1 mice. *Front Pharmacol*. 2022;13. doi:10.3389/fphar.2022.864591
 48. Aso E, Andrés-Benito P, Ferrer I. Delineating the efficacy of a cannabis-based medicine at advanced stages of dementia in a murine model. *J Alzheimers Dis*. 2016;54(3):903-912. doi:10.3233/JAD-160533
 49. Medina-Vera D, Rosell-Valle C, López-Gamero AJ, et al. Imbalance of endocannabinoid/lysophosphatidylinositol receptors marks the severity of Alzheimer's disease in a pre-clinical model: a therapeutic opportunity. *Biology (Basel)*. 2020;9(11):377. doi:10.3390/biology9110377
 50. Fernández-Moncada I, Eraso-Pichot A, Dalla Tor T, Fortunato-Marsol B, Marsicano G. An enquiry to the role of CB1 receptors in neurodegeneration. *Neurobiol Dis*. 2023;184:106235. doi:10.1016/j.nbd.2023.106235
 51. Manuel I, Lombardero L, LaFerla FM, Giménez-Llort L, Rodríguez-Puertas R. Activity of muscarinic, galanin and cannabinoid receptors in the prodromal and advanced stages in the triple transgenic mice model of Alzheimer's disease. *Neuroscience*. 2016;329:284-293. doi:10.1016/j.neuroscience.2016.05.012
 52. Rojo-Bustamante E, Abellanas MA, Clavero P, et al. The expression of cannabinoid type 1 receptor and 2-arachidonoyl glycerol synthesizing/degrading enzymes is altered in basal ganglia during the active phase of levodopa-induced dyskinesia. *Neurobiol Dis*. 2018;118:64-75. doi:10.1016/j.nbd.2018.06.019
 53. Kluger B, Triolo P, Jones W, Jankovic J. The therapeutic potential of cannabinoids for movement disorders. *Mov Disord*. 2015;30(3):313-327. doi:10.1002/mds.26142
 54. de Faria SM, de Moraes Fabrício D, Tumas V, et al. Effects of acute cannabidiol administration on anxiety and tremors induced by a simulated public speaking test in patients with Parkinson's disease. *J Psychopharmacol*. 2020;34(2):189-196. doi:10.1177/0269881119895536
 55. Peball M, Krismer F, Knaus HG, et al. Non-motor symptoms in Parkinson's disease are reduced by nabilone. *Ann Neurol*. 2020;88(4):712-722. doi:10.1002/ana.25864
 56. Sagredo O, Ramos JA, Decio A, Mechoulam R, Fernández-Ruiz J. Cannabidiol reduced the striatal atrophy caused 3-nitropropionic acid in vivo by mechanisms independent of the activation of cannabinoid, vanilloid TRPV1 and adenosine A2A receptors. *Eur J Neurosci*. 2007;26(4):843-851. doi:10.1111/j.1460-9568.2007.05717.x
 57. Sagredo O, González S, Aroyo I, et al. Cannabinoid CB2 receptor agonists protect the striatum against malonate toxicity: relevance for Huntington's disease. *Glia*. 2009;57(11):1154-1167. doi:10.1002/glia.20838
 58. Geraldo LHM, Spohr TCLS, Amaral RFD, et al. Role of lysophosphatidic acid and its receptors in health and disease: novel therapeutic strategies. *Signal Transduct Target Ther*. 2021;6(1):45. doi:10.1038/s41392-020-00367-5
 59. Majer A, Pesthy J, Besztercei B, et al. Characterization of native and human serum albumin-bound lysophosphatidic acid species and their effect on the viability of mesenchymal stem cells in vitro. *Applied Sciences*. 2022;12(16):8183. doi:10.3390/app12168183
 60. Erickson KI, Voss MW, Prakash RS, et al. Exercise training increases size of hippocampus and improves memory. *Proc Natl Acad Sci U S A*. 2011;108(7):3017-3022. doi:10.1073/pnas.1015950108
 61. Hu X, Haney N, Kropp D, Kabore AF, Johnston JB, Gibson SB. Lysophosphatidic acid (LPA) protects primary chronic lymphocytic leukemia cells from apoptosis through LPA receptor activation of the anti-apoptotic protein AKT/PKB. *J Biol Chem*. 2005;280(10):9498-9508. doi:10.1074/jbc.M410455200
 62. Zhang X, Li M, Yin N, Zhang J. The expression regulation and biological function of autotaxin. *Cells*. 2021;10(4):939. doi:10.3390/cells10040939
 63. Meduri B, Pujar GV, Durai Ananda Kumar T, et al. Lysophosphatidic acid (LPA) receptor modulators: structural features

- and recent development. *Eur J Med Chem.* 2021;222:113574. doi:10.1016/j.ejmech.2021.113574
64. Umemura K, Yamashita N, Yu X, et al. Autotaxin expression is enhanced in frontal cortex of Alzheimer-type dementia patients. *Neurosci Lett.* 2006;400(1–2):97–100. doi:10.1016/j.neulet.2006.02.008
 65. Zhang J, Cong YN, Li ZG, et al. Plasma phospholipids are associated with mild cognitive impairment in type 2 diabetic patients. *Curr Alzheimer Res.* 2017;14(6):592–597. doi:10.2174/1567205013666161201200722
 66. Shi J, Dong Y, Cui MZ, Xu X. Lysophosphatidic acid induces increased BACE1 expression and A β formation. *Biochim Biophys Acta Mol Basis Dis.* 2013;1832(1):29–38. doi:10.1016/j.bbadis.2012.09.010
 67. Sayas CL, Avila J, Wandosell F. Glycogen synthase kinase-3 is activated in neuronal cells by G α_{12} and G α_{13} by rho-independent and rho-dependent mechanisms. *J Neurosci.* 2002; 22(16):6863–6875. doi:10.1523/JNEUROSCI.22-16-06863.2002
 68. Matas-Rico E, García-Díaz B, Llebreg-Zayas P, et al. Deletion of lysophosphatidic acid receptor LPA1 reduces neurogenesis in the mouse dentate gyrus. *Molecular and Cellular Neuroscience.* 2008;39(3):342–355. doi:10.1016/j.mcn.2008.07.014
 69. Dash PK, Orsi SA, Moody M, Moore AN. A role for hippocampal Rho-ROCK pathway in long-term spatial memory. *Biochem Biophys Res Commun.* 2004;322(3):893–898. doi:10.1016/j.bbrc.2004.08.004
 70. Castilla-Ortega E, Sánchez-López J, Hoyo-Becerra C, et al. Exploratory, anxiety and spatial memory impairments are dissociated in mice lacking the LPA1 receptor. *Neurobiol Learn Memory.* 2010;94(1):73–82. doi:10.1016/j.nlm.2010.04.003
 71. Yang X, Zhao EY, Zhuang WX, et al. LPA signaling is required for dopaminergic neuron development and is reduced through low expression of the LPA1 receptor in a 6-OHDA lesion model of Parkinson's disease. *Neurol Sci.* 2015;36(11):2027–2033. doi:10.1007/s10072-015-2295-x
 72. Vodicka P, Mo S, Tousley A, et al. Mass spectrometry analysis of wild-type and knock-in Q140/Q140 Huntington's disease mouse brains reveals changes in glycerophospholipids including alterations in phosphatidic acid and lyso-phosphatidic acid. *Journal of Huntington's Disease.* 2015;4(2):187–201. doi:10.3233/JHD-150149
 73. Spiegel S, Toman RE. Lysophospholipid receptors in the nervous system. 2002;27(7/8):619–627. doi:10.1023/A:1020219915922
 74. Couttas TA, Kain N, Tran C, Chatterton Z, Kwok JB, Don AS. Age-dependent changes to sphingolipid balance in the human hippocampus are gender-specific and may sensitize to neurodegeneration. *J Alzheimers Dis.* 2018;63(2):503–514. doi:10.3233/JAD-171054
 75. Chua XY, Chai YL, Chew WS, et al. Immunomodulatory sphingosine-1-phosphates as plasma biomarkers of Alzheimer's disease and vascular cognitive impairment. *Alzheimer's Research & Therapy.* 2020;12(1):122. doi:10.1186/s13195-020-00694-3
 76. Oizumi H, Sugimura Y, Totsune T, et al. Plasma sphingolipid abnormalities in neurodegenerative diseases. *PLoS One.* 2022; 17(12 December). doi:10.1371/journal.pone.0279315
 77. Koal T, Klavins K, Seppi D, Kemmler G, Humpel C. Sphingomyelin SM(d18:1/18:0) is significantly enhanced in cerebrospinal fluid samples dichotomized by pathological amyloid- β 42, tau, and phospho-tau-181 levels. *J Alzheimers Dis.* 2015;44(4):1193–1201. doi:10.3233/JAD-142319
 78. Jung Y, Lopez-Benitez J, Tognoni CM, Carreras I, Dedeoglu A. Dysregulation of sphingosine-1-phosphate (S1P) and S1P receptor 1 signaling in the 5xFAD mouse model of Alzheimer's disease. *Brain Res.* 2023;1799. doi:10.1016/j.brainres.2022.148171
 79. Schwedhelm E, Englisch C, Niemann L, et al. Sphingosine-1-phosphate, motor severity, and progression in Parkinson's disease (MARK-PD). *Mov Disord.* 2021;36(9):2178–2182. doi:10.1002/mds.28652
 80. Xue G, Chen J, Li Y, Zhang ZQ, Zhu JL, Dong W. MicroRNA-6862 inhibition elevates sphingosine kinase 1 and protects neuronal cells from MPP⁺-induced apoptosis. *Aging.* 2021; 13(1):1369–1382. doi:10.18632/aging.202335
 81. di Pardo A, Amico E, Basit A, et al. Defective sphingosine-1-phosphate metabolism is a druggable target in Huntington's disease. *Sci Rep.* 2017;7(1):5280. doi:10.1038/s41598-017-05709-y
 82. Moruno-Manchon JF, Uzor NE, Blasco-Conesa MP, et al. Inhibiting sphingosine kinase 2 mitigates mutant Huntingtin-induced neurodegeneration in neuron models of Huntington disease. *Hum Mol Genet.* 2017;26(7):1305–1317. doi:10.1093/hmg/ddx046

How to cite this article: Pereira-Castelo G, Bengoetxea de Tena I, Martínez-Gardeazabal J, et al. Neurolipid systems: A new target for the treatment of dementia. *Basic Clin Pharmacol Toxicol.* 2024;135(3):225–236. doi:10.1111/bcpt.14059