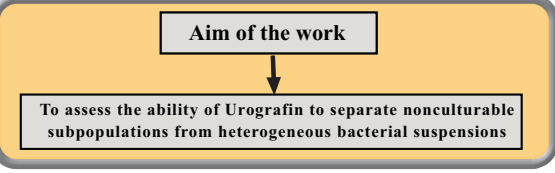
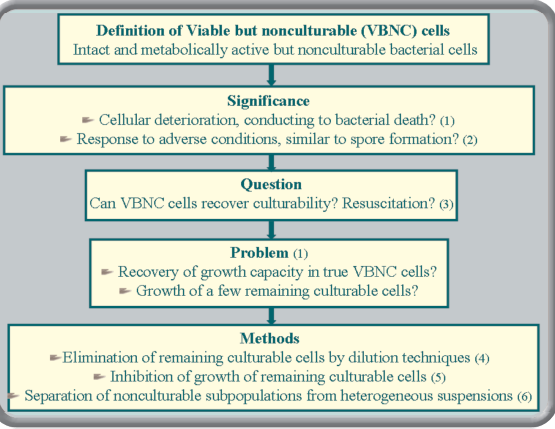


# Ability of Urografin Density Gradients to Separate Nonculturable Subpopulations of *Escherichia coli*

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**METHODOLOGY**

**Strain:** *E. coli* STCC 416  
**Origin of cells:** growth curves (MB, TSB, LB), mixed cultures (stationary phase and aged cultures) or stress exposure (starvation or acid pH)  
**External standard of density:** Density Marker Beads (DMBs, Amersham Biosciences)

**Bacterial determinations**  
Total number of bacteria (TDC) (8), bacteria with intact cytoplasmic membranes (MEMB<sup>+</sup>) (9), colony-forming units (CFU) on Tryptone Soya Agar plates. Flow cytometry measurements with a FACSCalibur flow cytometer (Becton Dickinson, San Jose, CA, USA).

Solution	Urografin (% v/v)	Volume (mL)
A	26.45	2.0
B	30.66	2.0
C	34.87	2.0
D	39.08	2.0
E	43.29	2.0
F	47.50	2.5
G	53.20	2.5

1h, 71,800 g, 20°C

4°C and 20°C 14 h

• Urografin (solution A or F)  
• Percoll (not diluted or IPS)  
• Saline solution

Inoculation 10<sup>8</sup> cells mL<sup>-1</sup> Stationary phase Bacterial determinations

Fraction	From
Culturable (C)	CFU x 100 TDC
Nonculturable (NC)	(TDC - CFU) x 100 TDC
Viable but nonculturable (VNC)	(MEMB <sup>+</sup> - CFU) x 100 TDC
No viable (NV)	(TDC - MEMB <sup>+</sup> ) x 100 TDC

140,700 g, 4°C

45', 71,800 g, 20°C

Urografin Percoll

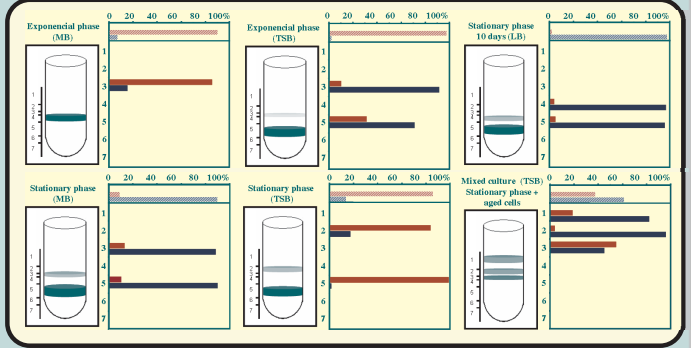
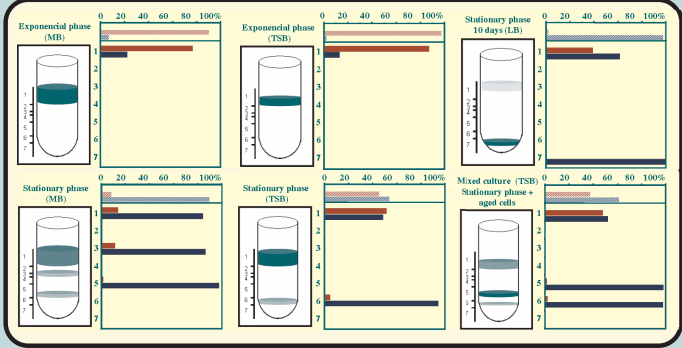
Bands collection for bacterial determinations

**REMARKABLE FACTS**

**Fractionation in Urografin gradients:** very diverse results were obtained when physiologically heterogeneous populations from different culture media, in different phases of growth, and even populations subjected to stress were examined. Formation of low- and high-density bands was observed. The high-density bands (horizon 5 and below) showed a homogeneous composition, mainly of nonculturable cells. In contrast, low-density bands had a heterogeneous composition, being mixtures of culturable and nonculturable cells.

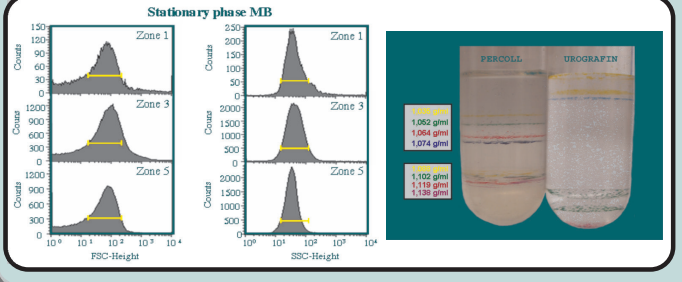
**Fractionation in Percoll gradients:** great differences with respect to the centrifugation in Urografin gradient were detected. A great heterogeneity was observed in number and distribution of bands regardless of the origin of layered cells. Moreover, there was no correlation between band location and the kind of cells

Cellular density	Fraction	C	NC
> 90%	Culture	■	■
50-90%	Band	■	■
10-50%	Band	■	■
1-10%	Band	■	■
< 1%	Band	■	■

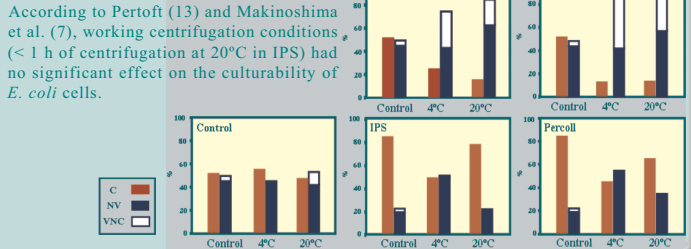


**Bases for separation of VBNC cells:** The separation of culturable and nonculturable cells by means of Urografin gradients has been reported (6, 10) to be based on slight difference in cell densities. A greater density is assigned to nonculturable cells, which are thus understood to concentrate in high-density band. From our results: 1) the increase in cells density would not be to all nonculturable cells (part of these cells banded in the same zone as culturable cells), 2) significant differences in cells size (FSC-Height) or complexity (SSC-Height) were no found between bands, 3) in Urografin gradients, DMBs were distributed in each band according to density, but vertical distribution was not observed.

**Toxicity of fractionation methods based on density gradient:** Urografin has been described as toxic to bacterial cells (11, 12). Centrifugation in density gradients of Urografin may have a harmful effect upon cellular state. This effect depends on time and temperature of centrifugation, as well as on concentration. Our results indicate that Urografin has a toxic effect on cell culturability. The decrease in percentage of culturable cell, and the concomitant increase in the nonculturable fraction, was mainly due to the formation of viable but nonculturable cells rather than the loss of viability.



Percoll is regarded as non-toxic to cells (13). Undiluted Percoll had a more negative effect upon culturability than isotonic Percoll solution (IPS) and the effect of Percoll was temperature dependent (exposure to isotonic solutions at 4°C had a negative effect on culturability).



**CONCLUSION**

We can thus conclude that Urografin density gradients are a good tool to obtain homogeneous nonculturable subpopulations from a heterogeneous *E. coli* populations. However, it is necessary to pay special attention to working conditions to avoid Urografin toxicity.

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